Allen SDK Documentation

Release dev

Allen Institute for Brain Science

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CONTENTS

CHAPTER

ONE

INSTALL GUIDE

This guide is a resource for using the Allen SDK package. It is maintained by the Allen Institute for Brain Science.

Attention: As of October 2019, we have dropped Python 2 support.

1.1 Quick Start

1. Use a virtual environment, e.g Anaconda. After the installation is complete, open up a terminal (in Windows open Anaconda3 Command Prompt).
2. Create a new conda environment and install the AllenSDK using pip

```
conda create -n allensdk
conda activate allensdk
pip install allensdk
```

3. Add conda env to ipykernel so that the notebook can use it

```
pip install ipykernel
python -m ipykernel install --user --name=allensdk
```

4. Explore notebooks.
   - Legacy notebooks
   - visual behavior/visual coding notebooks

   Download one of our many notebooks to a new folder.

   In your terminal, navigate to the directory where you downloaded the Jupyter Notebook example and start jupyer notebook

```
jupyter notebook
```

1.2 Other Distribution Formats

The Allen SDK is also available from the Github source repository.

CHAPTER

TWO

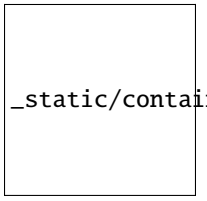
DATA RESOURCES

The Allen SDK features Python code to support data and model access for the Allen Cell Types Database. Resources for other Allen Brain Atlas data resources will come in future updates.

2.1 Brain Observatory

The Allen Brain Observatory is a database of the visually-evoked functional responses of neurons in mouse visual cortex based on 2-photon fluorescence imaging. Characterized responses include orientation tuning, spatial and temporal frequency tuning, temporal dynamics, and spatial receptive field structure.

The data is organized into experiments and experiment containers. An experiment container represents a group of experiments with the same targeted imaging area, imaging depth, and Cre line. The individual experiments within an experiment container have different stimulus protocols, but cover the same imaging field of view.



Note: Version 1.3 of scipy fixed an error in its 2 sample Kolmogorov-Smirnoff test implementation. The new version produces more accurate p values for small and medium-sized samples. This change impacts speed tuning analysis p values (as returned by *StimulusAnalysis.get_speed_tuning*). If you access precalculated analysis results via *BrainObservatoryCache.get_ophys_experiment_analysis*, you will see values calculated using an older version of scipy's *ks_2samp*. To access values calculated from the new version, install scipy>=1.3.0 in your environment and construct a *StimulusAnalysis* object from a *BrainObservatoryNwbDataSet* (as returned by *BrainObservatoryCache.get_ophys_experiment_data*).

Note: Data collected after September 2016 uses a new session C stimulus designed to better-characterize spatial receptive fields in higher visual areas. The original locally sparse noise stimulus used 4.65 visual degree pixels. Session C2 broke that stimulus into two separate stimulus blocks: one with 4.65 degree pixels and one with 9.3 degree pixels. Note that the `stimulus_info` module refers to these as *locally_sparse_noise_4deg* and *locally_sparse_noise_8deg*, respectively.

For more information on experimental design and a data overview, please visit the Allen Brain Observatory data portal.

2.1.1 Data Processing

For all data in Allen Brain Observatory, we perform the following processing:

1. Segment cell masks from each experiment's 2-photon fluorescence video
2. Associate cells from experiments belonging to the same experiment container and assign unique IDs
3. Extract each cell's mean fluorescence trace
4. Extract mean fluorescence traces from each cell's surrounding neuropil
5. Demix traces from overlapping ROIs
6. Estimate neuropil-corrected fluorescence traces
7. Compute dF/F
8. Compute stimulus-specific tuning metrics

All traces and masks for segmented cells in an experiment are stored in a Neurodata Without Borders (NWB) file. Stored traces include the raw fluoresence trace, neuropil trace, demixed trace, and dF/F trace. Code for extracting neuropil-corrected fluorescence traces, computing dF/F, and computing tuning metrics is available in the SDK.

New in June 2017: Trace demixing is a new addition as of June 2017. All past data was reprocessed using the new demixing algorithm. We have also developed a new module to better characterize a cell's receptive field. Take a look at the receptive field analysis example notebook

For more information about data processing, please read the technical whitepapers.

2.1.2 Getting Started

The Brain Observatory Jupyter notebook has many code samples to help get started with the available data:

- Download experimental metadata by visual area, imaging depth, and Cre line
- Find cells with specific response properties, like direction tuning
- Download data for an experiment
- Plot raw fluorescences traces, neuropil-corrected traces, and dF/F
- Find the ROI mask for a given cell
- Run neuropil correction
- Get pupil location and size

The code used to analyze and visualize data in the Allen Brain Observatory data portal is available as part of the SDK. Take a look at this Jupyter notebook to find out how to:

- Plot cell's response to its preferred stimulus condition
- Compute a cell's on/off receptive field based on the locally sparse noise stimulus

More detailed documentation is available demonstrating how to:

- Read and visualize the stimulus presentation tables in the NWB files
- Understand the layout of Brain Observatory NWB files
- Map previous cell specimen IDs to current cell specimen IDs

2.1.3 Precomputed Cell Metrics

A large table of precomputed metrics are available for download to support population analysis and filtering. The table below describes all of the metrics in the table. The `get_cell_specimens()` method will download this table as a list of dictionaries which can be converted to a pandas DataFrame as shown in this Jupyter notebook.

Stimulus	Metric	Field Name
drifting gratings	orientation selectivity	osi_dg
	direction selectivity	dsi_dg
	preferred direction	pref_dir_dg
	preferred temporal frequency	pref_tf_dg
	response p value	p_dg
	global ori. selectivity	g_osi_dg
	global dir. selectivity	g_dsi_dg
	response reliability	reliability_dg
	running modulation	run_mod_dg
	running modulation p value	p_run_mod_dg
	pref. condition mean df/f	peak_dff_dg
	TF discrimination index	tfdi_dg
static gratings	orientation selectivity	osi_sg
	preferred orientation	pref_ori_sg
	preferred spatial frequency	pref_sf_sg
	preferred phase	pref_phase_sg
	mean time to peak response	time_to_peak_sg
	response p value	p_sg
	global ori. selectivity	g_osi_sg
	reponse reliability	reliability_sg
	running modulation	run_mod_sg
	running modulation p value	p_run_mod_sg
	pref. condition mean df/f	peak_dff_ns
	SF discrimiation index	sfdi_sg
natural scenes	mean time to peak response	time_to_peak_ns
	preferred scene index	pref_scene_ns
	response p value	p_ns
	image selectivity	image_sel_ns
	running modulation	run_mod_ns
	running modulation p value	p_run_mod_ns
	pref. condition mean df/f	peak_dff_ns
natural movie 1	response reliability (session A)	reliability_nm1_a
	response reliability (session B)	reliability_nm1_b
	response reliability (session C)	reliability_nm1_c
natural movie 2	response reliability	reliability_nm2
natural movie 3	response reliability	reliability_nm3
locally sparse noise	RF area (on subunit)	rf_area_on_lsn
	RF area (off subunit)	rf_area_off_lsn
	RF center (on subunit)	rf_center_on_x, rf_center_on_y
	RF center (off subunit)	rf_center_off_x, rf_center_off_y
	RF chi^2	rf_chi2_lsn
	RF on-off subunit distance	rf_distance_lsn
	RF on-off subunit overlap index	rf_overlap_lsn

2.2 Cell Types

The Allen Cell Types data set is a database of mouse and human neuronal cell types based on multimodal characterization of single cells to enable data-driven approaches to classification and is fully integrated with other Allen Brain Atlas resources. The database currently includes:

- **electrophysiology**: whole cell current clamp recordings made from Cre-positive neurons
- **morphology**: 3D bright-field images of the complete structure of neurons from the visual cortex

This page describes how the SDK can be used to access data in the Cell Types Database. For more information, please visit the Cell Types Database home page and the API documentation.

2.2.1 Examples

The Cell Types Jupyter notebook has many code samples to help get started with analysis:

- Download and plot stimuli and responses from an NWB file for a cell
- Download and plot a cell's morphological reconstruction
- Download and plot precomputed electrophysiology features
- Download precomputed morphology features to a table
- Compute electrophysiology features for a single sweep

2.2.2 Cell Types Cache

The `CellTypesCache` class provides a Python interface for downloading data in the Allen Cell Types Database into well known locations so that you don't have to think about file names and directories. The following example demonstrates how to download meta data for all cells with 3D reconstructions, then download the reconstruction and electrophysiology recordings for one of those cells:

```
from allensdk.core.cell_types_cache import CellTypesCache

ctc = CellTypesCache(manifest_file='cell_types/manifest.json')

# a list of cell metadata for cells with reconstructions, download if necessary
cells = ctc.get_cells(require_reconstruction=True)

# open the electrophysiology data of one cell, download if necessary
data_set = ctc.get_ephys_data(cells[0]['id'])

# read the reconstruction, download if necessary
reconstruction = ctc.get_reconstruction(cells[0]['id'])
```

`CellTypesCache` takes takes care of knowing if you've already downloaded some files and reads them from disk instead of downloading them again. All data is stored in the same directory as the *manifest_file* argument to the constructor.

2.2.3 Feature Extraction

The `EphysFeatureExtractor` class calculates electrophysiology features from cell recordings. *`extract_cell_features()`* can be used to extract the precise feature values available in the Cell Types Database:

```
from allensdk.core.cell_types_cache import CellTypesCache
from allensdk.ephys.extract_cell_features import extract_cell_features
from collections import defaultdict

# initialize the cache
ctc = CellTypesCache(manifest_file='cell_types/manifest.json')

# pick a cell to analyze
specimen_id = 324257146

# download the ephys data and sweep metadata
data_set = ctc.get_ephys_data(specimen_id)
sweeps = ctc.get_ephys_sweeps(specimen_id)

# group the sweeps by stimulus
sweep_numbers = defaultdict(list)
for sweep in sweeps:
    sweep_numbers[sweep['stimulus_name']].append(sweep['sweep_number'])

# calculate features
cell_features = extract_cell_features(data_set,
                                      sweep_numbers['Ramp'],
                                      sweep_numbers['Short Square'],
                                      sweep_numbers['Long Square'])
```

2.2.4 File Formats

This section provides a short description of the file formats used for Allen Cell Types data.

Morphology SWC Files

Morphological neuron reconstructions are available for download as SWC files. The SWC file format is a white-space delimited text file with a standard set of headers. The file lists a set of 3D neuronal compartments, each of which has:

Column	Data Type	Description
id	string	compartment ID
type	integer	compartment type
x	float	3D compartment position (x)
y	float	3D compartment position (y)
z	float	3D compartment position (z)
radius	float	compartment radius
parent	string	parent compartment ID

Comment lines begin with a '#'. Reconstructions in the Allen Cell Types Database can contain the following compartment types:

Type	Description
0	unknown
1	soma
2	axon
3	basal dendrite
4	apical dendrite

The Allen SDK comes with a *swc* Python module that provides helper functions and classes for manipulating SWC files. Consider the following example:

```
import allensdk.core.swc as swc

# if you ran the examples above, you will have a reconstruction here
file_name = 'cell_types/specimen_485909730/reconstruction.swc'
morphology = swc.read_swc(file_name)

# subsample the morphology 3x. root, soma, junctions, and the first child of the root␣
↪are preserved.
sparse_morphology = morphology.sparsify(3)

# compartments in the order that they were specified in the file
compartment_list = sparse_morphology.compartment_list

# a dictionary of compartments indexed by compartment id
compartments_by_id = sparse_morphology.compartment_index

# the root soma compartment
soma = morphology.soma

# all compartments are dictionaries of compartment properties
# compartments also keep track of ids of their children
for child in morphology.children_of(soma):
    print(child['x'], child['y'], child['z'], child['radius'])
```

Neurodata Without Borders

The electrophysiology data collected in the Allen Cell Types Database is stored in the Neurodata Without Borders (NWB) file format. This format, created as part of the NWB initiative, is designed to store a variety of neurophysiology data, including data from intra- and extracellular electrophysiology experiments, optophysiology experiments, as well as tracking and stimulus data. It has a defined schema and metadata labeling system designed so software tools can easily access contained data.

The Allen SDK provides a basic Python class for extracting data from Allen Cell Types Database NWB files. These files store data from intracellular patch-clamp recordings. A stimulus current is presented to the cell and the cell's voltage response is recorded. The file stores both stimulus and response for several experimental trials, here called "sweeps." The following code snippet demonstrates how to extract a sweep's stimulus, response, sampling rate, and estimated spike times:

```
from allensdk.core.nwb_data_set import NwbDataSet

# if you ran the examples above, you will have a NWB file here
```

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```
file_name = 'cell_types/specimen_485909730/ephys.nwb'
data_set = NwbDataSet(file_name)

sweep_numbers = data_set.get_sweep_numbers()
sweep_number = sweep_numbers[0]
sweep_data = data_set.get_sweep(sweep_number)

# spike times are in seconds relative to the start of the sweep
spike_times = data_set.get_spike_times(sweep_number)

# stimulus is a numpy array in amps
stimulus = sweep_data['stimulus']

# response is a numpy array in volts
reponse = sweep_data['response']

# sampling rate is in Hz
sampling_rate = sweep_data['sampling_rate']

# start/stop indices that exclude the experimental test pulse (if applicable)
index_range = sweep_data['index_range']
```

HDF5 Overview

NWB is implemented in HDF5. HDF5 files provide a hierarchical data storage that mirrors the organization of a file system. Just as a file system has directories and files, and HDF5 file has groups and datasets. The best way to understand an HDF5 (and NWB) file is to open a data file in an HDF5 browser. HDFView is the recommended browser from the makers of HDF5.

There are HDF5 manipulation libraries for many languages and platorms. MATLAB and Python in particular have strong HDF5 support.

2.3 Mouse Connectivity

The Allen Mouse Brain Connectivity Atlas consists of high-resolution images of axonal projections targeting different anatomic regions or various cell types using Cre-dependent specimens. Each data set is processed through an informatics data analysis pipeline to obtain spatially mapped quantified projection information.

This page describes how to use the SDK to access experimental projection data and metadata. For more information, please visit the Connectivity Atlas home page and the API documentation

2.3.1 Structure-Level Projection Data

All AAV projection signal in the Allen Mouse Connectivity Atlas has been registered to the expert-annotated Common Coordinate Framework (CCF) and summarized to structures in the adult mouse structure ontology. Most commonly used for analysis are measures of the density of projection signal in all brain areas for every experiment. This data is available for download and is described in more detail on the structure unionizes page.

2.3.2 Voxel-Level Projection Data

The CCF-registered AAV projection signal is also available for download as a set of 3D volumes for each experiment. The following data volumes are available for download:

- **projection density**: sum of detected projection pixels / sum of all pixels in voxel
- **injection_fraction**: fraction of pixels belonging to manually annotated injection site
- **injection_density**: density of detected projection pixels within the manually annotated injection site
- **data_mask**: binary mask indicating if a voxel contains valid data. Only valid voxels should be used for analysis.

2.3.3 Code Examples

The Mouse Connectivity Jupyter notebook has many code samples to help get started with analysis:

- Download experimental metadata by injection structure and transgenic line
- Download projection signal statistics at a structure level
- Build a structure-to-structure matrix of projection signal values
- Download and visualize gridded projection signal volumes

2.3.4 Mouse Connectivity Cache

The *MouseConnectivityCache* class saves all of the data you can download via the MouseConenctivityApi in well known locations so that you don't have to think about file names and directories. It also takes care of knowing if you've already downloaded some files and reads them from disk instead of downloading them again. The following example demonstrates how to download meta data for all experiments with injections in the isocortex and download the projetion density volume for one of them:

```
from allensdk.core.mouse_connectivity_cache import MouseConnectivityCache

# tell the cache class what resolution (in microns) of data you want to download
mcc = MouseConnectivityCache(resolution=25)

# use the structure tree class to get information about the isocortex structure
structure_tree = mcc.get_structure_tree()
isocortex_id = structure_tree.get_structures_by_name(['Isocortex'])[0]['id']

# a list of dictionaries containing metadata for non-Cre experiments
experiments = mcc.get_experiments(file_name='non_cre.json',
                                  injection_structure_ids=[isocortex_id])

# download the projection density volume for one of the experiments
pd = mcc.get_projection_density(experiments[0]['id'])
```

2.3.5 File Formats

This section provides a short description of the file formats used for data in the Allen Mouse Connectivity Atlas.

NRRD Files

All of the volumetric data in the connectivity atlas are stored as NRRD (Nearly Raw Raster Data) files. A NRRD file consists of a short ASCII header followed by a binary array of data values.

To read these in Python, we recommend the pynrrd package. Usage is straightforward:

```
import nrrd

file_name = 'mouse_connectivity/experiment_644250774/projection_density_25.nrrd'
data_array, metadata = nrrd.read(file_name)
```

2.4 Reference Space

Allen Institute atlases and data are registered, when possible, to one of several common reference spaces. Working in such a space allows you to easily compare data across subjects and experimental modalities.

This page documents how to use the Allen SDK to interact with a reference space. For more information and a list of reference spaces, see the atlas drawings and ontologies API documentation and the 3D reference models API documentation. For details about the construction of the Common Coordinate Framework space, see the CCFv3 whitepaper.

2.4.1 Structure Tree

Brain structures in our reference spaces are arranged in trees. The leaf nodes of the tree describe the very fine anatomical divisions of the space, while nodes closer to the root correspond to gross divisions. The *StructureTree* class provides an interface for interacting with a structure tree.

To download a structure tree, use the *allensdk.core.reference_space_cache.ReferenceSpaceCache* class as seen in this example

2.4.2 Annotation Volumes

An annotation volume is a 3d raster image that segments the reference space into structures. Each voxel in the annotation volume is assigned an integer value that describes the finest structure to which that point in space definitely belongs.

To download a nrrd formatted annotation volume at a specified isometric resolution, use the *allensdk.core.reference_space_cache.ReferenceSpaceCache* class. There is an example in the notebook.

2.4.3 ReferenceSpaceCache Class

The *allensdk.core.reference_space_cache.ReferenceSpaceCache* class provides a Python interface for downloading structure trees and annotation volumes. It takes care of knowing if you've already downloaded the files and reads them from disk instead of downloading them again.

The class contains methods for working with our reference spaces. Some use cases might include:

- Building an indicator mask for one or more structures
- Viewing the annotation
- Querying the structure graph

Please see the example notebook for more code samples.

2.5 API Access

The *allensdk.api* package is designed to help retrieve data from the Allen Brain Atlas API. *api* contains methods to help formulate API queries and parse the returned results. There are several pre-made subclasses available that provide pre-made queries specific to certain data sets. Currently there are several subclasses in Allen SDK:

- *CellTypesApi*: data related to the Allen Cell Types Database
- *BiophysicalApi*: data related to biophysical models
- *GlifApi*: data related to GLIF models
- *AnnotatedSectionDataSetsApi*: search for experiments by intensity, density, pattern, and age
- *GridDataApi*: used to download 3-D expression grid data
- *ImageDownloadApi*: download whole or partial two-dimensional images
- *MouseConnectivityApi*: common operations for accessing the Allen Mouse Brain Connectivity Atlas
- *OntologiesApi*: data about neuroanatomical regions of interest
- *ConnectedServices*: schema of Allen Institute Informatics Pipeline services available through the RmaApi
- *RmaApi*: general-purpose HTTP interface to the Allen Institute API data model and services
- *SvgApi*: annotations associated with images as scalable vector graphics (SVG)
- *SynchronizationApi*: data about image alignment
- *TreeSearchApi*: list ancestors or descendents of structure and specimen trees

2.5.1 RMA Database and Service API

One API subclass is the *RmaApi* class. It is intended to simplify constructing an RMA query.

The RmaApi is a base class for much of the allensdk.api.queries package, but it may be used directly to customize queries or to build queries from scratch.

Often a query will simply request a table of data of one type:

```
from allensdk.api.queries.rma_api import RmaApi

rma = RmaApi()

```

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```
data = rma.model_query('Atlas',
                       criteria="[name$il'*Mouse*']")
```

This will construct the RMA query url, make the query and parse the resulting JSON into an array of Python dicts with the names, ids and other information about the atlases that can be accessed via the API.

Using the criteria, include and other parameter, specific data can be requested.

```
associations = ''.join(['[id$eq1]',
                        'structure_graph(ontology),',
                        'graphic_group_labels'])

atlas_data = rma.model_query('Atlas',
                             include=associations,
                             criteria=associations,
                             only=['atlases.id',
                                 'atlases.name',
                                 'atlases.image_type',
                                 'ontologies.id',
                                 'ontologies.name',
                                 'structure_graphs.id',
                                 'structure_graphs.name',
                                 'graphic_group_labels.id',
                                 'graphic_group_labels.name'])
```

Note that a 'class' name is used for the first parameter. 'Association' names are used to construct the include and criteria parameters nested using parentheses and commas. In the only clause, the 'table' form is used, which is generally a plural lower-case version of the class name. The only clause selects specific 'fields' to be returned. The schema that includes the classes, fields, associations and tables can be accessed in JSON form using:

```
# http://api.brain-map.org/api/v2/data.json
schema = rma.get_schema()
for entry in schema:
    data_description = entry['DataDescription']
    clz = list(data_description.keys())[0]
    info = list(data_description.values())[0]
    fields = info['fields']
    associations = info['associations']
    table = info['table']
    print("class: %s" % (clz))
    print("fields: %s" % (','.join(f['name'] for f in fields)))
    print("associations: %s" % (','.join(a['name'] for a in associations)))
    print("table: %s\n" % (table))
```

2.5.2 Using Pandas to Process Query Results

When it is difficult to get data in exactly the required form using only an RMA query, it may be helpful to perform additional operations on the client side. The pandas library can be useful for this.

Data from the API can be read directly into a pandas Dataframe object.

```
import pandas as pd

structures = pd.DataFrame(
    rma.model_query('Structure',
                    criteria='[graph_id$eq1]',
                    num_rows='all'))
```

Indexing subsets of the data (certain columns, certain rows) is one use of pandas: specifically .loc:

```
names_and_acronyms = structures.loc[:,['name', 'acronym']]
```

and Boolean indexing

```
mea = structures[structures.acronym == 'MEA']
mea_id = mea.iloc[0,:].id
mea_children = structures[structures.parent_structure_id == mea_id]
print(mea_children['name'])
```

Concatenate, merge and join are used to add columns or rows:

When an RMA call contains an include clause, the associated data will be represented as a python dict in a single column. The column may be converted to a proper Dataframe and optionally dropped.

```
criteria_string = "structure_sets[name$eq'Mouse Connectivity - Summary']"
include_string = "ontology"
summary_structures = \
    pd.DataFrame(
        rma.model_query('Structure',
                        criteria=criteria_string,
                        include=include_string,
                        num_rows='all'))
ontologies = \
    pd.DataFrame(
        list(summary_structures.ontology)).drop_duplicates()
flat_structures_dataframe = summary_structures.drop(['ontology'], axis=1)
```

Alternatively, it can be accessed using normal python dict and list operations.

```
print(summary_structures.ontology[0]['name'])
```

Pandas Dataframes can be written to a CSV file using to_csv and read using load_csv.

```
summary_structures[['id',
                    'parent_structure_id',
                    'acronym']].to_csv('summary_structures.csv',
                                       index_label='structure_id')
reread = pd.read_csv('summary_structures.csv')
```

Iteration over a Dataframe of API data can be done in several ways. The .itertuples method is one way to do it.

```
for id, name, parent_structure_id in summary_structures[['name',
                                                         'parent_structure_id']].
↪itertuples():
    print("%d %s %d" % (id, name, parent_structure_id))
```

2.5.3 Caching Queries on Disk

wrap() has several parameters for querying the API, saving the results as CSV or JSON and reading the results as a pandas dataframe.

```
from allensdk.api.warehouse_cache.cache import Cache

cache_writer = Cache()
do_cache=True
structures_from_api = \
    cache_writer.wrap(rma.model_query,
                          path='summary.csv',
                          cache=do_cache,
                          model='Structure',
                          criteria='[graph_id$eq1]',
                          num_rows='all')
```

If you change to_cache to False and run the code again it will read the data from disk rather than executing the query.

2.6 Visual Coding – Neuropixels

The Visual Coding – Neuropixels project uses high-density extracellular electrophysiology (**Ecephys**) probes to record spikes from a wide variety of regions in the mouse brain. Our experiments are designed to study the activity of the visual cortex and thalamus in the context of passive visual stimulation, but these data can be used to address a wide variety of topics.

Spike-sorted data and metadata are available via the AllenSDK as Neurodata Without Borders files. However, if you're using the AllenSDK to interact with the data, no knowledge of the NWB data format is required.

2.6.1 Getting Started

To jump right in, check out the quick start guide (download .ipynb), which will show you how to download the data, align spikes to a visual stimulus, and decode natural images from neural activity patterns. For a quick summary of experimental design and data access, see the cheat sheet.

If you would like more example code, the full example notebook (download .ipynb) covers all of the ways to access data for each experiment.

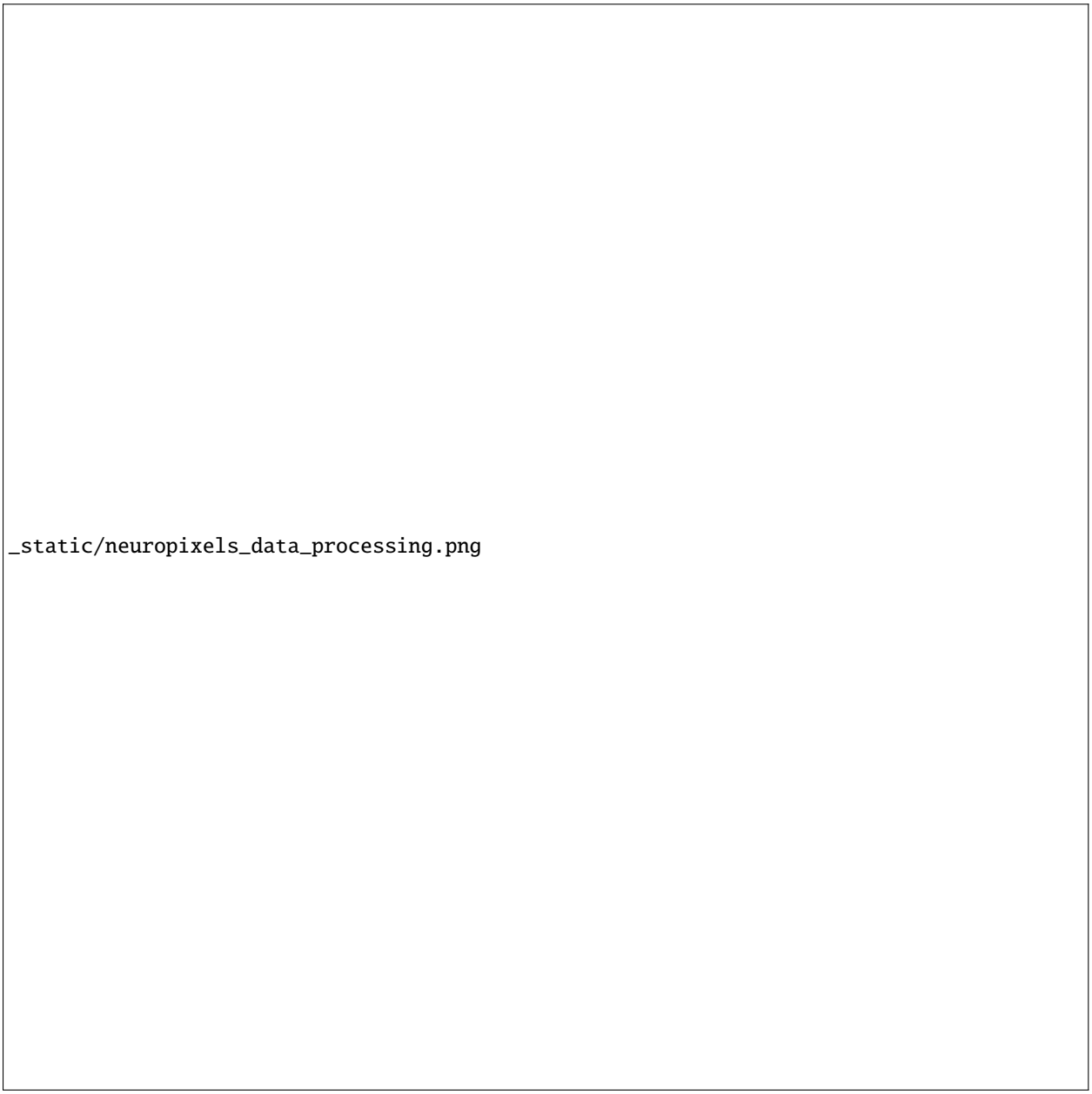
Additional tutorials are available on the following topics:

1. Data access (download .ipynb)
2. Unit quality metrics (download .ipynb)
3. LFP data analysis (download .ipynb)
4. Receptive field mapping (download .ipynb)
5. Optotagging (download .ipynb)

For detailed information about the experimental design, data acquisition, and informatics methods, please refer to our technical whitepaper. AllenSDK API documentation is available here.

A note on terminology: Throughout the SDK, we refer to neurons as "units," because we cannot guarantee that all the spikes assigned to one unit actually originate from a single cell. Unlike in two-photon imaging, where you can visualize each neuron throughout the entire experiment, with electrophysiology we can only "see" a neuron when it fires a spike. If a neuron moves relative to the probe, or if it's far away from the probe, some of its spikes may get mixed together with those from other neurons. Because of this inherent ambiguity, we provide a variety of quality metrics to allow you to find the right units for your analysis. Even highly contaminated units contain potentially valuable information about brain states, so we didn't want to leave them out of the dataset. But certain types of analysis require more stringent quality thresholds, to ensure that all of the included units are well isolated from their neighbors.

2.6.2 Data Processing



Neuropixels probes contain 374 or 383 channels that continuously detect voltage fluctuations in the surrounding neural tissue. Each channel is split into two separate data streams, or "bands," on the probes. The "spike band" is digitized at 30 kHz, and contains information about action potentials fired by neurons directly adjacent to the probe. The "LFP band" is digitized at 2.5 kHz, and records the low-frequency (<1000 Hz) fluctuations that result from synchronized neural activity over a wider area.

To go from the raw spike-band data to NWB files, we perform the following processing steps:

1. Median-subtraction to remove common-mode noise from the continuous traces
2. High-pass filtering (>150 Hz) and whitening across blocks of 32 channels
3. Spike sorting with Kilosort2, to detect spikes and assign them to individual units
4. Computing the mean waveform for each unit
5. Removing units with artifactual waveforms
6. Computing quality metrics for every unit
7. Computing stimulus-specific tuning metrics

For the LFP band, we:

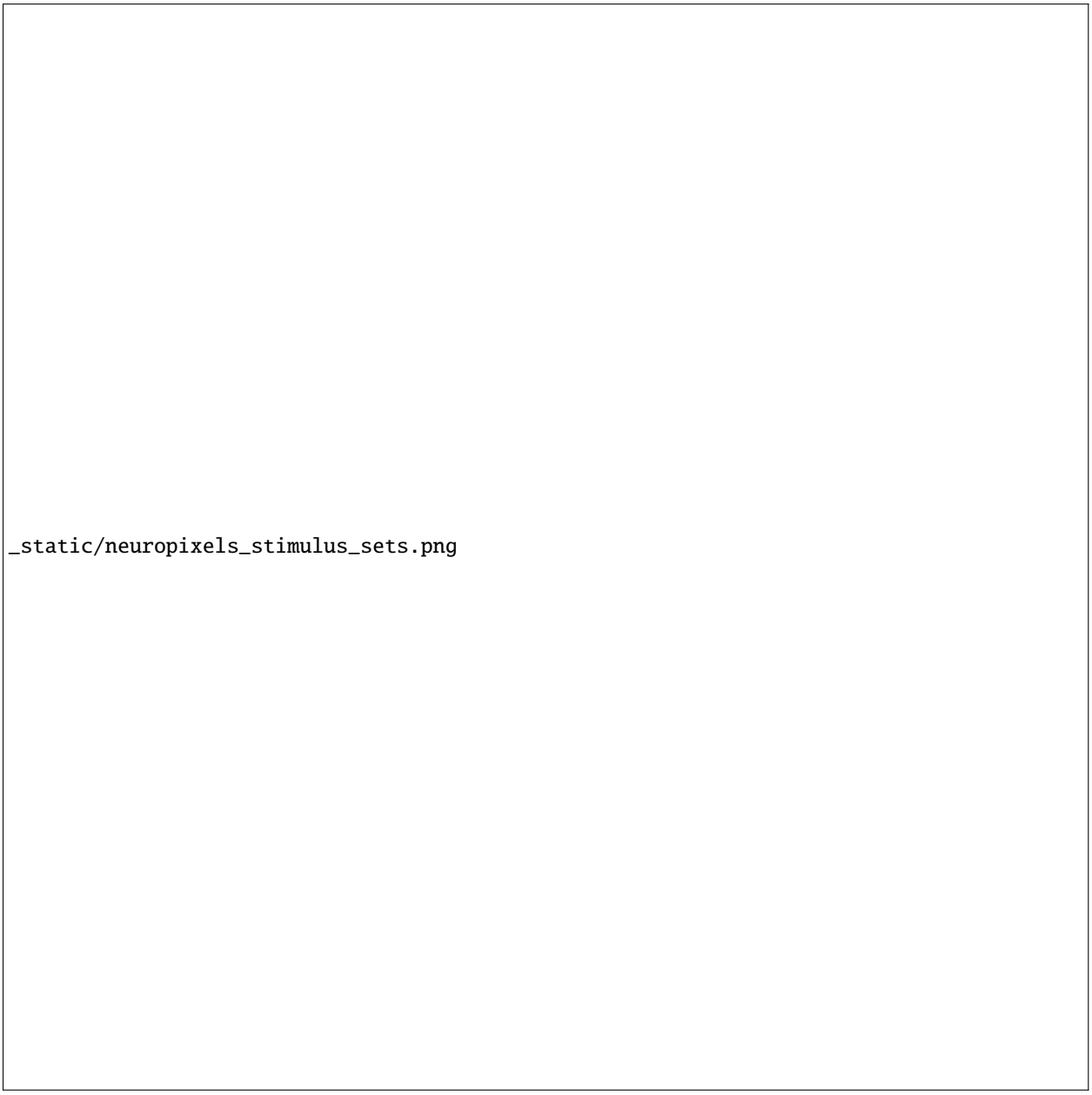
1. Downsample the signals in space and time (every 4th channel and every 2nd sample)
2. High-pass filter at 0.1 Hz to remove the DC offset from each channel
3. Re-reference to channels outside of the brain to remove common-mode noise

The packaged NWB files contain:

1. Spike times, spike amplitudes, mean waveforms, and quality metrics for every unit
2. Information about the visual stimulus
3. Time series of the mouse's running speed, pupil diameter, and pupil position
4. LFP traces for channels in the brain
5. Experiment metadata

All code for data processing and packaging is available in the ecephys_spike_sorting and the ecephys section of the AllenSDK.

2.6.3 Visual Stimulus Sets



A central aim of the Visual Coding – Neuropixels project is to measure the impact of visual stimuli on neurons throughout the mouse visual system. To that end, all mice viewed one of two possible stimulus sets, known as "Brain Observatory 1.1" or "Functional Connectivity". Both stimulus sets began with a Gabor stimulus flashed at 81 different locations on the screen, used to map receptive fields of visually responsive units. Next, the mice were shown brief flashes of light or dark, to measure the temporal dynamics of the visual response.

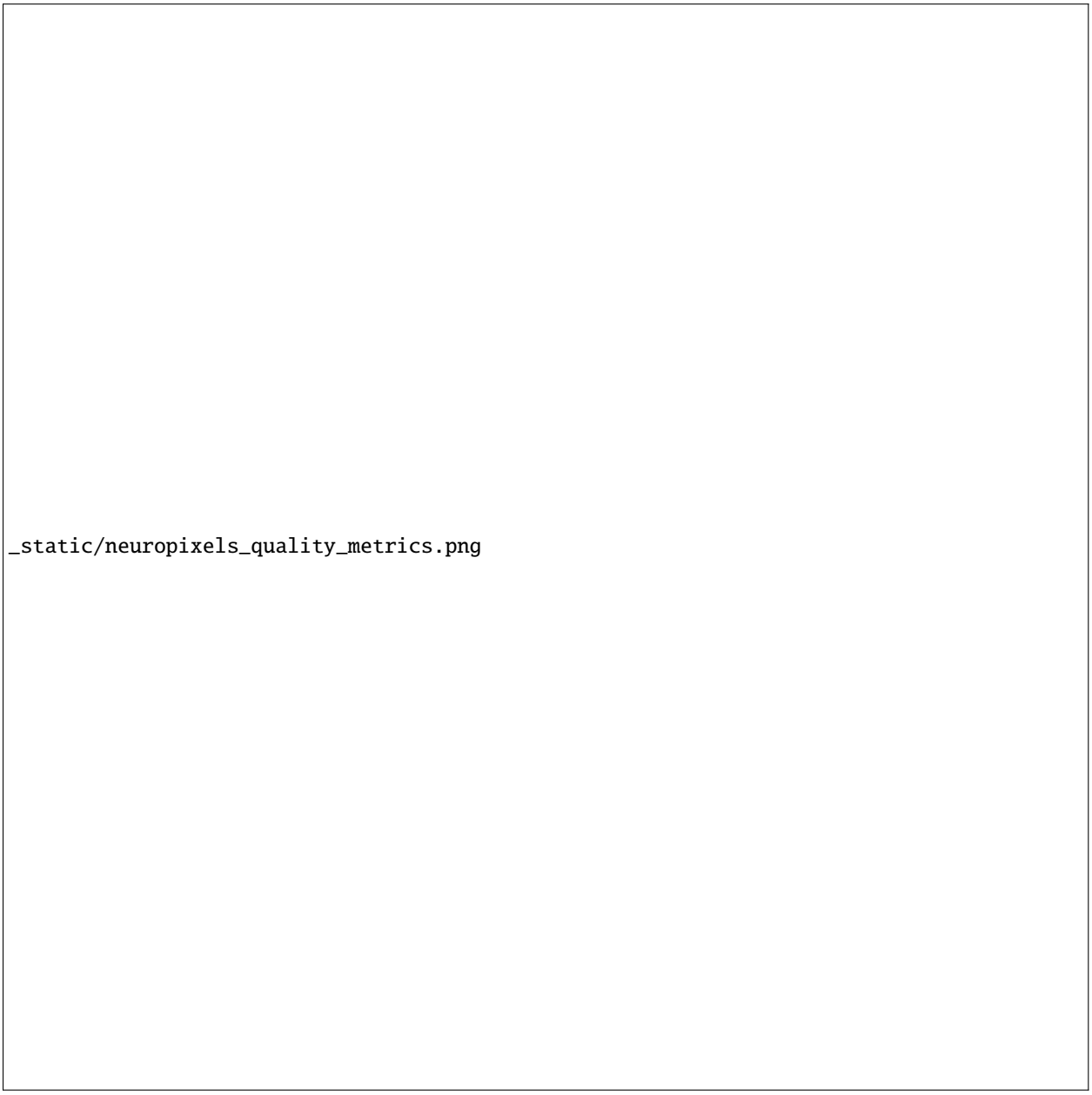
The remainder of the visual stimulus set either consisted of the same stimuli shown in the two-photon experiments ("Brain Observatory 1.1"), or a subset of those stimuli shown with a higher number of repeats. We also added a dot motion stimulus, to allow us to measure the speed tuning of units across the mouse visual system.

2.6.4 AllenSDK 2.0 and Data Compatability

AllenSDK version 2.0 marks a major update to released Visual Coding Neuropixels datasets. Due to newer versions of pynwb/hdmf having issues reading previously released Visual Coding Neuropixels NWB files and due to the significant reorganization of updated NWB file contents, this release contains breaking changes that necessitate a major version revision. NWB files released prior to 6/11/2020 are not guaranteed to work with the 2.0.0 version of AllenSDK. If you cannot or choose not to re-download the updated NWB files, you can continue using a prior version of AllenSDK (< 2.0.0) to access them. However, no further features or bugfixes for AllenSDK (< 2.0.0) are planned. Data released for other projects (Cell Types, Mouse Connectivity, etc.) are *NOT* affected and will *NOT* need to be re-downloaded.

When using the Visual Coding **EcephysProjectCache** from this updated AllenSDK version, if a **ManifestError** is encountered, this indicates that previously downloaded cached data files need to be removed and re-downloaded. The location these files as well as manifest are user defined and are set when instantiating an **EcephysProjectCache**.

2.6.5 Quality Metrics



Every NWB file includes a table of quality metrics, which can be used to assess the completeness, contamination, and stability of units in the recording. By default, we won't show you units below a pre-determined quality threshold; we hide any units that are not present for the whole session (presence_ratio < 0.95), that include many contaminating spikes (isi_violations > 0.5), or are likely missing a large fraction of spikes (amplitude_cutoff > 0.1). However, even contaminated or incomplete units contain information about brain states, and may be of interest to analyze. Therefore, the complete units table can be accessed via special flags in the AllenSDK.

In general, we do not make a distinction between 'single-unit' and 'multi-unit' activity. There is no obvious place to draw a boundary in the overall distributions of quality metrics, and setting a strict cutoff (e.g. isi_violations = 0) will remove a lot of potentially valuable data. We prefer to leave it up to the end user to decide what level of contamination is tolerable. But that means you need to be aware that different units will have different levels of cleanliness.

It should also be noted that all of these metrics assume that the spike waveform is stable throughout the experiment.

Given that the probe drifts, on average, about 40 microns over the course of the ~3 hour recordings, this assumption is almost never valid. The resulting changes in waveform shape can cause a unit's quality to fluctuate. If you're unsure about a unit's quality, it can be helpful to plot its spike amplitudes over time. This can make it obvious if it's drifting below threshold, or if it contains spikes from multiple neurons.

Documentation on the various quality metrics can be found in the ecephys_spike_sorting repository.

For a detailed discussion of the appropriate way to apply each of these metrics, please check out this tutorial (download .ipynb)

2.6.6 Precomputed Stimulus Metrics

Tables of precomputed metrics are available for download to support population analysis and filtering. The table below describes all of the available metrics. The `get_unit_analysis_metrics()` method will load this table as a pandas DataFrame.

Stimulus	Metric	Field Name
drifting gratings	preferred orientation	pref_ori_dg
	preferred temporal frequency	pref_tf_dg
	global ori. selectivity	g_osi_dg
	global dir. selectivity	g_dsi_dg
	running modulation	run_mod_dg
	running modulation p-value	p_run_mod_dg
	firing rate	firing_rate_dg
	fano factor	fano_dg
	modulation index	mod_idx_dg
	f1/f0	f1_f0_dg
	lifetime sparseness	lifetime_sparseness_dg
	c50 (contrast tuning stimulus)	c50_dg
static gratings	preferred orientation	pref_ori_sg
	preferred spatial frequency	pref_sf_sg
	preferred phase	pref_phase_sg
	global ori. selectivity	g_osi_sg
	running modulation	run_mod_sg
	running modulation p-value	p_run_mod_sg
	firing rate	firing_rate_sg
	fano factor	fano_sg
	lifetime sparseness	lifetime_sparseness_sg
natural scenes	preferred image index	pref_image_ns
	image selectivity	image_selectivity_ns
	running modulation	run_mod_ns
	running modulation p-value	p_run_mod_ns
	firing rate	firing_rate_ns
	fano factor	fano_factor_ns
	lifetime sparseness	lifetime_sparseness_ns
dot motion	preferred speed	pref_speed_dm
	preferred direction	pref_dir_dm
	running modulation	run_mod_dm
	running modulation p-value	p_run_mod_dm
	firing rate	firing_rate_dm
	fano factor	fano_factor_dm
	lifetime sparseness	lifetime_sparseness_dm

continues on next page

Table 2 – continued from previous page

Stimulus	Metric	Field Name
full-field flashes	on/off ratio	on_off_ratio_fl
	running modulation	run_mod_fl
	running modulation p-value	p_run_mod_fl
	firing rate	firing_rate_fl
	fano factor	fano_factor_fl
	lifetime sparseness	lifetime_sparseness_fl
gabors	RF area	area_rf
	RF elevation	elevation_rf
	RF azimuth	azimuth_rf
	RF p-value	p_value_rf
	running modulation	run_mod_rf
	running modulation p-value	p_run_mod_rf
	firing rate	firing_rate_rf
	fano factor	fano_factor_rf
	lifetime sparseness	lifetime_sparseness_rf

2.7 GETTING STARTED

2.7.1 Prerequisites

- **install or update the AllenSDK,**
  our Python based toolkit for accessing and working with Allen Institute datasets.
- Pandas familiarity

Data is provided in in NWB format and can be downloaded using the AllenSDK, or accessed directly via an S3 bucket (instructions provided in notebook #1 below). Regardless of which method of file download you choose, we recommend that you load and interact with the data using the tools provided in the AllenSDK, which have been designed to simplify data access and subsequent analysis. No knowledge of the NWB file format is required.

Specific information about how Visual Behavior Optical Physiology data is stored in NWB files and how AllenSDK accesses NWB files can be found here.

2.8 TUTORIALS

To get started, check out these jupyter notebooks to learn how to:

1) Download data using the AllenSDK or directly from our Amazon S3 bucket (download .ipynb) (Open in Colab)
2) Identify experiments of interest using the dataset manifest (download .ipynb) (Open in Colab)
3) Load and visualize data from a 2-photon imaging experiment (download .ipynb) (Open in Colab)
4) Examine the full training history of one mouse (download .ipynb) (Open in Colab)
5) Compare behavior and neural activity across different trial types in the task (download .ipynb) (Open in Colab)

2.9 DOCUMENTATION

To learn more about the experimental design and available data, read through the Visual Behavior and Visual Behavior Ophys chapters in the SWDB Databook.

For a description of available AllenSDK methods and attributes for data access, see this further documentation.

For detailed information about the experimental design, data acquisition, and informatics methods, please refer to our technical whitepaper.

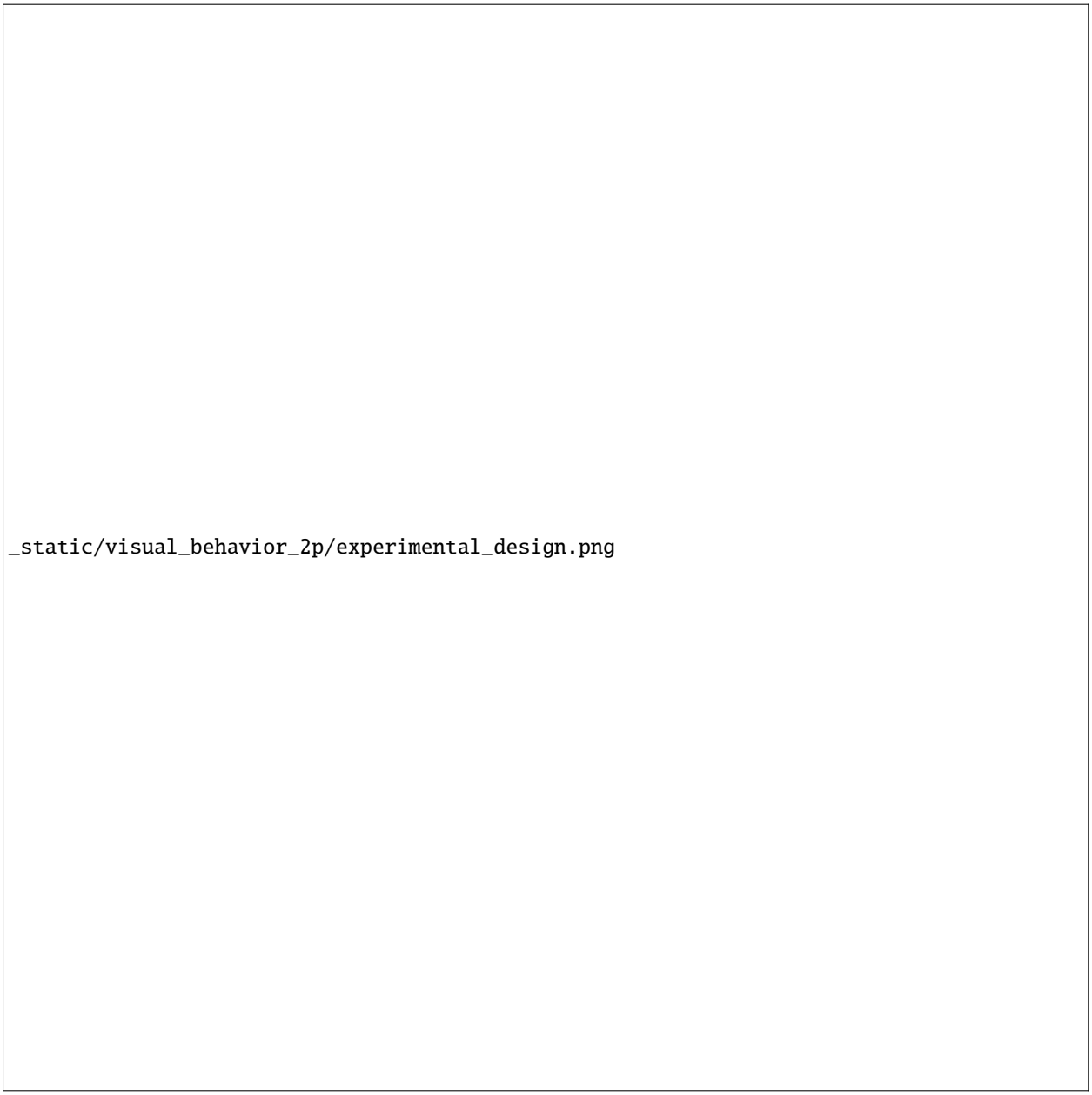
If you have questions about the dataset that aren't addressed by the whitepaper or any of our tutorials, please reach out by posting at https://community.brain-map.org/

2.10 VISUAL BEHAVIOR OPTICAL PHYSIOLOGY DATASETS

The Visual Behavior 2P project used in vivo 2-photon calcium imaging (also called optical physiology, or "ophys") to measure the activity of genetically identified neurons in the visual cortex of mice performing a go/no-go visual change detection task. This dataset can be used to evaluate the influence of experience, expectation, and task engagement on neural coding and dynamics in excitatory and inhibitory cell populations. A description of the experimental design and available data is provided below.

_static/visual_behavior_2p/visual_behavior_2p.png

We used single- and multi-plane imaging approaches to record the activity of populations of neurons across multiple cortical depths and visual areas during change detection behavior. Each population of neurons was imaged repeatedly over multiple days under different sensory and behavioral contexts, including familiar and novel stimuli, as well as active behavior and passive viewing conditions.



Different imaging configurations and stimulus sets were used in different groups of mice, resulting in four unique datasets (indicated by their **project_code** in SDK metadata tables). Two single-plane 2-photon datasets were acquired in the primary visual cortex (VISp). In the *VisualBehavior* dataset, mice were trained with image set A and tested with image set B which was novel to the mice. In the *VisualBehaviorTask1B* dataset, mice were trained with image set B and tested with image set A as the novel image set. One multi-plane dataset (*VisualBehahviorMultiscope*) was acquired at 4 cortical depths in 2 visual areas (VISp & VISl) using image set A for training and image set B for novelty. Another multi-plane dataset (*VisualBehaviorMultiscope4areasx2d*) was acquired at 2 cortical depths in 4 visual areas (VISp, VISl, VISal, VISam). In this dataset, two of the images that became highly familiar during training with image set G were interleaved among novel images in image set H.

For each dataset, we imaged the activity of GCaMP6 expressing cells in populations of excitatory (Slc17a7-IRES2-Cre;Camk2a-tTA;Ai93(TITL-GCaMP6f) or Ai94(TITL-GCaMP6s)), Vip inhibitory (Vip-IRES-Cre;Ai148(TIT2L-GCaMP6f-ICL-tTA2)), and Sst inhibitory (Sst-IRES-Cre;Ai148(TIT2L-GCaMP6f-ICL-tTA2)) neurons. Imaging took place between 75-400um below the cortical surface.

_static/visual_behavior_2p/cre_lines.png

The full dataset includes neural and behavioral measurements from 107 mice during 703 in vivo 2-photon imaging sessions from 326 unique fields of view, resulting in longitudinal recordings from 50,476 cortical neurons. The table below describes the numbers of mice, sessions, and unique recorded neurons for each transgenic line and experimental configuration:

_static/visual_behavior_2p/final_dataset_numbers.png

2.11 BEHAVIORAL TRAINING

Prior to 2-photon imaging, mice were trained to perform a go/no-go visual change detection task in which they learned to lick a spout in response to changes in stimulus identity to earn a water reward. The full behavioral training history of all imaged mice is provided as part of the dataset, allowing investigation into task learning, behavioral strategy, and inter-animal variability. There are a total of 4,787 behavior sessions available for analysis.

We used a standardized procedure to progress mice through a series of training stages, with transitions between stages determined by specific advancement criteria. First, mice learned to detect changes in the orientation of full field static grating stimuli. Next, a 500ms inter stimulus interval period with mean luminance gray screen was added between the 250ms stimulus presentations, incorporating a short-term memory component to the task. Once mice successfully and consistently performed orientation change detection with flashed gratings, they moved to the image change detection version of the task. During image change detection, 8 natural scene images were presented during each behavioral session, for a total of 64 possible image transitions. When behavioral performance again reached criterion (d-prime >1 for 2 out of 3 consecutive days), mice were transitioned to the 2-photon imaging stage in which they performed the task under a microscope to allow simultaneous measurement of neural activity and behavior.

Behavioral training data for mice progressing through these stages of task learning is accessible via the **BehaviorSession** class of the AllenSDK or the `get_behavior_session()` method of the **VisualBehaviorOphysProjectCache**. Each **BehaviorSession** contains the following data streams, event times, and metadata:

- Running speed

- Lick times
- Reward times
- Stimulus presentations
- Behavioral trial information
- Mouse metadata (age, sex, genotype, etc)

2.12 2-PHOTON IMAGING DURING BEHAVIOR

Once mice are well-trained on the image change detection task, they transition to performing the behavior under a 2-photon microscope. Each 2-photon field of view is imaged across multiple session types, allowing measurement of neural activity across different sensory and behavioral contexts.

Mice initially perform the task under the microscope with the same set of images they observed during training, which have become highly familiar (each image is viewed thousands of times during training). Mice also undergo several sessions with a novel image set that they had not seen prior to the 2-photon imaging portion of the experiment. Passive viewing sessions are interleaved between active behavior sessions. On passive days, mice are given their daily water before the session (and are thus satiated) and view the stimulus in open loop mode, with the lick spout retracted to indicate that rewards are not available. This allows investigation of the impact of motivation and attention on patterns of neural activity.

During imaging sessions (but not during training), stimulus presentations are randomly omitted with a 5% probability, resulting in an extended gray screen period between two presentations of the same stimulus and disrupting the expected cadence of stimulus presentations. The change and pre-change stimulus presentations are never omitted. Running speed, pupil diameter, licking, and reward delivery are measured and aligned to neural activity traces.

The **BehaviorOphysExperiment** class in the AllenSDK (or the `get_behavior_ophys_experiment()` method of the **VisualBehaviorOphysProjectCache**) provides all data for a single imaging plane, recorded in a single session, and contains the following data and metadata:

- Maximum intensity image
- Average intensity image
- Segmentation masks and ROI metadata
- dF/F traces (baseline corrected, normalized fluorescence traces)
- Corrected fluorescence traces (neuropil subtracted and demixed, but not normalized)
- Events (detected with an L0 event detection algorithm)
- Pupil position, diameter, and area

- Running speed (in cm/second)
- Lick times
- Reward times
- Stimulus presentation times
- Behavioral trial information
- Mouse metadata (age, sex, genotype, etc)

The data collected in a single continuous recording is defined as a **session** and receives a unique *ophys_session_id*. Each imaging plane in a given session is referred to as an **experiment** and receives a unique *ophys_experiment_id*. For single-plane imaging, there is only one imaging plane (i.e. one experiment) per session. For multi-plane imaging, there can be up to 8 imaging planes (i.e. 8 experiments) per session. Due to our strict QC process, described below, not all multi-plane imaging sessions have exactly 8 experiments, as some imaging planes may not meet our data quality criteria.

_static/visual_behavior_2p/data_structure.png

We aimed to track the activity of single neurons across the session types described above by targeting the same population of neurons over multiple recording sessions, with only one session recorded per day for a given mouse. The collection of imaging sessions for a given population of cells, belonging to a single imaging plane measured across days, is called a **container** and receives a unique *ophys_container_id*. A container can include between 3 and 11 separate sessions for that imaging plane. Mice imaged with the multi-plane 2-photon microscope can have multiple containers, one for each imaging plane recorded across multiple sessions. The session types available for a given container can vary, due to our selection criteria to ensure data quality (described below).

Thus, each mouse can have one or more **containers**, each representing a unique imaging plane (**experiment**) that has been targeted on multiple recording days (**sessions**), under different behavioral and sensory conditions (**session types**).

2.13 SESSION STRUCTURE

During behavioral training, sessions consist of 60 minutes of change detection behavior (other than the *TRAINING_0* sessions, which are 15 minutes of associative reward pairing).

During ophys sessions (*session_type* starting with *OPHYS*), there is a 5 minute period where no stimulus is shown before the change detection task begins, as well as 5 minutes of gray screen after the task ends. This allows evaluation of spontaneous activity in the absence of stimulus or task. After the second 5 minute gray screen period, a 30 second natural movie clip is shown 10 times. This movie clip is the same as the Visual Coding 2P stimulus called *natural_movie_one*. This allows evaluation of stimulus driven activity that is independent of the task.

Ophys session structure:

2.14 DATA PROCESSING

Each 2-photon movie is processed through a series of steps to obtain single cell traces of baseline-corrected fluorescence (dF/F) and detected events, and packaged into the NWB file format along with stimulus and behavioral information, as well as other metadata.

Detailed descriptions of data processing steps can be found in the technical white paper, as well as our data processing repository.

2.15 QUALITY CONTROL

Every 2-photon imaging session was carefully evaluated for a variety of quality control criteria to ensure that the final dataset is of the highest quality possible. Sessions or imaging planes that do not meet our criteria are excluded from the released dataset. These are a few of the key aspects of the data that are evaluated:

- intensity drift
- image saturation or bleaching
- z-drift over the course of a session
- accuracy of session-to-session field of view matching
- excessive or uncorrectable motion in the image
- uncorrectable crosstalk between simultaneously recorded multiscope planes
- errors affecting temporal alignment of data streams
- hardware or software failures
- brain health
- animal stress

2.16 SUMMARY OF AVAILABLE DATA

Behavior	Physiology	Metadata
Running speed	Max intensity projection image	Mouse genotype, age, sex
Licks	Average projection image	Date of acquisition
Rewards	Segmentation mask image	Imaging parameters
Pupil area	Cell specimen table	Task parameters
Pupil position	Cell ROI masks	Session type
Stimulus presentations table	Corrected fluorescence traces	Stimulus images
Trials table	dF/F activity traces	Performance metrics
Stimulus timestamps	Detected events	
	Ophys timestamps	

2.17 DATA FILE CHANGELOG

v1.1.0

Removed data

- **Removed behavior session with incorrect image presentations: 931566300.**
  - **This results in the removal several other subordinate data:**
    - ophys session removed: 931566300
    - ophys experiments removed: 932372699, 932372701, 932372705, 932372707, 932372711
- Removed ophys session 875259383 from the ophys metadata table. The behavior component of this session is available as behavior session id=875471358. No ophys data for this session was previously released.
- Removed truncated behavior sessions with ids: 934610593, 958310218, 975358131, 1011688792

- The above identifiers have also been removed from their respective metadata tables. Columns such as prior_exposures however, are correctly calculated with knowledge of these sessions.
- The cell ROIs associated with the above experiments have also been removed from the cell ROI metadata table.
- **Current data counts**
  - 107 mice (same as v1.0.0)
  - 4079 behavior training session (down from 4082)
  - 703 in vivo 2-photon imaging sessions (down from 704 sessions, previous releases erroneously included session 875259383 in their metadata tables and claimed 705 sessions.)
  - 50,476 logitudinal recordings (down from 50,489)

Metadata Changes

- **Additions to multiple tables**
  - Added project_code and behavior_type (active/passive) value to all tables.
  - Added imaging_plane_group_count, num_depths_per_area, num_targeted_structures, experience_level to Behavior and Ophys session tables.
- **Behavior session table**
  - Added trials summary columns: catch_trial_count, correct_reject_trial_count, engaged_trial_count, false_alarm_trial_count, miss_trial_count, trial_count.
  - Added image_set column.
- **Ophys experiment table**
  - Added targeted_imaging_depth to experiment table, representing the average depth of all experiments in the published container.
- Better consistency of integer typing throughout.

NWB Data Changes

- The value for Age in the metadata, Session/Experiment objects now consistent. NWBs now reflect the age of the animal at the time the session/experiment was taken.
- Enforced better and more consistent typing between the metadata tables and the session metadata.
- All date/times in NWBs and metadata tables are now explicitly UTC timezone.
- Stimulus presentations tables now contain information for additional stimulus conditions, including 10 repeats of a 30 second movie clip at the end of session_types starting with OPHYS, and 5 minute gray screen period before and after change detection behavior in session_types starting with OPHYS. These new stimuli are delineated by stimulus_block and stimulus_block_name. The previously released image behavior stimulus is accessible as the block with a name containing "change_detection". Use the following example code snippet to retrieve the original stimulus block from the pandas table:

  ```
  stimulus_presentations[stimulus_presentations.stimulus_block_name.str.contains('change_detection')]
  ```

  See "SESSION STRUCTURE" section above for more details.
- **New columns in the stimulus_presentations table:**
  - is_image_novel, is_sham_change, movie_frame_index, movie_repeat, stimulus_block, stimulus_block_name, stimulus_name, active
  - See in code documentation for stimulus_presentations table for definitions of these new columns.
- **New trials columns:**

  - change_time, change_frame, response_latency
  - See in code documentation for trials table for definitions of these new columns.

- **Include additional traces in the ophys experiment object:**
  - corrected_fluorescence_traces, demixed traces, neuropil traces, df/f traces
  - corrected_fluorescence_traces are now the correct trace.

Supplemental cache data

- New accessors in VisualBehaviorOphysProjectCache to download and cache the natural movie presented to the mice. Additional accessor to convert the movie to the format as shown on the screen during the session. (Warning this conversion is compute intensive). Methods are:
  - get_raw_natural_movie: Downloads the raw movie if needed and returns as an ndarray.
  - get_natural_movie_template: Converts the raw movie to the format as shown on the screen during the session. Return a pandas dataframe in a similar formate to the image templates. Warning this conversion is compute intensive.

v1.0.1

Metadata corrections - ophys_container_id columns contained extra IDs of incorrect containers.

v1.0.0

New Data

- 107 mice, up from 82
- 4082 behavior training sessions, up from 3021.
- 705 in vivo 2-photon imaging sessions, up from 551.
- 50,489 logitudinal recordings from cortical cells, up from 34,619

Metadata changes

- A new metadata table is present: ophys_cells_table. This table has a project-wide aggregate of cell_specimen_id, cell_roi_id, and ophys_experiment_id.
- Added 'experience_level', 'passive' and 'image_set' columns to ophys_experiments_table

Data Corrections

- 196 BehaviorOphysExperiments had excess invalid ROIs in the dataframe returned by the events field. These have been corrected to remove these invalid ROIs.

v0.3.0

13 sessions were labeled with the wrong session_type in v0.2.0. We have corrected that error. The offending sessions were

behavior_session_id	ophys_session_id	session_type_v0.2.0	session_type_v0.3.0
875020233		OPHYS_3_images_A	OPHYS_2_images_A_passive
902810506		TRAINING_4_images_B_training	TRAINING_3_images_B_10uL_reward
914219174		OPHYS_0_images_B_habituation	TRAINING_5_images_B_handoff_ready
863571063		TRAINING_5_images_A_handoff_ready	TRAINING_1_gratings
974330793		OPHYS_0_images_B_habituation	TRAINING_5_images_B_handoff_ready
863571072		OPHYS_5_images_B_passive	TRAINING_4_images_A_training
1010972317		OPHYS_4_images_A	OPHYS_3_images_B
1011659817		OPHYS_5_images_A_passive	OPHYS_4_images_A
1003302686	1003277121	OPHYS_6_images_A	OPHYS_5_images_A_passive
863571054		OPHYS_7_receptive_field_mappin	TRAINING_5_images_A_epilogue
974282914	974167263	OPHYS_6_images_B	OPHYS_5_images_B_passive
885418521		OPHYS_1_images_A	TRAINING_5_images_A_handoff_lapsed
915739774		OPHYS_1_images_A	OPHYS_0_images_A_habituation

CHAPTER

THREE

MODELS

The Allen SDK currently focuses on models generated from electrophysiology data in the Allen Cell Types Database. There are two classes of models available for download: biophysical models and generalize leaky integrate-and-fire models.

3.1 Generalized LIF Models

The Allen Cell Types Database contains Generalized Leaky Integrate and Fire (GLIF) models that simulate the firing behavior of neurons at five levels of complexity. Review the GLIF technical white paper for details on these models and how their parameters were optimized.

The Allen SDK GLIF simulation module is an explicit time-stepping simulator that evolves a neuron's simulated voltage over the course of an input current stimulus. The module also tracks the neuron's simulated spike threshold and registers action potentials whenever voltage surpasses threshold. Action potentials initiate reset rules that update voltage, threshold, and (optionally) trigger afterspike currents.

The GLIF simulator in this package has a modular architecture that enables users to choose from a number of dynamics and reset rules that update the simulation's voltage, spike threshold, and afterspike currents during the simulation. The GLIF package contains a built-in set of rules, however developers can plug in custom rule implementations provided they follow a simple argument specification scheme.

The Allen SDK GLIF simulator was developed and tested with Python 2.7.9, installed as part of Anaconda Python distribution version 2.1.0.

The rest of this page provides examples demonstrating how to download models, examples of simulating these models, and general GLIF model documentation.

Note: the GLIF simulator module is still under heavy development and may change significantly in the future.

3.1.1 Downloading GLIF Models

There are two ways to download files necessary to run a GLIF model. The first way is to visit http://celltypes.brain-map.org and find cells that have GLIF models available for download. The electrophysiology details page for a cell has a neuronal model download link. Specifically:

1. Click 'More Options +' and filter for GLIF models.
2. Click the electrophysiology thumbnail for a cell on the right hand panel.
3. Choose a GLIF model from the 'Show model responses' dropdown.
4. Scroll down to the model response click 'Download model'.

One such link (for a simple LIF neuronal model, ID 566302806), would look like this:

```
http://api.brain-map.org/neuronal_model/download/566302806
```

This link returns .zip archive containing the neuron configuration file and sweep metadata required to simulate the model with stimuli applied to the cell. Specifically, the .zip archive will contain:

- **472423251_neuron_config.json**: JSON config file for the GLIF model
- **ephys_sweeps.json**: JSON with metadata for sweeps presented to the cell
- **neuronal_model.json**: JSON with general metadata for the cell

If you would like to reproduce the model traces seen in the Cell Types Database, you can download an NWB file containing both the stimulus and cell response traces via a 'Download data' link on the cell's electrophysiology page. See the NWB description section for more details on the NWB file format.

You can also download all of these files, including the cell's NWB file, using the *GlifApi* class:

```
from allensdk.api.queries.glif_api import GlifApi
from allensdk.core.cell_types_cache import CellTypesCache
import allensdk.core.json_utilities as json_utilities

neuronal_model_id = 566302806

# download model metadata
glif_api = GlifApi()
nm = glif_api.get_neuronal_models_by_id([neuronal_model_id])[0]

# download the model configuration file
nc = glif_api.get_neuron_configs([neuronal_model_id])[neuronal_model_id]
neuron_config = glif_api.get_neuron_configs([neuronal_model_id])
json_utilities.write('neuron_config.json', neuron_config)

# download information about the cell
ctc = CellTypesCache()
ctc.get_ephys_data(nm['specimen_id'], file_name='stimulus.nwb')
ctc.get_ephys_sweeps(nm['specimen_id'], file_name='ephys_sweeps.json')
```

3.1.2 Running a GLIF Simulation

To run a GLIF simulation, the most important file you you need is the `neuron_config` JSON file. You can use this file to instantiate a simulator and feed in your own stimulus:

```
import allensdk.core.json_utilities as json_utilities
from allensdk.model.glif.glif_neuron import GlifNeuron

# initialize the neuron
neuron_config = json_utilities.read('neuron_config.json')['566302806']
neuron = GlifNeuron.from_dict(neuron_config)

# make a short square pulse. stimulus units should be in Amps.
stimulus = [ 0.0 ] * 100 + [ 10e-9 ] * 100 + [ 0.0 ] * 100

# important! set the neuron's dt value for your stimulus in seconds
```

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```
neuron.dt = 5e-6

# simulate the neuron
output = neuron.run(stimulus)

voltage = output['voltage']
threshold = output['threshold']
spike_times = output['interpolated_spike_times']
```

Note: The GLIF simulator does not simulate during action potentials. Instead it inserts `NaN` values for a fixed number of time steps when voltage surpasses threshold. The simulator skips `neuron.spike_cut_length` time steps after voltage surpasses threshold.

To reproduce the model's traces displayed on the Allen Cell Types Database web page, the Allen SDK provides the `allensdk.core.model.glif.simulate_neuron` module for simulating all sweeps presented to a cell and storing them in the NWB format:

```
import allensdk.core.json_utilities as json_utilities

from allensdk.model.glif.glif_neuron import GlifNeuron
from allensdk.model.glif.simulate_neuron import simulate_neuron

neuron_config = json_utilities.read('neuron_config.json')['566302806']
ephys_sweeps = json_utilities.read('ephys_sweeps.json')
ephys_file_name = 'stimulus.nwb'

neuron = GlifNeuron.from_dict(neuron_config)

sweep_numbers = [ s['sweep_number'] for s in ephys_sweeps if s['stimulus_units'] == 'Amps
↪' ]
sweep_numbers = sweep_numbers[:1] # for the sake of a speedy example, just run the first␣
↪one
simulate_neuron(neuron, sweep_numbers, ephys_file_name, ephys_file_name, 0.05)
```

Warning: These stimuli are sampled at a very high resolution (200kHz), and a given cell can have many sweeps. This process can take over an hour.

The `simulate_neuron` function call simulates all sweeps in the NWB file. Because the same NWB file is being used for both input and output, the cell's response traces will be overwritten as stimuli are simulated. `simulate_neuron` optionally accepts a value which will be used to overwrite these `NaN` values generated during action potentials (in this case 0.05 Volts).

If you would like to run a single sweep instead of all sweeps, try the following:

```
import allensdk.core.json_utilities as json_utilities
from allensdk.model.glif.glif_neuron import GlifNeuron
from allensdk.core.nwb_data_set import NwbDataSet

neuron_config = json_utilities.read('neuron_config.json')['566302806']
ephys_sweeps = json_utilities.read('ephys_sweeps.json')
```

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```
ephys_file_name = 'stimulus.nwb'

# pull out the stimulus for the current-clamp first sweep
ephys_sweep = next( s for s in ephys_sweeps
                    if s['stimulus_units'] == 'Amps' )
ds = NwbDataSet(ephys_file_name)
data = ds.get_sweep(ephys_sweep['sweep_number'])
stimulus = data['stimulus']

# initialize the neuron
# important! update the neuron's dt for your stimulus
neuron = GlifNeuron.from_dict(neuron_config)
neuron.dt = 1.0 / data['sampling_rate']

# simulate the neuron
output = neuron.run(stimulus)

voltage = output['voltage']
threshold = output['threshold']
spike_times = output['interpolated_spike_times']
```

Note: The `dt` value provided in the downloadable GLIF neuron configuration files does not correspond to the sampling rate of the original stimulus. Stimuli were subsampled and filtered for parameter optimization. Be sure to overwrite the neuron's `dt` with the correct sampling rate.

If you would like to plot the outputs of this simulation using numpy and matplotlib, try:

```
import numpy as np
import matplotlib.pyplot as plt

voltage = output['voltage']
threshold = output['threshold']
interpolated_spike_times = output['interpolated_spike_times']
spike_times = output['interpolated_spike_times']
interpolated_spike_voltages = output['interpolated_spike_voltage']
interpolated_spike_thresholds = output['interpolated_spike_threshold']
grid_spike_indices = output['spike_time_steps']
grid_spike_times = output['grid_spike_times']
after_spike_currents = output['AScurrents']

# create a time array for plotting
time = np.arange(len(stimulus))*neuron.dt

plt.figure(figsize=(10, 10))

# plot stimulus
plt.subplot(3,1,1)
plt.plot(time, stimulus)
plt.xlabel('time (s)')
plt.ylabel('current (A)')
plt.title('Stimulus')
```

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```
# plot model output
plt.subplot(3,1,2)
plt.plot(time,  voltage, label='voltage')
plt.plot(time,  threshold, label='threshold')

if grid_spike_indices is not None:
    plt.plot(interpolated_spike_times, interpolated_spike_voltages, 'x',
             label='interpolated spike')

    plt.plot((grid_spike_indices-1)*neuron.dt, voltage[grid_spike_indices-1], '.',
             label='last step before spike')

plt.xlabel('time (s)')
plt.ylabel('voltage (V)')
plt.legend(loc=3)
plt.title('Model Response')

# plot after spike currents
plt.subplot(3,1,3)
for ii in range(np.shape(after_spike_currents)[1]):
    plt.plot(time, after_spike_currents[:,ii])
plt.xlabel('time (s)')
plt.ylabel('current (A)')
plt.title('After Spike Currents')

plt.tight_layout()
plt.show()
```

Note: There both interpolated spike times and grid spike times. The grid spike is the first time step where the voltage is higher than the threshold. Note that if you try to plot the voltage at the grid spike indices the output will be `NaN`. The interpolated spike is the calculated intersection of the threshold and voltage between the time steps.

3.1.3 GLIF Configuration

Instances of the *GlifNeuron* class require many parameters for initialization. Fixed neuron parameters are stored directly as properties on the class instance:

Parameter	Description	Units	Type
El	resting potential	Volts	float
dt	time duration of each simulation step	seconds	float
R_input	input resistance	Ohms	float
C	capacitance	Farads	float
asc_vector	afterspike current coefficients	Amps	np.array
spike_cut_length	spike duration	time steps	int
th_inf	instantaneous threshold	Volts	float
th_adapt	adapted threshold	Volts	float

Some of these fixed parameters were optimized to fit Allen Cell Types Database electrophysiology data. Optimized

coefficients for these parameters are stored by name in the `neuron.coeffs` dictionary. For more details on which parameters were optimized, please see the technical white paper.

The *GlifNeuron* class has six methods that can be customized: three rules for updating voltage, threshold, and afterspike currents during the simulation; and three rules for updating those values when a spike is detected (voltage surpasses threshold).

Method Type	Description
voltage_dynamics_method	Update simulation voltage for the next time step.
threshold_dynamics_method	Update simulation threshold for the next time step.
AScurrent_dynamics_method	Update afterspike current coefficients for the next time step.
voltage_reset_method	Reset simulation voltage after a spike occurs.
threshold_reset_method	Reset simulation threshold after a spike occurs.
AScurrent_reset_method	Reset afterspike current coefficients after a spike occurs.

The GLIF neuron configuration files available from the Allen Brain Atlas API use built-in methods, however you can supply your own custom method if you like:

```
# define your own custom voltage reset rule
# this one linearly scales the input voltage
def custom_voltage_reset_rule(neuron, voltage_t0, custom_param_a, custom_param_b):
    return custom_param_a * voltage_t0 + custom_param_b

# initialize a neuron from a neuron config file
neuron_config = json_utilities.read('neuron_config.json')['566302806']
neuron = GlifNeuron.from_dict(neuron_config)

# configure a new method and overwrite the neuron's old method
method = neuron.configure_method('custom', custom_voltage_reset_rule,
                                 { 'custom_param_a': 0.1, 'custom_param_b': 0.0 })
neuron.voltage_reset_method = method

output = neuron.run(stimulus)
```

Notice that the function is allowed to take custom parameters (here `custom_param_a` and `custom_param_b`), which are configured on method initialization from a dictionary. For more details, see the documentation for the *GlifNeuron* and *GlifNeuronMethod* classes.

3.1.4 Built-in Dynamics Rules

The job of a dynamics rule is to describe how the simulator should update the voltage, spike threshold, and afterspike currents of the simulator at a given simulation time step.

Voltage Dynamics Rules

These methods update the output voltage of the simulation. They all expect a voltage, afterspike current vector, and current injection value to be passed in by the GlifNeuron. All other function parameters must be fixed using the GlifNeuronMethod class. They all return an updated voltage value.

allensdk.model.glif.glif_neuron_methods.dynamics_voltage_linear_forward_euler()

Threshold Dynamics Rules

These methods update the spike threshold of the simulation. They all expect the current threshold and voltage values of the simulation to be passed in by the GlifNeuron. All other function parameters must be fixed using the GlifNeuronMethod class. They all return an updated threshold value.

allensdk.model.glif.glif_neuron_methods.dynamics_threshold_three_components_exact()

allensdk.model.glif.glif_neuron_methods.dynamics_threshold_spike_component()

allensdk.model.glif.glif_neuron_methods.dynamics_threshold_inf()

Afterspike Current Dynamics Rules

These methods expect current afterspike current coefficients, current time step, and time steps of all previous spikes to be passed in by the GlifNeuron. All other function parameters must be fixed using the GlifNeuronMethod class. They all return an updated afterspike current array.

allensdk.model.glif.glif_neuron_methods.dynamics_AScurrent_exp()

allensdk.model.glif.glif_neuron_methods.dynamics_AScurrent_none()

3.1.5 Built-in Reset Rules

The job of a reset rule is to describe how the simulator should update the voltage, spike threshold, and afterspike currents of the simulator after the simulator has detected that the simulated voltage has surpassed threshold.

Voltage Reset Rules

These methods update the output voltage of the simulation after voltage has surpassed threshold. They all expect a voltageto be passed in by the GlifNeuron. All other function parameters must be fixed using the GlifNeuronMethod class. They all return an updated voltage value.

allensdk.model.glif.glif_neuron_methods.reset_voltage_zero()

allensdk.model.glif.glif_neuron_methods.reset_voltage_v_before()

Threshold Reset Rules

These methods update the spike threshold of the simulation after a spike has been detected. They all expect the current threshold and the reset voltage value of the simulation to be passed in by the GlifNeuron. All other function parameters must be fixed using the GlifNeuronMethod class. They all return an updated threshold value.

allensdk.model.glif.glif_neuron_methods.reset_threshold_inf()

allensdk.model.glif.glif_neuron_methods.reset_threshold_three_components()

Afterspike Reset Reset Rules

These methods expect current afterspike current coefficients to be passed in by the GlifNeuron. All other function parameters must be fixed using the GlifNeuronMethod class. They all return an updated afterspike current array.

allensdk.model.glif.glif_neuron_methods.reset_AScurrent_none()

allensdk.model.glif.glif_neuron_methods.reset_AScurrent_sum()

3.2 Biophysical Models

The Allen Cell Types Database contains biophysical models that characterize the firing behavior of neurons measured in slices through current injection by a somatic whole-cell patch clamp electrode. These models contain a set of 10 active conductances placed at the soma and use the reconstructed 3D morphologies of the modeled neurons. The biophysical modeling technical white paper contains details on the specific construction of these models and the optimization of the model parameters to match the experimentally-recorded firing behaviors.

The biophysical models are run with the NEURON simulation environment. The Allen SDK package contains libraries that assist in downloading and setting up the models available on the Allen Institute web site for users to run using NEURON. The examples and scripts provided run on Linux using the bash shell.

3.2.1 Prerequisites

You must have NEURON with the Python interpreter enabled and the Allen SDK installed.

The Allen Institute perisomatic biophysical models were generated using NEURON version v7.4.rel-1370. Instructions for compiling NEURON with the Python interpreter are available from the NEURON team under the heading Installation with Python as an alternative interpreter. The Allen SDK is compatible with Python version 2.7.9, included in the Anaconda 2.1.0 distribution.

Instructions for optional Docker installation are also available.

Note: Building and installing NEURON with the Python wrapper enabled is not always easy. This page targets users that have a background in NEURON usage and installation.

3.2.2 Downloading Biophysical Models

There are two ways to download files necessary to run a biophysical model. The first way is to visit http://celltypes.brain-map.org and find cells that have biophysical models available for download. The electrophysiology details page for a cell has a neuronal model download link. Specifically:

1. Click 'More Options+'
2. Check 'Models -> Biophysical - perisomatic' or 'Biophysical - all active'
3. Use the Filters, Cell Location and Cell Feature Filters to narrow your results.
4. Click on a Cell Summary to view the Mouse Experiment Electrophysiology.
5. Click the "download data" link to download the NWB stimulus and response file.
6. Click "show model response" and select 'Biophysical - perisomatic' or 'Biophysical - all active'.
7. Scroll down and click the 'Biophysical - perisomatic' or 'Biophysical - all active' "download model" link.

This may be also be done programmatically. The neuronal model id can be found to the left of the corresponding 'Biophysical - perisomatic' or 'Biophysical - all active' "download model" link.

```
from allensdk.api.queries.biophysical_api import BiophysicalApi

bp = BiophysicalApi()
bp.cache_stimulus = True # change to False to not download the large stimulus NWB file
neuronal_model_id = 472451419    # get this from the web site as above
bp.cache_data(neuronal_model_id, working_directory='neuronal_model')
```

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```
```

More help can be found in the online help for the Allen Cell Types Database web application.

3.2.3 Directory Structure

The structure of the directory created looks like this. It includes stimulus files, model parameters, morphology, cellular mechanisms and application configuration.

```
neuronal_model
|-- manifest.json
|-- 472451419_fit.json
|-- Nr5a1-Cre_Ai14_IVSCC_-169248.04.02.01.nwb
|-- Nr5a1-Cre_Ai14_IVSCC_-169248.04.02.01_403165543_m.swc
|-- modfiles
|   |--CaDynamics.mod
|   |--Ca_HVA.mod
|   |--Ca_LVA.mod
|   |--Ih.mod
|   `--...etc.
|
|--x86_64
`---work
```

3.2.4 Running the Simulation (Linux shell prompt)

All of the sweeps available from the web site are included in manifest.json and will be run by default. This can take some time.

```
cd neuronal_model
nrnivmodl ./modfiles   # compile the model (only needs to be done once)
python -m allensdk.model.biophysical.runner manifest.json # perisomatic models
python -m allensdk.model.biophysical.runner manifest.json # legacy all-active models
# new all-active models (axon replaced by a 60 micron long 1 micron diameter stub)
python -m allensdk.model.biophysical.runner manifest.json --axon_type stub
```

3.2.5 Selecting a Specific Sweep

The sweeps are listed in manifest.json. You can remove all of the sweep numbers that you do not want run.

3.2.6 Simulation Main Loop

The top level script is in the *run()* method of the *allensdk.model.biophysical.runner* module. The implementation of the method is discussed here step-by-step:

First configure NEURON based on the configuration file, which was read in from the command line at the very bottom of the script.

run():

```
# configure NEURON -- this will infer model type (perisomatic vs. all-active)
utils = Utils.create_utils(description)
h = utils.h
```

The next step is to get the path of the morphology file and pass it to NEURON.

```
# configure model
manifest = description.manifest
morphology_path = description.manifest.get_path('MORPHOLOGY')
utils.generate_morphology(morphology_path.encode('ascii', 'ignore'))
utils.load_cell_parameters()
```

Then read the stimulus and recording configuration and configure NEURON

```
# configure stimulus and recording
stimulus_path = description.manifest.get_path('stimulus_path')
nwb_out_path = manifest.get_path("output")
output = NwbDataSet(nwb_out_path)
run_params = description.data['runs'][0]
sweeps = run_params['sweeps']
junction_potential = description.data['fitting'][0]['junction_potential']
mV = 1.0e-3
```

Loop through the stimulus sweeps and write the output.

```
# run sweeps
for sweep in sweeps:
    utils.setup_iclamp(stimulus_path, sweep=sweep)
    vec = utils.record_values()

    h.finitialize()
    h.run()

    # write to an NWB File
    output_data = (numpy.array(vec['v']) - junction_potential) * mV
    output.set_sweep(sweep, None, output_data)
```

3.2.7 Customization

Much of the code in the perisomatic simulation is not core Allen SDK code. The runner.py script largely reads the configuration file and calls into methods in the *Utils* class. Utils is a subclass of the *HocUtils* class, which provides access to objects in the NEURON package. The various methods called by the runner.script are implemented here, including: *generate_morphology()*, *load_cell_parameters()*, *setup_iclamp()*, *read_stimulus()* and *record_values()*.

Utils:

```
from allensdk.model.biophys_sim.neuron.hoc_utils import HocUtils

.....

class Utils(HocUtils):
.....

    def __init__(self, description):
        super(Utils, self).__init__(description)
....
```

To create a biophysical model using your own software or data, simply model your directory structure on one of the downloaded simulations or one of the examples below. Add your own runner.py and utils.py module to the simulation directory.

Compile the .mod files using NEURON's nrnivmodl command (Linux shell):

```
nrnivmodl modfiles
```

Then call your runner script directly, passing in the manifest file to your script:

```
python runner.py manifest.json
```

The output from your simulation and any intermediate files will go in the work directory.

3.2.8 Examples

A `minimal example (simple_example.tgz)` and a `multicell example (multicell_example.tgz)` are available to download as a starting point for your own projects.

Each example provides its own utils.py file along with a main script (Linux shell) and supporting configuration files.

simple_example.tgz:

```
tar xvzf simple_example.tgz
cd simple
nrnivmodl modfiles
python simple.py
```

multicell_example.tgz:

```
tar xvzf multicell_example.tgz
cd multicell
nrnivmodl modfiles
python multi.py
python multicell_diff.py
```

3.2.9 Exporting Output to Text Format or Image

This is an example of using the AllenSDK to save a response voltage to other formats.

```
from allensdk.core.dat_utilities import \
    DatUtilities
from allensdk.core.nwb_data_set import \
    NwbDataSet
import numpy as np
import matplotlib
matplotlib.use("Agg")
import matplotlib.pyplot as plt

nwb_file = '313862020.nwb'
sweep_number = 52
dat_file = '313862020_%d.dat' % (sweep_number)

nwb = NwbDataSet(nwb_file)
sweep = nwb.get_sweep(sweep_number)

# read v and t as numpy arrays
v = sweep['response']
dt = 1.0e3 / sweep['sampling_rate']
num_samples = len(v)
t = np.arange(num_samples) * dt

# save as text file
data = np.transpose(np.vstack((t, v)))
with open (dat_file, "w") as f:
    np.savetxt(f, data)

# save image using matplotlib
fig, ax = plt.subplots(nrows=1, ncols=1)
ax.plot(t, v)
ax.set_title("Sweep %s" % (sweep_number))
fig.savefig('out.png')
```

3.2.10 Model Description Files

Basic Structure

A model description file is simply a JSON object with several sections at the top level and an array of JSON objects within each section.

```
{
   "cell_section": [
       {
         "name": "cell 1",
         "shape": "pyramidal"
         "position": [ 0.1, 0.2, 0.3 ]
       },
       {
```

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```
            "name": "cell 2",
            "shape": "glial",
            "position": [ 0.1, 0.2, 0.3 ]
         }
    ],
    "extra": [
       { "what": "wood",
         "who": "woodchuck"
       }
    ]
}
```

Even if a section contains no objects or only one object the array brackets must be present.

Objects Within Sections

While no restrictions are enforced on what kinds of objects are stored in a section, some rules of thumb make the file easier to work with.

1. All objects within a section are the same structure. Common operations on a section are to display it as a table, iterate over it, load from or write to a spreadsheet or csv file. These operations are all easier if the section is fairly homogeneous.
2. Objects are not deeply nested. While some shallow nesting is often useful, deep nesting such as a tree structure is not recommended. It makes interoperability with other tools and data formats more difficult.
3. Arrays are allowed, though they should not be deeply nested either.
4. Object member values should be literals. Do not use pickled classes, for example.

Comment Lines

The JSON specification does not allow comments. However, the Allen SDK library applies a preprocessing stage to remove C++-style comments, so they can be used in description files.

Multi-line comments should be surrounded by /* */ and single-line comments start with //. Commented description files will not be recognized by strict json parsers unless the comments are stripped.

commented.json:

```
{
   /*
    *  multi-line comment
    */
   "section1": [
       {
          "name": "simon"  // single line comment
       }]
   }
```

Split Description Files by Section

A model description can be split into multiple files by putting some sections in one file and other sections into another file. This can be useful if you want to put a topology of cells and connections in one file and experimental conditions and stimulus in another file. The resulting structure in memory will behave the same way as if the files were not split. This allows a small experiment to be described in a single file and large experiments to be more modular.

cells.json:

```
{
   "cell_section": [
       {
         "name": "cell 1",
         "shape": "pyramidal"
         "position": [ 0.1, 0.2, 0.3 ]
       },
       {
         "name": "cell 2",
         "shape": "glial",
         "position": [ 0.1, 0.2, 0.3 ]
       }
   ]
}
```

extras.json:

```
{
    "extra": [
       {
         "what": "wood",
         "who": "woodchuck"
       }
    ]
}
```

Split Sections Between Description Files

If two description files containing the same sections are combined, the resulting description will contain objects from both files. This feature allows sub-networks to be described in separate files. The sub-networks can then be composed into a larger network with an additional description of the interconnections.

network1.json:

```
/* A self-contained sub-network */
{
    "cells": [
        { "name": "cell1" },
        { "name": "cell2" }
    ],
    /* intra-network connections /*
    "connections": [
        { "source": "cell1", "target" : "cell2" }
```

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```
    ]
}
```

network2.json:

```
/* Another self-contained sub-network */
{
    "cells": [
        { "name": "cell3" },
        { "name": "cell4" }
    ],
    "connections": [
        { "source": "cell3", "target" : "cell4" }
    ]
}
```

interconnect.json:

```
{
    // the additional connections needed to
    // connect the network1 and network2
    // into a ring topology.
    "connections": [
       { "source": "cell2", "target": "cell3" },
       { "source": "cell4", "target": "cell1" }
    ]
}
```

3.2.11 Resource Manifest

JSON has many advantages. It is widely supported, readable and easy to parse and edit. As data sets get larger or specialized those advantages diminish. Large or complex models and experiments generally need more than a single model description file to completely describe an experiment. A manifest file is a way to describe all of the resources needed within the Allen SDK description format itself.

The manifest section is named "manifest" by default, though it is configurable. The objects in the manifest section each specify a directory, file, or file pattern. Files and directories may be organized in a parent-child relationship.

A Simple Manifest

This is a simple manifest file that specifies the BASEDIR directory using ".", meaning the current directory:

```
{
    "manifest": [
        {   "key": "BASEDIR",
            "type": "dir",
            "spec": "."
        }
    ] }
}
```

Parent Child Relationships

Adding the optional "parent_key" member to a manifest object creates a parent-child relation. In this case WORKDIR will be found in "./work":

```
{
    "manifest": [
        {   "key": "BASEDIR",
            "type": "dir",
            "spec": "."
        },
        {   "key": "WORKDIR",
            "type": "dir",
            "spec": "/work",
            "parent_key": "BASEDIR"
        }
    ] }
}
```

File Spec Patterns

Files can be specified using the type "file" instead of "dir". If a sequence of many files is needed, the spec may contain patterns to indicate where the sequence number (%d) or string (%s) will be interpolated:

```
{
    "manifest": [
        {   "key": "BASEDIR",
            "type": "dir",
            "spec": "."
        },
        {
            "key": "voltage_out_cell_path",
            "type": "file",
            "spec": "v_out-cell-%d.dat",
            "parent_key": "BASEDIR"
        }
    ] }
}
```

Split Manifest Files

Manifest files can be split like any description file. This allows the specification of a general directory structure in a shared file and specific files in a separate configuration (i.e. stimulus and working directory)

Extensions

To date, manifest description files have not been used to reference URLs that provide model data, but it is a planned future use case.

3.2.12 Further Reading

- NEURON
- Python
- JSON

CHAPTER

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EXAMPLES

Take a look at the table below for a list of SDK example notebooks and scripts.

Description	Link
Introduction to the Mouse Connectivity Atlas	Jupyter notebook (download .ipynb)
Introduction to the Cell Types Database	Jupyter notebook (download .ipynb)
Introduction to the Brain Observatory	Jupyter notebook (download .ipynb)
Brain Observatory Stimulus Manipulation	Jupyter notebook (download .ipynb)
Brain Observatory Tuning Analysis	Jupyter notebook (download .ipynb)
Brain Observatory Receptive Field Analysis	Jupyter notebook (download .ipynb)
Brain Observatory Cell Specimen ID Mapping	Jupyter notebook (download .ipynb)
Brain Observatory Monitor	Jupyter notebook (download .ipynb)
Dynamic Brain Workshop 2015 experiment detail	Jupyter notebook (download .ipynb)
Stimulating a biophysical model with a square pulse	Jupyter notebook (download .ipynb)
Using a Reference Space	Jupyter notebook (download .ipynb)
Downloading Images	Jupyter notebook (download .ipynb)
Visual Coding Neuropixels Quick Start	Jupyter notebook (download .ipynb)
Visual Coding Neuropixels Reference	Jupyter notebook (download .ipynb)

CHAPTER

FIVE

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CHAPTER

SIX

ALLENSDK PACKAGE

6.1 Subpackages

6.1.1 allensdk.api package

Subpackages

allensdk.api.cloud_cache package

Submodules

allensdk.api.cloud_cache.cloud_cache module

allensdk.api.cloud_cache.file_attributes module

class allensdk.api.cloud_cache.file_attributes.**CacheFileAttributes**(*url: str*, *version_id: str*, *file_hash: str*, *local_path: Path*)

Bases: object

This class will contain the attributes of a remotely stored file so that they can easily and consistently be passed around between the methods making up the remote file cache and manifest classes

Parameters

url: str
The full URL of the remote file

version_id: str
A string specifying the version of the file (probably calculated by S3)

file_hash: str
The (hexadecimal) file hash of the file

local_path: pathlib.Path
The path to the location where the file's local copy should be stored (probably computed by the Manifest class)

property file_hash: str

property local_path: Path

property url: str

property version_id: str

allensdk.api.cloud_cache.manifest module

class allensdk.api.cloud_cache.manifest.**Manifest**(*cache_dir: str | Path*, *json_input*, *use_static_project_dir: bool = False*)

Bases: object

A class for loading and manipulating the online manifest.json associated with a dataset release

Each Manifest instance should represent the data for 1 and only 1 manifest.json file.

Parameters

cache_dir: str or pathlib.Path
The path to the directory where local copies of files will be stored

json_input:
A ''.read()"-supporting file-like object containing a JSON document to be deserialized (i.e. same as the first argument to json.load)

use_static_project_dir: bool
When determining what the local path of a remote resource (data or metadata file) should be, the Manifest class will typically create a versioned project subdirectory under the user provided *cache_dir* (e.g. f"{cache_dir}/{project_name}-{manifest_version}") to allow the possibility of multiple manifest (and data) versions to be used. In certain cases, like when using a project's s3 bucket directly as the cache_dir, the project directory name needs to be static (e.g. f"{cache_dir}/{project_name}"). When set to True, the Manifest class will use a static project directory to determine local paths for remote resources. Defaults to False.

data_file_attributes(*file_id*) → *CacheFileAttributes*

Return the CacheFileAttributes associated with a data file

Parameters

file_id:
The identifier of the data file whose attributes are to be returned. Must be a key in self._data['data_files']

Raises

RuntimeError
If you try to run this method when self._data is None (meaning you haven't yet loaded a manifest.json file)

ValueError
If the file_id is not a valid option

property file_id_column

The column in the metadata files used to uniquely identify data files

property file_id_values

List of valid file_id values

metadata_file_attributes(*metadata_file_name: str*) → *CacheFileAttributes*

Return the CacheFileAttributes associated with a metadata file

Parameters

metadata_file_name: str
Name of the metadata file. Must be in self.metadata_file_names

Raises

RuntimeError
If you try to run this method when self._data is None (meaning you haven't yet loaded a manifest.json)

ValueError
If the metadata_file_name is not a valid option

`property metadata_file_names`
List of metadata file names associated with this dataset

`property project_name`
The name of the project whose data and metadata files this manifest tracks.

`property version`
The version of the dataset currently loaded

allensdk.api.cloud_cache.utils module

`allensdk.api.cloud_cache.utils.bucket_name_from_url(url: str) → str | None`
Read in a URL and return the name of the AWS S3 bucket it points towards.

Parameters

URL: str
A generic URL, suitable for retrieving an S3 object via an HTTP GET request.

Returns

str
An AWS S3 bucket name. Note: if 's3.amazonaws.com' does not occur in the URL, this method will return None and emit a warning.

`allensdk.api.cloud_cache.utils.file_hash_from_path(file_path: str | Path) → str`
Return the hexadecimal file hash for a file

Parameters

file_path: Union[str, Path]
path to a file

Returns

str:
The file hash (Blake2b; hexadecimal) of the file

`allensdk.api.cloud_cache.utils.relative_path_from_url(url: str) → str`
Read in a url and return the relative path of the object

Parameters

url: str
The url of the object whose path you want

Returns

str:
Relative path of the object

Notes

This method returns a str rather than a pathlib.Path because it is used to get the S3 object Key from a URL. If using Pathlib.path on a Windows system, the '/' will get transformed into '', confusing S3.

Module contents

allensdk.api.queries package

Submodules

allensdk.api.queries.annotated_section_data_sets_api module

class allensdk.api.queries.annotated_section_data_sets_api.**AnnotatedSectionDataSetsApi**(*base_uri=None*)

Bases: *RmaApi*

See: Searching Annotated SectionDataSets

get_annotated_section_data_sets(*structures*, *intensity_values=None*, *density_values=None*, *pattern_values=None*, *age_names=None*)

For a list of target structures, find the SectionDataSet that matches the parameters for intensity_values, density_values, pattern_values, and Age.

Parameters

structure_graph_id
[dict of integers] what to retrieve

intensity_values
[array of strings, optional] 'High','Low', 'Medium' (default)

density_values
[array of strings, optional] 'High', 'Low'

pattern_values
[array of strings, optional] 'Full'

age_names
[array of strings, options] for example 'E11.5', '13.5'

Returns

data
[dict] The parsed JSON repsonse message.

Notes

This method uses the non-RMA Annotated SectionDataSet endpoint.

get_annotated_section_data_sets_via_rma(*structures*, *intensity_values=None*, *density_values=None*, *pattern_values=None*, *age_names=None*)

For a list of target structures, find the SectionDataSet that matches the parameters for intensity_values, density_values, pattern_values, and Age.

Parameters

structure_graph_id
[dict of integers] what to retrieve

intensity_values
[array of strings, optional] intensity values, 'High','Low', 'Medium' (default)

density_values
[array of strings, optional] density values, 'High', 'Low'

pattern_values
[array of strings, optional] pattern values, 'Full'

age_names
[array of strings, options] for example 'E11.5', '13.5'

Returns

data
[dict] The parsed JSON response message.

Notes

This method uses the RMA endpoint to search annotated SectionDataSet data.

`get_compound_annotated_section_data_sets`(*queries*, *fmt='json'*)

Find the SectionDataSet that matches several annotated_section_data_sets queries linked together with a Boolean 'and' or 'or'.

Parameters

queries
[array of dicts] dicts with args like build_query

fmt
[string, optional] 'json' or 'xml'

Returns

data
[dict] The parsed JSON repsonse message.

allensdk.api.queries.biophysical_api module

`class` allensdk.api.queries.biophysical_api.`BiophysicalApi`(*base_uri=None*)

Bases: *RmaTemplate*

`BIOPHYSICAL_MODEL_TYPE_IDS = (491455321, 329230710)`

`build_rma`(*neuronal_model_id*, *fmt='json'*)

Construct a query to find all files related to a neuronal model.

Parameters

neuronal_model_id
[integer or string representation] key of experiment to retrieve.

fmt
[string, optional] json (default) or xml

Returns

string
RMA query url.

`cache_data`(*neuronal_model_id*, *working_directory=None*)

Take a an experiment id, query the Api RMA to get well-known-files download the files, and store them in the working directory.

Parameters

neuronal_model_id
[int or string representation] found in the neuronal_model table in the api

working_directory
[string] Absolute path name where the downloaded well-known files will be stored.

`create_manifest`(*fit_path=''*, *model_type=''*, *stimulus_filename=''*, *swc_morphology_path=''*, *marker_path=''*, *sweeps=[]*)

Generate a json configuration file with parameters for a a biophysical experiment.

Parameters

fit_path
[string] filename of a json configuration file with cell parameters.

stimulus_filename
[string] path to an NWB file with input currents.

swc_morphology_path
[string] file in SWC format.

sweeps
[array of integers] which sweeps in the stimulus file are to be used.

`get_neuronal_models`(*specimen_ids*, *num_rows='all'*, *count=False*, *model_type_ids=None*, ***kwargs*)

Fetch all of the biophysically detailed model records associated with a particular specimen_id

Parameters

specimen_ids
[list] One or more integer ids identifying specimen records.

num_rows
[int, optional] how many records to retrieve. Default is 'all'.

count
[bool, optional] If True, return a count of the lines found by the query. Default is False.

model_type_ids
[list, optional] One or more integer ids identifying categories of neuronal model. Defaults to all-active and perisomatic biophysical_models.

Returns

List of dict
Each element is a biophysical model record, containing a unique integer id, the id of the associated specimen, and the id of the model type to which this model belongs.

`get_well_known_file_ids`(*neuronal_model_id*)

Query the current RMA endpoint with a neuronal_model id to get the corresponding well known file ids.

Returns

list
A list of well known file id strings.

`is_well_known_file_type`(*wkf*, *name*)

Check if a structure has the expected name.

Parameters

wkf
[dict] A well-known-file structure with nested type information.

name
[string] The expected type name

See also:

read_json
where this helper function is used.

`read_json`(*json_parsed_data*)

Get the list of well_known_file ids from a response body containing nested sample,microarray_slides,well_known_files.

Parameters

json_parsed_data
[dict] Response from the Allen Institute Api RMA.

Returns

list of strings
Well known file ids.

```
rma_templates = {'model_queries': [{'name': 'models_by_specimen', 'description':
'see name', 'model': 'NeuronalModel', 'num_rows': 'all', 'count': False,
'criteria': '[neuronal_model_template_id$in{{biophysical_model_types}}],
[specimen_id$in{{specimen_ids}}]', 'criteria_params': ['specimen_ids',
'biophysical_model_types']}]}
```

allensdk.api.queries.brain_observatory_api module

allensdk.api.queries.cell_types_api module

`class` allensdk.api.queries.cell_types_api.`CellTypesApi`(*base_uri=None*)

Bases: *RmaApi*

`HUMAN = 'Homo Sapiens'`

`MARKER_FILE_TYPE = '3DNeuronMarker'`

`MOUSE = 'Mus musculus'`

`NWB_FILE_TYPE = 'NWBDownload'`

`SWC_FILE_TYPE = '3DNeuronReconstruction'`

filter_cells(*cells*, *require_morphology*, *require_reconstruction*, *reporter_status*, *species*)

Filter a list of cell specimens to those that optionally have morphologies or have morphological reconstructions.

Parameters

cells: list
List of cell metadata dictionaries to be filtered

require_morphology: boolean
Filter out cells that have no morphological images.

require_reconstruction: boolean
Filter out cells that have no morphological reconstructions.

reporter_status: list
Filter for cells that have a particular cell reporter status

species: list
Filter for cells that belong to one or more species. If None, return all. Must be one of [CellTypesApi.MOUSE, CellTypesApi.HUMAN].

filter_cells_api(*cells*, *require_morphology=False*, *require_reconstruction=False*, *reporter_status=None*, *species=None*, *simple=True*)

get_cell(*id*)

Query the API for a one cells in the Cell Types Database.

Returns

list
Meta data for one cell.

get_ephys_features()

Query the API for the full table of EphysFeatures for all cells.

get_ephys_sweeps(*specimen_id*)

Query the API for a list of sweeps for a particular cell in the Cell Types Database.

Parameters

specimen_id: int
Specimen ID of a cell.

Returns

list: List of sweep dictionaries belonging to a cell

get_morphology_features()

Query the API for the full table of morphology features for all cells

Notes

by default the tags column is removed because it isn't useful

`list_cells`(*id=None*, *require_morphology=False*, *require_reconstruction=False*, *reporter_status=None*, *species=None*)

Query the API for a list of all cells in the Cell Types Database.

Parameters

id: int
ID of a cell. If not provided returns all matching cells.

require_morphology: boolean
Only return cells that have morphology images.

require_reconstruction: boolean
Only return cells that have morphological reconstructions.

reporter_status: list
Return cells that have a particular cell reporter status.

species: list
Filter for cells that belong to one or more species. If None, return all. Must be one of [CellTypesApi.MOUSE, CellTypesApi.HUMAN].

Returns

list
Meta data for all cells.

`list_cells_api`(*id=None*, *require_morphology=False*, *require_reconstruction=False*, *reporter_status=None*, *species=None*)

`save_ephys_data`(*specimen_id*, *file_name*)

Save the electrophysology recordings for a cell as an NWB file.

Parameters

specimen_id: int
ID of the specimen, from the Specimens database model in the Allen Institute API.

file_name: str
Path to save the NWB file.

`save_reconstruction`(*specimen_id*, *file_name*)

Save the morphological reconstruction of a cell as an SWC file.

Parameters

specimen_id: int
ID of the specimen, from the Specimens database model in the Allen Institute API.

file_name: str
Path to save the SWC file.

`save_reconstruction_markers`(*specimen_id*, *file_name*)

Save the marker file for the morphological reconstruction of a cell. These are comma-delimited files indicating points of interest in a reconstruction (truncation points, early tracing termination, etc).

Parameters

specimen_id: int
ID of the specimen, from the Specimens database model in the Allen Institute API.

file_name: str
Path to save the marker file.

simplify_cells_api(*cells*)

allensdk.api.queries.connected_services module

class allensdk.api.queries.connected_services.**ConnectedServices**

Bases: object

A class representing a schema of informatics web services.

Notes

See Connected Services and Pipes for a human-readable list of services and parameters.

The URL format is documented at Service Pipelines.

Connected Services only include API services that are accessed via the RMA endpoint using an rma::services stage.

ARRAY = 'array'

BOOLEAN = 'boolean'

FLOAT = 'float'

INTEGER = 'integer'

STRING = 'string'

build_url(*service_name*, *kwargs*)

Create a single stage RMA url from a service name and parameters.

class property schema

Dictionary of service names and parameters.

Notes

See Connected Services and Pipes for a human-readable list of connected services and their parameters.

allensdk.api.queries.glif_api module

class allensdk.api.queries.glif_api.**GlifApi**(*base_uri=None*)

Bases: *RmaTemplate*

GLIF_TYPES = [395310498, 395310469, 395310475, 395310479, 471355161]

NWB_FILE_TYPE = None

cache_stimulus_file(*output_file_name*)

DEPRECATED Download the NWB file for the current neuronal model and save it to a file.

Parameters

output_file_name: string
File name to store the NWB file.

get_ephys_sweeps()

DEPRECATED Retrieve ephys sweep information out of downloaded metadata for a neuronal model

Returns

list
A list of sweeps metadata dictionaries

get_neuron_config(*output_file_name=None*)

DEPRECATED Retrieve a model configuration file from the API, optionally save it to disk, and return the contents of that file as a dictionary.

Parameters

output_file_name: string
File name to store the neuron configuration (optional).

get_neuron_configs(*neuronal_model_ids=None*)

get_neuronal_model(*neuronal_model_id*)

DEPRECATED Query the current RMA endpoint with a neuronal_model id to get the corresponding well known files and meta data.

Returns

dict
A dictionary containing

get_neuronal_model_templates()

get_neuronal_models(*ephys_experiment_ids=None*)

get_neuronal_models_by_id(*neuronal_model_ids=None*)

list_neuronal_models()

DEPRECATED Query the API for a list of all GLIF neuronal models.

Returns

list
Meta data for all GLIF neuronal models.

```
rma_templates = {'glif_queries': [{'name': 'neuronal_model_templates',
'description': 'see name', 'model': 'NeuronalModelTemplate', 'num_rows': 'all',
'count': False}, {'name': 'neuronal_models', 'description': 'see name', 'model':
'Specimen', 'include':
'neuronal_models(well_known_files,neuronal_model_template[id$in395310498,395310469,
395310475,395310479,471355161],neuronal_model_runs(well_known_files))', 'criteria':
'{% if ephys_experiment_ids is defined %}[id$in{{ ephys_experiment_ids
}}]{%endif%}', 'num_rows': 'all', 'criteria_params': ['ephys_experiment_ids'],
'count': False}, {'name': 'neuron_config', 'description': 'see name', 'model':
'NeuronalModel', 'include': 'well_known_files(well_known_file_type)', 'criteria':
'[id$in{{ neuronal_model_ids }}]', 'num_rows': 'all', 'criteria_params':
['neuronal_model_ids'], 'count': False}]}
```

allensdk.api.queries.grid_data_api module

class allensdk.api.queries.grid_data_api.**GridDataApi**(*resolution=None*, *base_uri=None*)

Bases: *RmaApi*

HTTP Client for the Allen 3-D Expression Grid Data Service.

See: Downloading 3-D Expression Grid Data

DATA_MASK = 'data_mask'

DENSITY = 'density'

ENERGY = 'energy'

INJECTION_DENSITY = 'injection_density'

INJECTION_ENERGY = 'injection_energy'

INJECTION_FRACTION = 'injection_fraction'

INTENSITY = 'intensity'

PROJECTION_DENSITY = 'projection_density'

PROJECTION_ENERGY = 'projection_energy'

download_alignment3d(*section_data_set_id*, *num_rows='all'*, *count=False*, ***kwargs*)

Download the parameters of the 3D affine tranformation mapping this section data set's image-space stack to CCF-space (or vice-versa).

Parameters

section_data_set_id
[int] download the parameters for this data set.

Returns

dict
parameters of this section data set's alignment3d

download_deformation_field(*section_data_set_id*, *header_path=None*, *voxel_path=None*, *voxel_type='DeformationFieldVoxels'*, *header_type='DeformationFieldHeader'*)

Download the local alignment parameters for this dataset. This a 3D vector image (3 components) describing a deformable local mapping from CCF voxels to this section data set's affine-aligned image stack.

Parameters

section_data_set_id
[int] Download the deformation field for this data set

header_path
[str, optional] If supplied, the deformation field header will be downloaded to this path.

voxel_path
[str, optiona] If supplied, the deformation field voxels will be downloaded to this path.

voxel_type
[str] WellKnownFileType of this dataset's data file

header_type
[str] WellKnownFileType of this dataset's header file

`download_expression_grid_data`(*section_data_set_id*, *include=None*, *path=None*)

Download in zipped metaimage format.

Parameters

section_data_set_id
[integer] What to download.

include
[list of strings, optional] Image volumes. 'energy' (default), 'density', 'intensity'.

path
[string, optional] File name to save as.

Returns

file
[3-D expression grid data packaged into a compressed archive file (.zip).]

`download_gene_expression_grid_data`(*section_data_set_id*, *volume_type*, *path*)

Download a metaimage file containing registered gene expression grid data

Parameters

section_data_set_id
[int] Download data from this experiment

volume_type
[str] Download this type of data (options are GridDataApi.ENERGY, GridDataApi.DENSITY, GridDataApi.INTENSITY)

path
[str] Download to this path

`download_projection_grid_data`(*section_data_set_id*, *image=None*, *resolution=None*, *save_file_path=None*)

Download in NRRD format.

Parameters

section_data_set_id
[integer] What to download.

image
[list of strings, optional] Image volume. 'projection_density', 'projection_energy', 'injection_fraction', 'injection_density', 'injection_energy', 'data_mask'.

resolution
[integer, optional] in microns. 10, 25, 50, or 100 (default).

save_file_path
[string, optional] File name to save as.

Notes

See Downloading 3-D Projection Grid Data for additional documentation.

allensdk.api.queries.image_download_api module

class allensdk.api.queries.image_download_api.**ImageDownloadApi**(*base_uri=None*)

Bases: *RmaTemplate*

HTTP Client to download whole or partial two-dimensional images from the Allen Institute with the SectionImage, AtlasImage and ProjectionImage Download Services.

See Downloading an Image for more documentation.

COLORMAPS = {'aba': 8, 'aibsmap_alt': 9, 'blue': 6, 'colormap': 10, 'expression': 4, 'gray': 0, 'green': 7, 'hotmetal': 1, 'jet': 2, 'projection': 11, 'red': 5, 'redtemp': 3}

atlas_image_query(*atlas_id*, *image_type_name=None*)

List atlas images belonging to a specified atlas

Parameters

atlas_id
[integer, optional] Find images from this atlas.

image_type_name
[string, optional] Restrict response to images of this type. If not provided, the query will get it from the atlas id.

Returns

list of dict
Each element is an AtlasImage record.

Notes

See Downloading Atlas Images and Graphics for additional documentation. *allensdk.api.queries.ontologies_api.OntologiesApi.get_atlases()* can also be used to list atlases along with their ids.

download_atlas_image(*atlas_image_id*, *file_path=None*, ***kwargs*)

download_image(*image_id*, *file_path=None*, *endpoint=None*, ***kwargs*)

Download whole or partial two-dimensional images from the Allen Institute with the SectionImage or AtlasImage service.

Parameters

image_id
[integer] SubImage to download.

file_path
[string, optional] where to put it, defaults to image_id.jpg

downsample
[int, optional] Number of times to downsample the original image.

quality
[int, optional] jpeg quality of the returned image, 0 to 100 (default)

expression
[boolean, optional] Request the expression mask for the SectionImage.

view
[string, optional] 'expression', 'projection', 'tumor_feature_annotation' or 'tumor_feature_boundary'

top
[int, optional] Index of the topmost row of the region of interest.

left :int, optional
Index of the leftmost column of the region of interest.

width
[int, optional] Number of columns in the output image.

height
[int, optional] Number of rows in the output image.

range
[list of ints, optional] Filter to specify the RGB channels. low,high,low,high,low,high

colormap
[list of floats, optional] Filter to specify the RGB channels. [lower_threshold,colormap] gain 0-1, colormap id is a string from ImageDownloadApi.COLORMAPS

rgb
[list of floats, optional] Filter to specify the RGB channels. [red,green,blue] 0-1

contrast
[list of floats, optional] Filter to specify contrast parameters. [gain,bias] 0-1

annotation
[boolean, optional] Request the annotated AtlasImage

atlas
[int, optional] Specify the desired Atlas' annotations.

projection
[boolean, optional] Request projection for the specified image.

downsample_dimensions
[boolean, optional] Indicates if the width and height should be adjusted to account for downsampling.

Returns

None
the file is downloaded and saved to the path.

Notes

By default, an unfiltered full-sized image with the highest quality is returned as a download if no parameters are provided.

'downsample=1' halves the number of pixels of the original image both horizontally and vertically. range_list = kwargs.get('range', None)

Specifying 'downsample=2' quarters the height and width values.

Quality must be an integer from 0, for the lowest quality, up to as high as 100. If it is not specified, it defaults to the highest quality.

Top is specified in full-resolution (largest tier) pixel coordinates. SectionImage.y is the default value.

Left is specified in full-resolution (largest tier) pixel coordinates. SectionImage.x is the default value.

Width is specified in tier-resolution (desired tier) pixel coordinates. SectionImage.width is the default value. It is automatically adjusted when downsampled.

Height is specified in tier-resolution (desired tier) pixel coordinates. SectionImage.height is the default value. It is automatically adjusted when downsampled.

The range parameter consists of 6 comma delimited integers that define the lower (0) and upper (4095) bound for each channel in red-green-blue order (i.e. "range=0,1500,0,1000,0,4095"). The default range values can be determined by referring to the following fields on the Equalization model associated with the SectionDataSet: red_lower, red_uppper, green_lower, green_upper, blue_lower, blue_upper. For more information, see the Image Controls section of the Allen Mouse Brain Connectivity Atlas: Projection Dataset help topic. See: `Image Download Service `<http://help.brain-map.org/display/api/Downloading+an+Image>_

download_projection_image(*projection_image_id*, *file_path=None*, ***kwargs*)

download_section_image(*section_image_id*, *file_path=None*, ***kwargs*)

get_section_data_sets_by_product(*product_ids*, *include_failed=False*, *num_rows='all'*, *count=False*, ***kwargs*)

List all of the section data sets produced as part of one or more products

Parameters

product_ids
[list of int] Integer specifiers for Allen Institute products. A product is a set of related data.

include_failed
[bool, optional] If True, find both failed and passed datasets. Default is False

num_rows
[int, optional] how many records to retrieve. Default is 'all'.

count
[bool, optional] If True, return a count of the lines found by the query. Default is False.

Returns

list of dict
Each returned element is a section data set record.

Notes

See http://api.brain-map.org/api/v2/data/query.json?criteria=model::Product for a list of products.

get_section_image_ranges(*section_image_ids*, *num_rows='all'*, *count=False*, *as_lists=True*, ***kwargs*)

Section images from the Mouse Connectivity Atlas are displayed on connectivity.brain-map.org after having been linearly windowed and leveled. This method obtains parameters defining channelwise upper and lower bounds of the windows used for one or more images.

Parameters

section_image_ids
[list of int] Each element is a unique identifier for a section image.

num_rows
[int, optional] how many records to retrieve. Default is 'all'.

count
[bool, optional] If True, return a count of the lines found by the query. Default is False.

as_lists
[bool, optional] If True, return the window parameters in a list, rather than a dict (this is the format of the range parameter on ImageDownloadApi.download_image). Default is False.

Returns

list of dict or list of list
For each section image id provided, return the window bounds for each channel.

```
rma_templates = {'image_queries': [{'name': 'section_image_ranges', 'description':
'see name', 'model': 'Equalization', 'num_rows': 'all', 'count': False, 'only':
['blue_lower', 'blue_upper', 'red_lower', 'red_upper', 'green_lower',
'green_upper'], 'criteria': 'section_data_set(section_images[id$in{{
section_image_ids }}])', 'criteria_params': ['section_image_ids']}, {'name':
'section_images_by_data_set_id', 'description': 'see name', 'model':
'SectionImage', 'num_rows': 'all', 'count': False, 'criteria': '[data_set_id$eq{{
data_set_id }}]', 'criteria_params': ['data_set_id']}, {'name':
'section_data_sets_by_product_id', 'description': 'see name', 'model':
'SectionDataSet', 'num_rows': 'all', 'count': False, 'criteria':
'[failed$in{{failed}}],products[id$in{{ product_ids }}]', 'criteria_params':
['product_ids', 'failed']}]}
```

section_image_query(*section_data_set_id*, *num_rows='all'*, *count=False*, ***kwargs*)

List section images belonging to a specified section data set

Parameters

atlas_id
[integer, optional] Find images from this section data set.

num_rows
[int] how many records to retrieve. Default is 'all'

count
[bool] If True, return a count of the lines found by the query.

Returns

list of dict
Each element is an SectionImage record.

Notes

The SectionDataSet model is used to represent single experiments which produce an array of images. This includes Mouse Connectivity and Mouse Brain Atlas experiments, among other projects. You may see references to the ids of experiments from those projects. These are the same as section data set ids.

allensdk.api.queries.mouse_atlas_api module

class allensdk.api.queries.mouse_atlas_api.**MouseAtlasApi**(*base_uri=None*)

Bases: *ReferenceSpaceApi*, *GridDataApi*

Downloads Mouse Brain Atlas grid data, reference volumes, and metadata.

DEVMOUSE_ATLAS_PRODUCTS = (3,)

HUMAN_ORGANISM = (1,)

MOUSE_ATLAS_PRODUCTS = (1,)

MOUSE_ORGANISM = (2,)

download_expression_density(*path*, *experiment_id*)

download_expression_energy(*path*, *experiment_id*)

download_expression_intensity(*path*, *experiment_id*)

get_genes(*organism_ids=None*, *chromosome_ids=None*, ***kwargs*)

Download a list of genes

Parameters

organism_ids
[list of int, optional] Filter genes to those appearing in these organisms. Defaults to mouse (2).

chromosome_ids
[list of int, optional] Filter genes to those appearing on these chromosomes. Defaults to all.

Returns

list of dict:
Each element is a gene record, with a nested chromosome record (also a dict).

get_section_data_sets(*gene_ids=None*, *product_ids=None*, ***kwargs*)

Download a list of section data sets (experiments) from the Mouse Brain Atlas project.

Parameters

gene_ids
[list of int, optional] Filter results based on the genes whose expression was characterized in each experiment. Default is all.

product_ids
[list of int, optional] Filter results to a subset of products. Default is the Mouse Brain Atlas.

Returns

list of dict
Each element is a section data set record, with one or more gene records nested in a list.

allensdk.api.queries.mouse_connectivity_api module

class allensdk.api.queries.mouse_connectivity_api.**MouseConnectivityApi**(*base_uri=None*)

Bases: *ReferenceSpaceApi*, *GridDataApi*

HTTP Client for the Allen Mouse Brain Connectivity Atlas.

See: Mouse Connectivity API

PRODUCT_IDS = [5, 31]

build_reference_aligned_image_channel_volumes_url(*data_set_id*)

Construct url to download the red, green, and blue channels aligned to the 25um adult mouse brain reference space volume.

Parameters

data_set_id
[integerallensdk.api.queries] aka attachable_id

Notes

See: Reference-aligned Image Channel Volumes for additional documentation.

calculate_injection_centroid(*injection_density*, *injection_fraction*, *resolution=25*)

Compute the centroid of an injection site.

Parameters

injection_density: np.ndarray
The injection density volume of an experiment

injection_fraction: np.ndarray
The injection fraction volume of an experiment

download_data_mask(*path*, *experiment_id*, *resolution*)

download_injection_density(*path*, *experiment_id*, *resolution*)

download_injection_fraction(*path*, *experiment_id*, *resolution*)

download_projection_density(*path*, *experiment_id*, *resolution*)

download_reference_aligned_image_channel_volumes(*data_set_id*, *save_file_path=None*)

Returns

The well known file is downloaded

experiment_correlation_search(***kwargs*)

Select a seed experiment and a domain over which the similarity comparison is to be made.

Parameters

row
[integer] SectionDataSet.id to correlate against.

structures
[list of integers or strings, optional] Integer Structure.id or String Structure.acronym.

hemisphere
[string, optional] Use ‘right’ or ‘left’. Defaults to both hemispheres.

transgenic_lines
[list of integers or strings, optional] Integer TransgenicLine.id or String TransgenicLine.name. Specify ID 0 to exclude all TransgenicLines.

injection_structures
[list of integers or strings, optional] Integer Structure.id or String Structure.acronym.

primary_structure_only
[boolean, optional]

product_ids
[list of integers, optional] Integer Product.id

start_row
[integer, optional] For paging purposes. Defaults to 0.

num_rows
[integer, optional] For paging purposes. Defaults to 2000.

Notes

See Correlation Search and service::mouse_connectivity_correlation.

`experiment_injection_coordinate_search`(***kwargs*)

User specifies a seed location within the 3D reference space. The service returns a rank list of experiments by distance of its injection site to the specified seed location.

Parameters

seed_point
[list of floats] The coordinates of a point in 3-D SectionDataSet space.

transgenic_lines
[list of integers or strings, optional] Integer TransgenicLine.id or String TransgenicLine.name. Specify ID 0 to exclude all TransgenicLines.

injection_structures
[list of integers or strings, optional] Integer Structure.id or String Structure.acronym.

primary_structure_only
[boolean, optional]

product_ids
[list of integers, optional] Integer Product.id

start_row
[integer, optional] For paging purposes. Defaults to 0.

num_rows
[integer, optional] For paging purposes. Defaults to 2000.

Notes

See Injection Coordinate Search and service::mouse_connectivity_injection_coordinate.

experiment_source_search(***kwargs*)

Search over the whole projection signal statistics dataset to find experiments with specific projection profiles.

Parameters

injection_structures
[list of integers or strings] Integer Structure.id or String Structure.acronym.

target_domain
[list of integers or strings, optional] Integer Structure.id or String Structure.acronym.

injection_hemisphere
[string, optional] 'right' or 'left', Defaults to both hemispheres.

target_hemisphere
[string, optional] 'right' or 'left', Defaults to both hemispheres.

transgenic_lines
[list of integers or strings, optional] Integer TransgenicLine.id or String TransgenicLine.name. Specify ID 0 to exclude all TransgenicLines.

injection_domain
[list of integers or strings, optional] Integer Structure.id or String Structure.acronym.

primary_structure_only
[boolean, optional]

product_ids
[list of integers, optional] Integer Product.id

start_row
[integer, optional] For paging purposes. Defaults to 0.

num_rows
[integer, optional] For paging purposes. Defaults to 2000.

Notes

See Source Search, Target Search, and service::mouse_connectivity_injection_structure.

experiment_spatial_search(***kwargs*)

Displays all SectionDataSets with projection signal density >= 0.1 at the seed point. This service also returns the path along the most dense pixels from the seed point to the center of each injection site..

Parameters

seed_point
[list of floats] The coordinates of a point in 3-D SectionDataSet space.

transgenic_lines
[list of integers or strings, optional] Integer TransgenicLine.id or String TransgenicLine.name. Specify ID 0 to exclude all TransgenicLines.

section_data_sets
[list of integers, optional] Ids to filter the results.

injection_structures
[list of integers or strings, optional] Integer Structure.id or String Structure.acronym.

primary_structure_only
[boolean, optional]

product_ids
[list of integers, optional] Integer Product.id

start_row
[integer, optional] For paging purposes. Defaults to 0.

num_rows
[integer, optional] For paging purposes. Defaults to 2000.

Notes

See Spatial Search and service::mouse_connectivity_target_spatial.

`get_experiment_detail`(*experiment_id*)

Retrieve the experiments data.

`get_experiments`(*structure_ids*, ***kwargs*)

Fetch experiment metadata from the Mouse Brain Connectivity Atlas.

Parameters

structure_ids
[integer or list, optional] injection structure

Returns

url
[string] The constructed URL

`get_experiments_api`()

Fetch experiment metadata from the Mouse Brain Connectivity Atlas via the ApiConnectivity table.

Returns

url
[string] The constructed URL

`get_manual_injection_summary`(*experiment_id*)

Retrieve manual injection summary.

`get_projection_image_info`(*experiment_id*, *section_number*)

Fetch meta-information of one projection image.

Parameters

experiment_id
[integer]

section_number
[integer]

Notes

See: image examples under Experimental Overview and Metadata for additional documentation. Download the image using *allensdk.api.queries.image_download_api.ImageDownloadApi.download_section_image()*

get_reference_aligned_image_channel_volumes_url(*data_set_id*)

Retrieve the download link for a specific data set. Notes —– See Reference-aligned Image Channel Volumes for additional documentation.

get_structure_unionizes(*experiment_ids*, *is_injection=None*, *structure_name=None*, *structure_ids=None*, *hemisphere_ids=None*, *normalized_projection_volume_limit=None*, *include=None*, *debug=None*, *order=None*)

allensdk.api.queries.ontologies_api module

class allensdk.api.queries.ontologies_api.**OntologiesApi**(*base_uri=None*)

Bases: *RmaTemplate*

See: Atlas Drawings and Ontologies

get_atlases()

get_atlases_table(*atlas_ids=None*, *brief=True*)

List Atlases available through the API with associated ontologies and structure graphs.

Parameters

atlas_ids
[integer or list of integers, optional] only select specific atlases

brief
[boolean, optional] True (default) requests only name and id fields.

Returns

dict
[atlas metadata]

Notes

This query is based on the table of available Atlases. See also: Class: Atlas

get_structure_graphs()

get_structure_sets(*structure_set_ids=None*)

get_structures(*structure_graph_ids=None*, *structure_graph_names=None*, *structure_set_ids=None*, *structure_set_names=None*, *order=['structures.graph_order']*, *num_rows='all'*, *count=False*, ***kwargs*)

Retrieve data about anatomical structures.

Parameters

structure_graph_ids
[int or list of ints, optional] database keys to get all structures in particular graphs

structure_graph_names
[string or list of strings, optional] list of graph names to narrow the query

structure_set_ids
[int or list of ints, optional] database keys to get all structures in a particular set

structure_set_names
[string or list of strings, optional] list of set names to narrow the query.

order
[list of strings] list of RMA order clauses for sorting

num_rows
[int] how many records to retrieve

Returns

dict
the parsed json response containing data from the API

Notes

Only one of the methods of limiting the query should be used at a time.

`get_structures_with_sets`(*structure_graph_ids*, *order=['structures.graph_order']*, *num_rows='all'*, *count=False*, ***kwargs*)

Download structures along with the sets to which they belong.

Parameters

structure_graph_ids
[int or list of int] Only fetch structure records from these graphs.

order
[list of strings] list of RMA order clauses for sorting

num_rows
[int] how many records to retrieve

Returns

dict
the parsed json response containing data from the API

```
rma_templates = {'ontology_queries': [{'name': 'structures_by_graph_ids',
'description': 'see name', 'model': 'Structure', 'criteria': '[graph_id$in{{
graph_ids }}]', 'order': ['structures.graph_order'], 'num_rows': 'all', 'count':
False, 'criteria_params': ['graph_ids']}, {'name': 'structures_by_graph_names',
'description': 'see name', 'model': 'Structure', 'criteria':
'graph[structure_graphs.name$in{{ graph_names }}]', 'order':
['structures.graph_order'], 'num_rows': 'all', 'count': False, 'criteria_params':
['graph_names']}, {'name': 'structures_by_set_ids', 'description': 'see name',
'model': 'Structure', 'criteria': '[structure_set_id$in{{ set_ids }}]', 'order':
['structures.graph_order'], 'num_rows': 'all', 'count': False, 'criteria_params':
['set_ids']}, {'name': 'structures_by_set_names', 'description': 'see name',
'model': 'Structure', 'criteria': 'structure_sets[name$in{{ set_names }}]',
'order': ['structures.graph_order'], 'num_rows': 'all', 'count': False,
'criteria_params': ['set_names']}, {'name': 'structure_graphs_list',
'description': 'see name', 'model': 'StructureGraph', 'num_rows': 'all', 'count':
False}, {'name': 'structure_sets_list', 'description': 'see name', 'model':
'StructureSet', 'num_rows': 'all', 'count': False}, {'name': 'atlases_list',
'description': 'see name', 'model': 'Atlas', 'num_rows': 'all', 'count': False},
{'name': 'atlases_table', 'description': 'see name', 'model': 'Atlas',
'criteria': '{% if atlas_ids is defined %}[id$in{{ atlas_ids
}}],{%endif%}structure_graph(ontology),graphic_group_labels', 'include':
'structure_graph(ontology),graphic_group_labels', 'only': ['atlases.id',
'atlases.name', 'atlases.image_type', 'ontologies.id', 'ontologies.name',
'structure_graphs.id', 'structure_graphs.name', 'graphic_group_labels.id',
'graphic_group_labels.name'], 'num_rows': 'all', 'count': False,
'criteria_params': ['atlas_ids']}, {'name': 'structures_with_sets', 'description':
'see name', 'model': 'Structure', 'include': 'structure_sets', 'criteria':
'[graph_id$in{{ graph_ids }}]', 'order': ['structures.graph_order'], 'num_rows':
'all', 'count': False, 'criteria_params': ['graph_ids']}, {'name':
'structure_sets_by_id', 'description': 'see name', 'model': 'StructureSet',
'criteria': '[id$in{{ set_ids }}]', 'num_rows': 'all', 'count': False,
'criteria_params': ['set_ids']}]}
```

unpack_structure_set_ancestors(*structure_dataframe*)

Convert a slash-separated structure_id_path field to a list.

Parameters

structure_dataframe
[DataFrame] structure data from the API

Returns

None
A new column is added to the dataframe containing the ancestor list.

allensdk.api.queries.reference_space_api module

class allensdk.api.queries.reference_space_api.**ReferenceSpaceApi**(*base_uri=None*)

Bases: *RmaApi*

ARA_NISSL = 'ara_nissl'

AVERAGE_TEMPLATE = 'average_template'

CCF_2015 = 'annotation/ccf_2015'

CCF_2016 = 'annotation/ccf_2016'

CCF_2017 = 'annotation/ccf_2017'

CCF_VERSION_DEFAULT = 'annotation/ccf_2017'

DEVMOUSE_2012 = 'annotation/devmouse_2012'

MOUSE_2011 = 'annotation/mouse_2011'

VOXEL_RESOLUTION_100_MICRONS = 100

VOXEL_RESOLUTION_10_MICRONS = 10

VOXEL_RESOLUTION_25_MICRONS = 25

VOXEL_RESOLUTION_50_MICRONS = 50

build_volumetric_data_download_url(*data_path*, *file_name*, *voxel_resolution=None*, *release=None*, *coordinate_framework=None*)

Construct url to download 3D reference model in NRRD format.

Parameters

data_path
[string] 'average_template', 'ara_nissl', 'annotation/ccf_{year}', 'annotation/mouse_2011', or 'annotation/devmouse_2012'

voxel_resolution
[int] 10, 25, 50 or 100

coordinate_framework
[string] 'mouse_ccf' (default) or 'mouse_annotation'

Notes

See: 3-D Reference Models for additional documentation.

download_annotation_volume(*ccf_version*, *resolution*, *file_name*)

Download the annotation volume at a particular resolution.

Parameters

ccf_version: string
Which reference space version to download. Defaults to "annotation/ccf_2017"

resolution: int
Desired resolution to download in microns. Must be 10, 25, 50, or 100.

file_name: string
Where to save the annotation volume.

Note: the parameters must be used as positional parameters, not keywords

`download_mouse_atlas_volume`(*age*, *volume_type*, *file_name*)

Download a reference volume (annotation, grid annotation, atlas volume) from the mouse brain atlas project

Parameters

age
[str] Specify a mouse age for which to download the reference volume

volume_type
[str] Specify the type of volume to download

file_name
[str] Specify the path to the downloaded volume

`download_structure_mask`(*structure_id*, *ccf_version*, *resolution*, *file_name*)

Download an indicator mask for a specific structure.

Parameters

structure_id
[int] Unique identifier for the annotated structure

ccf_version
[string] Which reference space version to download. Defaults to "annotation/ccf_2017"

resolution
[int] Desired resolution to download in microns. Must be 10, 25, 50, or 100.

file_name
[string] Where to save the downloaded mask.

`download_structure_mesh`(*structure_id*, *ccf_version*, *file_name*)

Download a Wavefront obj file containing a triangulated 3d mesh built from an annotated structure.

Parameters

structure_id
[int] Unique identifier for the annotated structure

ccf_version
[string] Which reference space version to download. Defaults to "annotation/ccf_2017"

file_name
[string] Where to save the downloaded mask.

`download_template_volume`(*resolution*, *file_name*)

Download the registration template volume at a particular resolution.

Parameters

resolution: int
Desired resolution to download in microns. Must be 10, 25, 50, or 100.

file_name: string
Where to save the registration template volume.

`download_volumetric_data`(*data_path*, *file_name*, *voxel_resolution=None*, *save_file_path=None*, *release=None*, *coordinate_framework=None*)

Download 3D reference model in NRRD format.

Parameters

data_path
[string] 'average_template', 'ara_nissl', 'annotation/ccf_{year}', 'annotation/mouse_2011', or 'annotation/devmouse_2012'

file_name
[string] server-side file name. 'annotation_10.nrrd' for example.

voxel_resolution
[int] 10, 25, 50 or 100

coordinate_framework
[string] 'mouse_ccf' (default) or 'mouse_annotation'

Notes

See: 3-D Reference Models for additional documentation.

allensdk.api.queries.rma_api module

class allensdk.api.queries.rma_api.**RmaApi**(*base_uri=None*)

Bases: *Api*

See: RESTful Model Access (RMA)

ALL = 'all'

COUNT = 'count'

CRITERIA = 'rma::criteria'

DEBUG = 'debug'

EQ = '$eq'

EXCEPT = 'except'

EXCPT = 'excpt'

FALSE = 'false'

INCLUDE = 'rma::include'

IS = '$is'

MODEL = 'model::'

NUM_ROWS = 'num_rows'

ONLY = 'only'

OPTIONS = 'rma::options'

ORDER = 'order'

PIPE = 'pipe::'

PREVIEW = 'preview'

SERVICE = 'service::'

START_ROW = 'start_row'

TABULAR = 'tabular'

TRUE = 'true'

build_query_url(*stage_clauses*, *fmt='json'*)

Combine one or more RMA query stages into a single RMA query.

Parameters

stage_clauses
[list of strings] subqueries

fmt
[string, optional] json (default), xml, or csv

Returns

string
complete RMA url

build_schema_query(*clazz=None*, *fmt='json'*)

Build the URL that will fetch the data schema.

Parameters

clazz
[string, optional] Name of a specific class or None (default).

fmt
[string, optional] json (default) or xml

Returns

url
[string] The constructed URL

Notes

If a class is specified, only the schema information for that class will be requested, otherwise the url requests the entire schema.

debug_clause(*debug_value=None*)

Construct a debug clause for use in an rma::options clause. Parameters ———- debug_value : string or boolean

True, False, None (default) or 'preview'

Returns

clause
[string] The query clause for inclusion in an RMA query URL.

Notes

True will request debugging information in the response. False will request no debugging information. None will return an empty clause. 'preview' will request debugging information without the query being run.

`filter`(*key*, *value*)

serialize a single RMA query filter clause.

Parameters

key
[string] keys for narrowing a query.

value
[string] value for narrowing a query.

Returns

string
a single filter clause for an RMA query string.

`filters`(*filters*)

serialize RMA query filter clauses.

Parameters

filters
[dict] keys and values for narrowing a query.

Returns

string
filter clause for an RMA query string.

`get_schema`(*clazz=None*)

Retrieve schema information.

`model_query`(**args*, ***kwargs*)

Construct and execute a model stage of an RMA query string.

Parameters

model
[string] The top level data type

filters
[dict] key, value comparisons applied to the top-level model to narrow the results.

criteria
[string] raw RMA criteria clause to choose what object are returned

include
[string] raw RMA include clause to return associated objects

only
[list of strings, optional] to be joined into an rma::options only filter to limit what data is returned

except
[list of strings, optional] to be joined into an rma::options except filter to limit what data is returned

excpt
[list of strings, optional] synonym for except parameter to avoid a reserved word conflict.

tabular
[list of string, optional] return columns as a tabular data structure rather than a nested tree.

count
[boolean, optional] False to skip the extra database count query.

debug
[string, optional] 'true', 'false' or 'preview'

num_rows
[int or string, optional] how many database rows are returned (may not correspond directly to JSON tree structure)

start_row
[int or string, optional] which database row is start of returned data (may not correspond directly to JSON tree structure)

Notes

See RMA Path Syntax for a brief overview of the normalized RMA syntax. Normalized RMA syntax differs from the legacy syntax used in much of the RMA documentation. Using the &debug=true option with an RMA URL will include debugging information in the response, including the normalized query.

`model_stage`(*model*, ***kwargs*)

Construct a model stage of an RMA query string.

Parameters

model
[string] The top level data type

filters
[dict] key, value comparisons applied to the top-level model to narrow the results.

criteria
[string] raw RMA criteria clause to choose what object are returned

include
[string] raw RMA include clause to return associated objects

only
[list of strings, optional] to be joined into an rma::options only filter to limit what data is returned

except
[list of strings, optional] to be joined into an rma::options except filter to limit what data is returned

tabular
[list of string, optional] return columns as a tabular data structure rather than a nested tree.

count
[boolean, optional] False to skip the extra database count query.

debug
[string, optional] 'true', 'false' or 'preview'

num_rows
[int or string, optional] how many database rows are returned (may not correspond directly to JSON tree structure)

start_row
[int or string, optional] which database row is start of returned data (may not correspond directly to JSON tree structure)

Notes

See RMA Path Syntax for a brief overview of the normalized RMA syntax. Normalized RMA syntax differs from the legacy syntax used in much of the RMA documentation. Using the &debug=true option with an RMA URL will include debugging information in the response, including the normalized query.

`only_except_tabular_clause`(*filter_type*, *attribute_list*)

Construct a clause to filter which attributes are returned for use in an rma::options clause.

Parameters

filter_type
[string] 'only', 'except', or 'tabular'

attribute_list
[list of strings] for example ['acronym', 'products.name', 'structure.id']

Returns

clause
[string] The query clause for inclusion in an RMA query URL.

Notes

The title of tabular columns can be set by adding '+as+<title>' to the attribute. The tabular filter type requests a response that is row-oriented rather than a nested structure. Because of this, the tabular option can mask the lazy query behavior of an rma::include clause. The tabular option does not mask the inner-join behavior of an rma::include clause. The tabular filter is required for .csv format RMA requests.

`options_clause`(***kwargs*)

build rma:: options clause.

Parameters

only
[list of strings, optional]

except
[list of strings, optional]

tabular
[list of string, optional]

count
[boolean, optional]

debug
[string, optional] 'true', 'false' or 'preview'

num_rows
[int or string, optional]

start_row
[int or string, optional]

`order_clause`(*order_list=None*)

Construct a debug clause for use in an rma::options clause.

Parameters

order_list
[list of strings] for example ['acronym', 'products.name+asc', 'structure.id+desc']

Returns

clause
[string] The query clause for inclusion in an RMA query URL.

Notes

Optionally adding '+asc' (default) or '+desc' after an attribute will change the sort order.

`pipe_stage`(*pipe_name*, *parameters*)

Connect model and service stages via their JSON responses.

Notes

See: Service Pipelines and Connected Services and Pipes

`quote_string`(*the_string*)

Wrap a clause in single quotes.

Parameters

the_string
[string] a clause to be included in an rma query that needs to be quoted

Returns

string
input wrapped in single quotes

`service_query`(**args*, ***kwargs*)

Construct and Execute a single-stage RMA query to send a request to a connected service.

Parameters

service_name
[string] Name of a documented connected service.

parameters
[dict] key-value pairs as in the online documentation.

Notes

See: Service Pipelines and Connected Services and Pipes

service_stage(*service_name*, *parameters=None*)

Construct an RMA query fragment to send a request to a connected service.

Parameters

service_name
[string] Name of a documented connected service.

parameters
[dict] key-value pairs as in the online documentation.

Notes

See: Service Pipelines and Connected Services and Pipes

tuple_filters(*filters*)

Construct an RMA filter clause.

Notes

See RMA Path Syntax - Square Brackets for Filters for additional documentation.

allensdk.api.queries.rma_pager module

class allensdk.api.queries.rma_pager.**RmaPager**

Bases: object

static pager(*fn*, **args*, ***kwargs*)

allensdk.api.queries.rma_pager.**pageable**(*total_rows=None*, *num_rows=None*)

allensdk.api.queries.rma_template module

class allensdk.api.queries.rma_template.**RmaTemplate**(*base_uri=None*, *query_manifest=None*)

Bases: *RmaApi*

See: Atlas Drawings and Ontologies

template_query(*template_name*, *entry_name*, ***kwargs*)

to_filter_rhs(*rhs*)

allensdk.api.queries.svg_api module

class allensdk.api.queries.svg_api.**SvgApi**(*base_uri=None*)

Bases: *Api*

build_query(*section_image_id*, *groups=None*, *download=False*)

Build the URL that will fetch meta data for the specified structure.

Parameters

section_image_id
[integer] Key of the object to be retrieved.

groups
[array of integers] Keys of the group labels to filter the svg types that are returned.

Returns

url
[string] The constructed URL

download_svg(*section_image_id*, *groups=None*, *file_path=None*)

Download the svg file

get_svg(*section_image_id*, *groups=None*)

Get the svg document.

allensdk.api.queries.synchronization_api module

class allensdk.api.queries.synchronization_api.**SynchronizationApi**(*base_uri=None*)

Bases: *Api*

HTTP client for image synchronization services uses the image alignment results from the Informatics Data Processing Pipeline. Note: all locations on SectionImages are reported in pixel coordinates and all locations in 3-D ReferenceSpaces are reported in microns.

See Image to Image Synchronization for additional documentation.

get_image_to_atlas(*section_image_id*, *x*, *y*, *atlas_id*)

For a specified Atlas, find the closest annotated SectionImage and (x,y) location as defined by a seed SectionImage and seed (x,y) location.

Parameters

section_image_id
[integer] Seed for spatial sync.

x
[float] Pixel coordinate of the seed location in the seed SectionImage.

y
[float] Pixel coordinate of the seed location in the seed SectionImage.

atlas_id
[int] Target Atlas for image sync.

Returns

dict
The parsed json response

`get_image_to_image`(*section_image_id*, *x*, *y*, *section_data_set_ids*)

For a list of target SectionDataSets, find the closest SectionImage and (x,y) location as defined by a seed SectionImage and seed (x,y) pixel location.

Parameters

section_image_id
[integer] Seed for spatial sync.

x
[float] Pixel coordinate of the seed location in the seed SectionImage.

y
[float] Pixel coordinate of the seed location in the seed SectionImage.

section_data_set_ids
[list of integers] Target SectionDataSet IDs for image sync.

Returns

dict
The parsed json response

`get_image_to_image_2d`(*section_image_id*, *x*, *y*, *section_image_ids*)

For a list of target SectionImages, find the closest (x,y) location as defined by a seed SectionImage and seed (x,y) location.

Parameters

section_image_id
[integer] Seed for image sync.

x
[float] Pixel coordinate of the seed location in the seed SectionImage.

y
[float] Pixel coordinate of the seed location in the seed SectionImage.

section_image_ids
[list of ints] Target SectionImage IDs for image sync.

Returns

dict
The parsed json response

`get_image_to_reference`(*section_image_id*, *x*, *y*)

For a specified SectionImage and (x,y) location, return the (x,y,z) location in the ReferenceSpace of the associated SectionDataSet.

Parameters

section_image_id
[integer] Seed for image sync.

x
[float] Pixel coordinate on the specified SectionImage.

y
[float] Pixel coordinate on the specified SectionImage.

Returns

dict
The parsed json response

`get_reference_to_image`(*reference_space_id*, *x*, *y*, *z*, *section_data_set_ids*)

For a list of target SectionDataSets, find the closest SectionImage and (x,y) location as defined by a (x,y,z) location in a specified ReferenceSpace.

Parameters

reference_space_id
[integer] Seed for spatial sync.

x
[float] Coordinate (in microns) of the seed location in the seed ReferenceSpace.

y
[float] Coordinate (in microns) of the seed location in the seed ReferenceSpace.

z
[float] Coordinate (in microns) of the seed location in the seed ReferenceSpace.

section_data_set_ids
[list of ints] Target SectionDataSets IDs for image sync.

Returns

dict
The parsed json response

`get_structure_to_image`(*section_data_set_id*, *structure_ids*)

For a list of target structures, find the closest SectionImage and (x,y) location as defined by the centroid of each Structure.

Parameters

section_data_set_id
[integer] primary key

structure_ids
[list of integers] primary key

Returns

dict
The parsed json response

allensdk.api.queries.tree_search_api module

`class allensdk.api.queries.tree_search_api.TreeSearchApi`(*base_uri=None*)

Bases: *Api*

See Searching a Specimen or Structure Tree for additional documentation.

`get_tree`(*kind*, *db_id*, *ancestors=None*, *descendants=None*)

Fetch meta data for the specified structure or specimen.

Parameters

kind
[string] ‘Structure’ or ‘Specimen’

db_id
[integer] The id of the structure or specimen to search.

ancestors
[boolean, optional] whether to include ancestors in the response (defaults to False)

descendants
[boolean, optional] whether to include descendants in the response (defaults to False)

Returns

dict
parsed json response data

Module contents

allensdk.api.warehouse_cache package

Submodules

allensdk.api.warehouse_cache.cache module

`class allensdk.api.warehouse_cache.cache.Cache(manifest=None, cache=True, version=None, **kwargs)`

Bases: `object`

`add_manifest_paths(manifest_builder)`
Add cache-class specific paths to the manifest. In derived classes, should call super.

`build_manifest(file_name)`
Creation of default path specifications.

Parameters

file_name
[string] where to save it

`static cache_csv()`

`static cache_csv_dataframe()`

`static cache_csv_json()`

`static cache_json()`

`static cache_json_dataframe()`

`static cacher(fn, *args, **kwargs)`
make an rma query, save it and return the dataframe.

Parameters

fn
[function reference] makes the actual query using kwargs.

path
[string] where to save the data

strategy
[string or None, optional] 'create' always generates the data, 'file' loads from disk, 'lazy' queries the server if no file exists, None generates the data and bypasses all caching behavior

pre
[function] df|json->df|json, takes one data argument and returns filtered version, None for pass-through

post
[function] df|json->?, takes one data argument and returns Object

reader
[function, optional] path -> data, default NOP

writer
[function, optional] path, data -> None, default NOP

kwargs
[objects] passed through to the query function

Returns

Object or None
data type depends on fn, reader and/or post methods.

`static csv_writer`(*pth*, *gen*)

`get_cache_path`(*file_name*, *manifest_key*, **args*)

Helper method for accessing path specs from manifest keys.

Parameters

file_name
[string]

manifest_key
[string]

args
[ordered parameters]

Returns

string or None
path

`static json_remove_keys`(*data*, *keys*)

`static json_rename_columns`(*data*, *new_old_name_tuples=None*)

Convenience method to rename columns in a pandas dataframe.

Parameters

data
[dataframe] edited in place.

new_old_name_tuples
[list of string tuples (new, old)]

`load_csv`(*path*, *rename=None*, *index=None*)

Read a csv file as a pandas dataframe.

Parameters

rename
[list of string tuples (new old), optional] columns to rename

index
[string, optional] post-rename column to use as the row label.

`load_json`(*path*, *rename=None*, *index=None*)

Read a json file as a pandas dataframe.

Parameters

rename
[list of string tuples (new old), optional] columns to rename

index
[string, optional] post-rename column to use as the row label.

`load_manifest`(*file_name*, *version=None*)

Read a keyed collection of path specifications.

Parameters

file_name
[string] path to the manifest file

Returns

Manifest

`manifest_dataframe`()

Convenience method to view manifest as a pandas dataframe.

`static nocache_dataframe`()

`static nocache_json`()

`static pathfinder`(*file_name_position*, *secondary_file_name_position=None*, *path_keyword=None*)

helper method to find path argument in legacy methods written prior to the @cacheable decorator. Do not use for new @cacheable methods.

Parameters

file_name_position
[integer] zero indexed position in the decorated method args where file path may be found.

secondary_file_name_position
[integer] zero indexed position in the decorated method args where the file path may be found.

path_keyword
[string] kwarg that may have the file path.

Notes

This method is only intended to provide backward-compatibility for some methods that otherwise do not follow the path conventions of the @cacheable decorator.

static remove_keys(*data*, *keys=None*)

DataFrame version

static rename_columns(*data*, *new_old_name_tuples=None*)

Convenience method to rename columns in a pandas dataframe.

Parameters

data
[dataframe] edited in place.

new_old_name_tuples
[list of string tuples (new, old)]

wrap(*fn*, *path*, *cache*, *save_as_json=True*, *return_dataframe=False*, *index=None*, *rename=None*, ***kwargs*)

make an rma query, save it and return the dataframe.

Parameters

fn
[function reference] makes the actual query using kwargs.

path
[string] where to save the data

cache
[boolean] True will make the query, False just loads from disk

save_as_json
[boolean, optional] True (default) will save data as json, False as csv

return_dataframe
[boolean, optional] True will cast the return value to a pandas dataframe, False (default) will not

index
[string, optional] column to use as the pandas index

rename
[list of string tuples, optional] (new, old) columns to rename

kwargs
[objects] passed through to the query function

Returns

dict or DataFrame
data type depends on return_dataframe option.

Notes

Column renaming happens after the file is reloaded for json

allensdk.api.warehouse_cache.cache.**cacheable**(*strategy=None*, *pre=None*, *writer=None*, *reader=None*, *post=None*, *pathfinder=None*)

decorator for rma queries, save it and return the dataframe.

Parameters

fn
[function reference] makes the actual query using kwargs.

path
[string] where to save the data

strategy
[string or None, optional] 'create' always gets the data from the source (server or generated), 'file' loads from disk, 'lazy' creates the data and saves to file if no file exists, None queries the server and bypasses all caching behavior

pre
[function] df|json->df|json, takes one data argument and returns filtered version, None for pass-through

post
[function] df|json->?, takes one data argument and returns Object

reader
[function, optional] path -> data, default NOP

writer
[function, optional] path, data -> None, default NOP

kwargs
[objects] passed through to the query function

Returns

dict or DataFrame
data type depends on dataframe option.

Notes

Column renaming happens after the file is reloaded for json

allensdk.api.warehouse_cache.cache.**get_default_manifest_file**(*cache_name*)

allensdk.api.warehouse_cache.cache.**memoize**(*f*)

Creates an unbound cache of function calls and results. Note that arguments of different types are not cached separately (so f(3.0) and f(3) are not treated as distinct calls)

Arguments to the cached function must be hashable.

View the cache size with f.cache_size(). Clear the cache with f.cache_clear(). Access the underlying function with f.__wrapped__.

allensdk.api.warehouse_cache.caching_utilities module

allensdk.api.warehouse_cache.caching_utilities.**call_caching**(*fetch: Callable[[], Q]*, *write: Callable[[Q], None]*, *read: Callable[[], P]*, *pre_write: Callable[[Q], Q] | None = None*, *cleanup: Callable[[], None] | None = None*, *lazy: bool = True*, *num_tries: int = 1*, *failure_message: str = ''*) → P

allensdk.api.warehouse_cache.caching_utilities.**call_caching**(*fetch: Callable[[], Q]*, *write: Callable[[Q], None]*, *read: None = None*, *pre_write: Callable[[Q], Q] | None = None*, *cleanup: Callable[[], None] | None = None*, *lazy: bool = True*, *num_tries: int = 1*, *failure_message: str = ''*) → None

Access data, caching on a local store for future accesses.

Parameters

fetch
Function which pulls data from a remote/expensive source.

write
Function which stores data in a local/inexpensive store.

read
Function which pulls data from a local/inexpensive store.

pre_write
Function applied to obtained data after fetching, but before writing.

cleanup
Function for fixing a failed fetch. e.g. unlinking a partially downloaded file. Exceptions raised by cleanup are not themselves handled

lazy
If True, attempt to read the data from the local/inexpensive store before fetching it. If False, forcibly fetch from the remote/expensive store.

num_tries
How many fetches to attempt before (re)raising an exception. A fetch is failed if reading the result raises an exception.

failure_message
Provides additional context in the event of a failed download. Emitted when retrying, and when a fetch failure occurs after tries are exhausted

Returns

The result of calling read

allensdk.api.warehouse_cache.caching_utilities.**one_file_call_caching**(*path: Path | str*, *fetch: Callable[[], Q]*, *write: Callable[[Path | str, Q], None]*, *read: Callable[[Path | str], P] | None = None*, *pre_write: Callable[[Q], Q] | None = None*, *cleanup: Callable[[], None] | None = None*, *lazy: bool = True*, *num_tries: int = 1*, *failure_message: str = ''*) → P | None

A call_caching variant where the local store is a single file. See call_caching for complete documentation.

Parameters

path
Path at which the data will be stored

Module contents

Submodules

allensdk.api.api module

class allensdk.api.api.**Api**(*api_base_url_string=None*)

Bases: object

cleanup_truncated_file(*file_path*)

Helper for removing files.

Parameters

file_path
[string] Absolute path including the file name to remove.

construct_well_known_file_download_url(*well_known_file_id*)

Join data api endpoint and id.

Parameters

well_known_file_id
[integer or string representing an integer] well known file id

Returns

string
the well-known-file download url for the current api api server

See also:

retrieve_file_over_http
Can be used to retrieve the file from the url.

default_api_url = 'http://api.brain-map.org'

`do_query`(*url_builder_fn*, *json_traversal_fn*, **args*, ***kwargs*)

Bundle an query url construction function with a corresponding response json traversal function.

Parameters

url_builder_fn
[function] A function that takes parameters and returns an rma url.

json_traversal_fn
[function] A function that takes a json-parsed python data structure and returns data from it.

post
[boolean, optional kwarg] True does an HTTP POST, False (default) does a GET

args
[arguments] Arguments to be passed to the url builder function.

kwargs
[keyword arguments] Keyword arguments to be passed to the rma builder function.

Returns

any type
The data extracted from the json response.

Examples

A simple Api subclass example.

`do_rma_query`(*rma_builder_fn*, *json_traversal_fn*, **args*, ***kwargs*)

Bundle an RMA query url construction function with a corresponding response json traversal function.

..note:: Deprecated in AllenSDK 0.9.2
do_rma_query will be removed in AllenSDK 1.0, it is replaced by *do_query* because the latter is more general.

Parameters

rma_builder_fn
[function] A function that takes parameters and returns an rma url.

json_traversal_fn
[function] A function that takes a json-parsed python data structure and returns data from it.

args
[arguments] Arguments to be passed to the rma builder function.

kwargs
[keyword arguments] Keyword arguments to be passed to the rma builder function.

Returns

any type
The data extracted from the json response.

Examples

A simple Api subclass example.

`download_url = 'http://download.alleninstitute.org'`

`json_msg_query`(*url*, *dataframe=False*)

Common case where the url is fully constructed
and the response data is stored in the 'msg' field.

Parameters

url
[string] Where to get the data in json form

dataframe
[boolean] True converts to a pandas dataframe, False (default) doesn't

Returns

dict or DataFrame
returned data; type depends on dataframe option

`load_api_schema`()

Download the RMA schema from the current RMA endpoint

Returns

dict
the parsed json schema message

Notes

This information and other Allen Brain Atlas Data Portal Data Model documentation is also available as a Class Hierarchy and Class List.

`read_data`(*parsed_json*)

Return the message data from the parsed query.

Parameters

parsed_json
[dict] A python structure corresponding to the JSON data returned from the API.

Notes

See API Response Formats - Response Envelope for additional documentation.

`retrieve_file_over_http`(*url*, *file_path*, *zipped=False*)

Get a file from the data api and save it.

Parameters

url
[string] Url[R099781a1d33c-1]_ from which to get the file.

file_path
[string] Absolute path including the file name to save.

zipped
[bool, optional] If true, assume that the response is a zipped directory and attempt to extract contained files into the directory containing file_path. Default is False.

See also:

construct_well_known_file_download_url
Can be used to construct the url.

References

[1]

retrieve_parsed_json_over_http(*url*, *post=False*)

Get the document and put it in a Python data structure

Parameters

url
[string] Full API query url.

post
[boolean] True does an HTTP POST, False (default) encodes the URL and does a GET

Returns

dict
Result document as parsed by the JSON library.

retrieve_xml_over_http(*url*)

Get the document and put it in a Python data structure

Parameters

url
[string] Full API query url.

Returns

string
Unparsed xml string.

set_api_urls(*api_base_url_string*)

Set the internal RMA and well known file download endpoint urls based on a api server endpoint.

Parameters

api_base_url_string
[string] url of the api to point to

set_default_working_directory(*working_directory*)

Set the working directory where files will be saved.

Parameters

working_directory
[string] the absolute path string of the working directory.

allensdk.api.api.**stream_file_over_http**(*url*, *file_path*, *timeout=(9.05, 31.1)*)

Supply an http get request and stream the response to a file.

Parameters

url
[str] Send the request to this url

file_path
[str] Stream the response to this path

timeout
[float or tuple of float, optional] Specify a timeout for the request. If a tuple, specify seperate connect and read timeouts.

allensdk.api.api.**stream_zip_directory_over_http**(*url*, *directory*, *members=None*, *timeout=(9.05, 31.1)*)

Supply an http get request and stream the response to a file.

Parameters

url
[str] Send the request to this url

directory
[str] Extract the response to this directory

members
[list of str, optional] Extract only these files

timeout
[float or tuple of float, optional] Specify a timeout for the request. If a tuple, specify seperate connect and read timeouts.

Module contents

Subclasses of allensdk.api.api.Api to implement specific queries to the Allen Brain Atlas Data Portal.

6.1.2 allensdk.brain_observatory package

Subpackages

allensdk.brain_observatory.behavior package

Subpackages

allensdk.brain_observatory.behavior.behavior_project_cache package

Subpackages

allensdk.brain_observatory.behavior.behavior_project_cache.project_metadata_writer package

Submodules

allensdk.brain_observatory.behavior.behavior_project_cache.project_metadata_writer.behavior_project_metadata_ module

allensdk.brain_observatory.behavior.behavior_project_cache.project_metadata_writer.schemas module

Module contents

allensdk.brain_observatory.behavior.behavior_project_cache.tables package

Subpackages

allensdk.brain_observatory.behavior.behavior_project_cache.tables.util package

Submodules

allensdk.brain_observatory.behavior.behavior_project_cache.tables.util.experiments_table_utils module

allensdk.brain_observatory.behavior.behavior_project_cache.tables.util.image_presentation_utils module

allensdk.brain_observatory.behavior.behavior_project_cache.tables.util.prior_exposure_processing module

Module contents

Submodules

allensdk.brain_observatory.behavior.behavior_project_cache.tables.experiments_table module

allensdk.brain_observatory.behavior.behavior_project_cache.tables.metadata_table_schemas module

allensdk.brain_observatory.behavior.behavior_project_cache.tables.ophys_mixin module

allensdk.brain_observatory.behavior.behavior_project_cache.tables.ophys_sessions_table module

allensdk.brain_observatory.behavior.behavior_project_cache.tables.project_table module

allensdk.brain_observatory.behavior.behavior_project_cache.tables.sessions_table module

Module contents

Submodules

allensdk.brain_observatory.behavior.behavior_project_cache.behavior_neuropixels_project_cache module

allensdk.brain_observatory.behavior.behavior_project_cache.behavior_project_cache module

allensdk.brain_observatory.behavior.behavior_project_cache.project_cache_base module

allensdk.brain_observatory.behavior.behavior_project_cache.utils module

Module contents

allensdk.brain_observatory.behavior.data_files package

Submodules

allensdk.brain_observatory.behavior.data_files.avg_projection_file module

allensdk.brain_observatory.behavior.data_files.demix_file module

allensdk.brain_observatory.behavior.data_files.dff_file module

allensdk.brain_observatory.behavior.data_files.event_detection_file module

allensdk.brain_observatory.behavior.data_files.eye_tracking_file module

allensdk.brain_observatory.behavior.data_files.eye_tracking_metadata_file module

allensdk.brain_observatory.behavior.data_files.eye_tracking_video module

allensdk.brain_observatory.behavior.data_files.max_projection_file module

allensdk.brain_observatory.behavior.data_files.neuropil_corrected_file module

allensdk.brain_observatory.behavior.data_files.neuropil_file module

allensdk.brain_observatory.behavior.data_files.rigid_motion_transform_file module

allensdk.brain_observatory.behavior.data_files.stimulus_file module

allensdk.brain_observatory.behavior.data_files.sync_file module

Module contents

allensdk.brain_observatory.behavior.data_objects package

Subpackages

allensdk.brain_observatory.behavior.data_objects.cell_specimens package

Subpackages

allensdk.brain_observatory.behavior.data_objects.cell_specimens.traces package

Submodules

allensdk.brain_observatory.behavior.data_objects.cell_specimens.traces.corrected_fluorescence_traces module

allensdk.brain_observatory.behavior.data_objects.cell_specimens.traces.demixed_traces module

allensdk.brain_observatory.behavior.data_objects.cell_specimens.traces.dff_traces module

allensdk.brain_observatory.behavior.data_objects.cell_specimens.traces.neuropil_traces module

Module contents

Submodules

allensdk.brain_observatory.behavior.data_objects.cell_specimens.cell_specimens module

allensdk.brain_observatory.behavior.data_objects.cell_specimens.events module

allensdk.brain_observatory.behavior.data_objects.cell_specimens.rois_mixin module

Module contents

allensdk.brain_observatory.behavior.data_objects.eye_tracking package

Submodules

allensdk.brain_observatory.behavior.data_objects.eye_tracking.eye_tracking_table module

allensdk.brain_observatory.behavior.data_objects.eye_tracking.rig_geometry module

Module contents

allensdk.brain_observatory.behavior.data_objects.metadata package

Subpackages

allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata package

Submodules

allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.behavior_metadata module

allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.behavior_session_id module

allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.behavior_session_uuid module

allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.date_of_acquisition module

allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.equipment module

class allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.equipment.**Equipment**(*eq*
st

Bases: DataObject, JsonReadableInterface, LimsReadableInterface, NwbReadableInterface, NwbWritableInterface

the name of the experimental rig.

classmethod from_json(*dict_repr: dict*) → *Equipment*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*behavior_session_id: int*, *lims_db:* PostgresQueryMixin) → *Equipment*

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_nwb(*nwbfile: NWBFile*) → *Equipment*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

to_nwb(*nwbfile: NWBFile*) → NWBFile

Given an already populated DataObject, return an pyNWB file object that had had DataObject data added.

Parameters

nwbfile
[NWBFile] An NWB file object

Returns

NWBFile
An NWB file object that has had data from the DataObject added to it.

property type

class allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.equipment.**EquipmentTyp**

Bases: Enum

An enumeration.

MESOSCOPE = 'MESOSCOPE'

OTHER = 'OTHER'

allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.foraging_id module

class allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.foraging_id.**ForagingI**

Bases: DataObject, LimsReadableInterface, JsonReadableInterface

Foraging id

classmethod from_json(*dict_repr: dict*) → *ForagingId*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*behavior_session_id: int*, *lims_db:* PostgresQueryMixin) → *ForagingId*

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.project_code module

class allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.project_code.**ProjectC**

Bases: DataObject, LimsReadableInterface, NwbReadableInterface

Unique identifier for the project this BehaviorSession is associated with. Project ids can be used internally to extract a project code.

If project_Code is null/None, we set the value to string 'Not Available'.

classmethod from_lims(*behavior_session_id: int*, *lims_db:* PostgresQueryMixin) → *ProjectCode*

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_nwb(*nwbfile: NWBFile*) → *ProjectCode*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.session_type module

allensdk.brain_observatory.behavior.data_objects.metadata.behavior_metadata.stimulus_frame_rate module

Module contents

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata package

Subpackages

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.multi_plane_metadata package

Submodules

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.multi_plane_metadata.im module

class allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.multi_plane_m

Bases: `DataObject`, `LimsReadableInterface`, `JsonReadableInterface`, `NwbReadableInterface`

classmethod from_json(*dict_repr: dict*) → *ImagingPlaneGroup*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*ophys_experiment_id: int*, *lims_db:* PostgresQueryMixin) → *ImagingPlaneGroup* | None

Parameters

ophys_experiment_id
lims_db

Returns

ImagingPlaneGroup instance if ophys_experiment given by
ophys_experiment_id is part of a plane group

else None

`classmethod from_nwb(nwbfile: NWBFile) → ImagingPlaneGroup`

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

`property plane_group`

`property plane_group_count`

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.multi_plane_metadata.m module

`class allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.multi_plane_m`

Bases: *OphysExperimentMetadata*

classmethod from_json(*dict_repr: dict*) → *MultiplaneMetadata*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*ophys_experiment_id: int*, *lims_db:* PostgresQueryMixin) → *MultiplaneMetadata*

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_nwb(*nwbfile: NWBFile*) → *MultiplaneMetadata*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

property imaging_plane_group: int

property imaging_plane_group_count: int

Module contents

Submodules

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.field_of_view_shape module

class allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.field_of_view_

Bases: DataObject, LimsReadableInterface, NwbReadableInterface, JsonReadableInterface

classmethod from_json(*dict_repr: dict*) → *FieldOfViewShape*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*ophys_experiment_id: int*, *lims_db:* PostgresQueryMixin) → *FieldOfViewShape*

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_nwb(*nwbfile: NWBFile*) → *FieldOfViewShape*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

property height

property width

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.imaging_depth module

class allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.imaging_depth

Bases: DataObject, LimsReadableInterface, NwbReadableInterface, JsonReadableInterface

Data object loads and stores the imaging_depth (microns) for an experiments. This is the calculated difference between measured z-depths of the surface and imaging_depth.

classmethod from_json(*dict_repr: dict*) → *ImagingDepth*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*ophys_experiment_id: int*, *lims_db:* PostgresQueryMixin) → *ImagingDepth*

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_nwb(*nwbfile: NWBFile*) → *ImagingDepth*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.imaging_plane module

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.ophys_container_id module

class allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.ophys_contain

Bases: DataObject, LimsReadableInterface, JsonReadableInterface, NwbReadableInterface

"experiment container id

classmethod from_json(*dict_repr: dict*) → ExperimentContainerId

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*ophys_experiment_id: int*, *lims_db:* PostgresQueryMixin) → ExperimentContainerId

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_nwb(*nwbfile: NWBFile*) → ExperimentContainerId

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.ophys_experiment_meta module

class allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.ophys_experime

Bases: DataObject, LimsReadableInterface, JsonReadableInterface, NwbReadableInterface

Container class for ophys experiment metadata

property field_of_view_shape: *FieldOfViewShape*

classmethod from_json(*dict_repr: dict*) → *OphysExperimentMetadata*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*ophys_experiment_id: int*, *lims_db:* PostgresQueryMixin) → *OphysExperimentMetadata*

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_nwb(*nwbfile: NWBFile*) → *OphysExperimentMetadata*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

property imaging_depth: int

property ophys_container_id: int

property ophys_experiment_id: int

property ophys_session_id: int

property project_code: str

property targeted_imaging_depth: int

update_targeted_imaging_depth(*ophys_experiment_ids: List[int]*, *lims_db:* PostgresQueryMixin)

Update the value for targeted imaging depth given a set of experiments to be published.

Compute the targeted_imaging_depth (average over experiments in a container) only for those experiments input.

Parameters

ophys_experiment_ids
[list of ints] Set of experiments to calculate targeted_imaging_depth for. Needs to contain the experiment this metadata represents.

lims_db
[PostgresQueryMixin] Connection to the LIMS2 database.

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.ophys_project_code module

class allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.ophys_project_

Bases: *ProjectCode*

Unique identifier for the project this OphysExperiment is associated with. Project ids can be used internally to extract a project code.

If the returned project id is null/None, return "Not available"

classmethod from_lims(*ophys_experiment_id: int*, *lims_db:* PostgresQueryMixin) → *OphysProjectCode*

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.ophys_session_id module

class allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.ophys_session_

Bases: DataObject, LimsReadableInterface, JsonReadableInterface, NwbReadableInterface

"Ophys session id

classmethod from_json(*dict_repr: dict*) → *OphysSessionId*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*ophys_experiment_id: int*, *lims_db:* PostgresQueryMixin) → *OphysSessionId*

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_nwb(*nwbfile: NWBFile*) → *OphysSessionId*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.targeted_imaging_depth module

class allensdk.brain_observatory.behavior.data_objects.metadata.ophys_experiment_metadata.targeted_imag

Bases: DataObject, LimsReadableInterface, NwbReadableInterface, JsonReadableInterface

Data object loads and stores the average `imaging_depth`s (microns) across experiments in the container that an experiment is associated with.

classmethod from_json(*dict_repr: dict*) → *TargetedImagingDepth*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*ophys_experiment_id: int*, *lims_db:* PostgresQueryMixin, *ophys_experiment_ids: List[int] | None = None*) → *TargetedImagingDepth*

Load targeted imaging depth.

Parameters

ophys_experiment_id
[int] Id of experiment to calculate targeted depth for.

lims_db
[PostgresQueryMixin] Connection to the LIMS2 database.

ophys_experiment_ids
[list of int] Subset of experiments in the container of `ophys_experiment_id` to calculate the target_imaging_depth. List should contain the value of `ophys_experiment_id`.

classmethod from_nwb(*nwbfile: NWBFile*) → *TargetedImagingDepth*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

Module contents

allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata package

Submodules

allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.age module

allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.driver_line module

class allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.driver_line.**DriverLine**

Bases: `DataObject`, `LimsReadableInterface`, `JsonReadableInterface`, `NwbReadableInterface`

the genotype name(s) of the driver line(s)

classmethod from_json(*dict_repr: dict*) → *DriverLine*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*behavior_session_id: int*, *lims_db:* PostgresQueryMixin, *allow_none: bool* = *True*) → *DriverLine*

Parameters

behavior_session_id: int
lims_db: PostgresQueryMixin
allow_none: bool
if True, allow None as a valid result

Returns

An instance of DriverLine with the value resulting from a query to the LIMS database

`classmethod from_nwb(nwbfile: NWBFile) → DriverLine`

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.full_genotype module

`class allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.full_genotype.FullGeno`

Bases: `DataObject`, `LimsReadableInterface`, `JsonReadableInterface`, `NwbReadableInterface`

the name of the subject's genotype

`classmethod from_json(dict_repr: dict) → FullGenotype`

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

`classmethod from_lims(behavior_session_id: int, lims_db: PostgresQueryMixin) → FullGenotype`

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

`classmethod from_nwb(nwbfile: NWBFile) → FullGenotype`

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

parse_cre_line(*warn=False*) → str | None

Parameters

warn
Whether to output warning if parsing fails

Returns

cre_line
just the Cre line, e.g. Vip-IRES-Cre, or None if not possible to parse

allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.mouse_id module

class allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.mouse_id.**MouseId**(*mouse_*
str)

Bases: DataObject, LimsReadableInterface, JsonReadableInterface, NwbReadableInterface

the LabTracks ID

classmethod from_json(*dict_repr: dict*) → *MouseId*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*behavior_session_id: int*, *lims_db:* PostgresQueryMixin) → *MouseId*

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_nwb(*nwbfile: NWBFile*) → *MouseId*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.reporter_line module

class allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.reporter_line.**ReporterL**

Bases: DataObject, LimsReadableInterface, JsonReadableInterface, NwbReadableInterface

the genotype name(s) of the reporter line(s)

classmethod from_json(*dict_repr: dict*) → *ReporterLine*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*behavior_session_id: int*, *lims_db:* PostgresQueryMixin, *allow_none: bool = True*) → *ReporterLine*

Parameters

behavior_session_id: int
lims_db: PostgresQueryMixin
allow_none: bool
if True, allow None as a valid result

Returns

An instance of ReporterLine with the value resulting from a query to the LIMS database

classmethod from_nwb(*nwbfile: NWBFile*) → *ReporterLine*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

static parse(*reporter_line: List[str] | None | str*, *warn=False*) → str | None

There can be multiple reporter lines, so it is returned from LIMS as a list. But there shouldn't be more than 1 for behavior. This tries to convert to str

Parameters

reporter_line
List of reporter line

warn
Whether to output warnings if parsing fails

Returns

single reporter line, or None if not possible

parse_indicator(*warn=False*) → str | None

Parses indicator from reporter

allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.sex module

class allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.sex.**Sex**(*sex: str*)

Bases: DataObject, LimsReadableInterface, JsonReadableInterface, NwbReadableInterface

sex of the animal (M/F)

classmethod from_json(*dict_repr: dict*) → *Sex*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_lims(*behavior_session_id: int*, *lims_db:* PostgresQueryMixin) → *Sex*

Populate a DataObject from an internal database (likely LIMS)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_nwb(*nwbfile: NWBFile*) → *Sex*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

allensdk.brain_observatory.behavior.data_objects.metadata.subject_metadata.subject_metadata module

Module contents

Submodules

allensdk.brain_observatory.behavior.data_objects.metadata.behavior_ophys_metadata module

Module contents

allensdk.brain_observatory.behavior.data_objects.running_speed package

Submodules

allensdk.brain_observatory.behavior.data_objects.timestamps.ophys_timestamps module

allensdk.brain_observatory.behavior.data_objects.timestamps.util module

Module contents

allensdk.brain_observatory.behavior.data_objects.trials package

Submodules

allensdk.brain_observatory.behavior.data_objects.trials.trial module

allensdk.brain_observatory.behavior.data_objects.trials.trials module

Module contents

Submodules

allensdk.brain_observatory.behavior.data_objects.licks module

allensdk.brain_observatory.behavior.data_objects.motion_correction module

allensdk.brain_observatory.behavior.data_objects.projections module

allensdk.brain_observatory.behavior.data_objects.rewards module

allensdk.brain_observatory.behavior.data_objects.task_parameters module

Module contents

allensdk.brain_observatory.behavior.sync package

Submodules

allensdk.brain_observatory.behavior.sync.process_sync module

allensdk.brain_observatory.behavior.sync.process_sync.**calculate_delay**(*sync_data*, *stim_vsync_fall*, *sample_frequency*)

allensdk.brain_observatory.behavior.sync.process_sync.**filter_digital**(*rising*, *falling*, *threshold=0.0001*)

Removes short transients from digital signal.

Rising and falling should be same length and units
in seconds.

Kwargs:
threshold (float): transient width

Module contents

Created on Sunday July 15 2018

@author: marinag

allensdk.brain_observatory.behavior.sync.**get_behavior_monitoring**(*dataset: Dataset*, *permissive: bool = False*) → ndarray | None

Report the timestamps of each frame of the behavior monitoring video

Parameters

dataset
[describes experiment timing]

permissive
[If True, None will be returned if timestamps are not found.] If False, a KeyError will be raised

Returns

array of timestamps (floating point; seconds; relative to experiment start) or None. If None, no behavior monitoring timestamps were found in this sync dataset.

allensdk.brain_observatory.behavior.sync.**get_eye_tracking**(*dataset: Dataset*, *permissive: bool = False*) → ndarray | None

Report the timestamps of each frame of the eye tracking video

Parameters

dataset
[describes experiment timing]

permissive
[If True, None will be returned if timestamps are not found.] If False, a KeyError will be raised

Returns

array of timestamps (floating point; seconds; relative to experiment start) or None. If None, no eye tracking timestamps were found in this sync dataset.

allensdk.brain_observatory.behavior.sync.**get_lick_times**(*dataset: Dataset*, *permissive: bool = False*) → ndarray | None

Report the timestamps of each detected lick

Parameters

dataset
[describes experiment timing]

permissive
[If True, None will be returned if timestamps are not found.] If False, a KeyError will be raised

Returns

array of timestamps (floating point; seconds; relative to experiment start)
or None. If None, no lick timestamps were found in this sync dataset.

allensdk.brain_observatory.behavior.sync.**get_ophys_frames**(*dataset: Dataset*, *permissive: bool = False*) → ndarray

Report the timestamps of each optical physiology video frame

Parameters

dataset
[describes experiment timing]

permissive
[If True, None will be returned if timestamps are not found.] If False, a KeyError will be raised

Returns

array of timestamps (floating point; seconds; relative to experiment start).

Notes

use rising edge for Scientifica, falling edge for Nikon http://confluence.corp.alleninstitute.org/display/IT/Ophys+Time+Sync This function uses rising edges

allensdk.brain_observatory.behavior.sync.**get_raw_stimulus_frames**(*dataset: Dataset*, *permissive: bool = False*) → ndarray

Report the raw timestamps of each stimulus frame. This corresponds to the time at which the psychopy window's flip method returned, but not necessarily to the time at which the stimulus frame was displayed.

Parameters

dataset
[describes experiment timing]

permissive
[If True, None will be returned if timestamps are not found.] If False, a KeyError will be raised

Returns

array of timestamps (floating point; seconds; relative to experiment start).

allensdk.brain_observatory.behavior.sync.**get_stim_photodiode**(*dataset: Dataset*, *permissive: bool = False*) → List[float] | None

Report the timestamps of each detected sync square transition (both black -> white and white -> black) in this experiment.

Parameters

dataset
[describes experiment timing]

permissive
[If True, None will be returned if timestamps are not found.] If False, a KeyError will be raised

Returns

array of timestamps (floating point; seconds; relative to experiment start) or None. If None, no photodiode timestamps were found in this sync dataset.

allensdk.brain_observatory.behavior.sync.**get_sync_data**(*sync_path: str*, *permissive: bool = False*) → Dict[str, List | ndarray | None]

Convenience function for extracting several timestamp arrays from a sync file.

Parameters

sync_path
[The hdf5 file here ought to be a Visual Behavior sync output]

file. See allensdk.brain_observatory.sync_dataset for more details of this format.
permissive
[If True, None will be returned if timestamps are not found.] If False, a KeyError will be raised

Returns

A dictionary with the following keys. All timestamps in seconds:
ophys_frames : timestamps of each optical physiology frame lick_times : timestamps of each detected lick ophys_trigger : The time at which ophys acquisition was started eye_tracking : timestamps of each eye tracking video frame behavior_monitoring : timestamps of behavior monitoring video frame stim_photodiode : timestamps of each photodiode transition stimulus_times_no_delay : raw stimulus frame timestamps

Some values may be None. This indicates that the corresponding timestamps were not located in this sync file.

allensdk.brain_observatory.behavior.sync.**get_trigger**(*dataset:* Dataset, *permissive: bool = False*) → ndarray | None

Returns (as a 1-element array) the time at which optical physiology
acquisition was started.

Parameters

dataset
[describes experiment timing]

permissive
[If True, None will be returned if timestamps are not found.] If False, a KeyError will be raised

Returns

timestamps (floating point; seconds; relative to experiment start) or None. If None, no timestamps were found in this sync dataset.

Notes

Ophys frame timestamps can be recorded before acquisition start when experimenters are setting up the recording session. These do not correspond to acquired ophys frames.

allensdk.brain_observatory.behavior.utils package

Submodules

allensdk.brain_observatory.behavior.utils.metadata_parsers module

allensdk.brain_observatory.behavior.utils.metadata_parsers.**parse_behavior_context**(*session_type: str*) → str

Return if session is passive or active.

Parameters

session_type
[str] String session type name.

Returns

behavior_type
[str] Return string describing active or passive session.

allensdk.brain_observatory.behavior.utils.metadata_parsers.**parse_num_cortical_structures**(*project_code: str*) → str | None

Return the number of structures that were targeted for this session.

Parameters

project_code
[str] Full project name of the experiment.

Returns

num_structures
[int] Number of structures targeted for the session.

allensdk.brain_observatory.behavior.utils.metadata_parsers.**parse_num_depths**(*project_code: str*) → str | None

Return the number of depths that were imaged for the session.

Parameters

project_code
[str] Full project name of the experiment.

Returns

num_depths
[int] Number of depths imaged for the session.

allensdk.brain_observatory.behavior.utils.metadata_parsers.**parse_stimulus_set**(*session_type: str*) → str

Return the name of the image set or gratings.

Parameters

session_type
[str] String session type name.

Returns

stim_set
[str] Name of stimulus set shown for session.

Module contents

allensdk.brain_observatory.behavior.write_nwb package

Subpackages

allensdk.brain_observatory.behavior.write_nwb.behavior package

Submodules

allensdk.brain_observatory.behavior.write_nwb.behavior.schemas module

Module contents

allensdk.brain_observatory.behavior.write_nwb.extensions package

Subpackages

allensdk.brain_observatory.behavior.write_nwb.extensions.event_detection package

Submodules

allensdk.brain_observatory.behavior.write_nwb.extensions.event_detection.extension_builder module

allensdk.brain_observatory.behavior.write_nwb.extensions.event_detection.extension_builder.**main**()

allensdk.brain_observatory.behavior.write_nwb.extensions.event_detection.ndx_ophys_events module

Module contents

allensdk.brain_observatory.behavior.write_nwb.extensions.stimulus_template package

Submodules

allensdk.brain_observatory.behavior.write_nwb.extensions.stimulus_template.extension_builder module

allensdk.brain_observatory.behavior.write_nwb.extensions.stimulus_template.extension_builder.**main**()

allensdk.brain_observatory.behavior.write_nwb.extensions.stimulus_template.ndx_stimulus_template module

Module contents

Module contents

allensdk.brain_observatory.behavior.write_nwb.ophys package

Submodules

allensdk.brain_observatory.behavior.write_nwb.ophys.schemas module

Module contents

Submodules

allensdk.brain_observatory.behavior.write_nwb.nwb_writer_utils module

Module contents

Submodules

allensdk.brain_observatory.behavior.behavior_ophys_analysis module

allensdk.brain_observatory.behavior.behavior_ophys_experiment module

allensdk.brain_observatory.behavior.behavior_ophys_session module

allensdk.brain_observatory.behavior.behavior_session module

allensdk.brain_observatory.behavior.criteria module

Functions for calculating mtrain state transitions. If criteria are met, return true. Otherwise, return false.

allensdk.brain_observatory.behavior.criteria.**consistency_is_key**(*session_summary*)

need some way to judge consistency of various parameters

- dprime
- num trials
- hit rate
- fa rate
- lick timing

allensdk.brain_observatory.behavior.criteria.**consistent_behavior_within_session**(*session_summary*)

need some way to measure consistent performance within a session

- compare peak to overall dprime?
- variance in rolling window dprime?

allensdk.brain_observatory.behavior.criteria.**meets_engagement_criteria**(*session_summary*)

Returns true if engagement criteria were met for the past 3 days, else false. Args:

session_summary (pd.DataFrame): Pandas dataframe with daily values for 'dprime_peak' and 'num_engaged_trials', ordered ascending by training day, for at least 3 days. If dataframe is not properly ordered, criterion may not be correctly calculated. This function does not sort the data to preserve prior behavior (sorting column was not required by mtrain function) The mtrain implementation created the required columns if they didn't exist, so a more informative error is raised here to assist end-users in debugging.

Returns:

bool: True if criterion is met, False otherwise

allensdk.brain_observatory.behavior.criteria.**mostly_useful**(*trials*)

Returns True if fewer than half the trial time on the last day were aborted trials.

Args: trials (pd.DataFrame): Pandas dataframe with columns 'training_day', 'trial_type', and 'trial_length'.

Returns:

bool: True if criterion is met, False otherwise

allensdk.brain_observatory.behavior.criteria.**n_complete**(*threshold*, *count*)

For compatibility with original API. If count >= threshold, return True. Otherwise return False. Args:

threshold (numeric): Threshold for the count to meet. count (numeric): The count to compare to the threshold.

Returns:

True if count >= threshold, otherwise False.

allensdk.brain_observatory.behavior.criteria.**no_response_bias**(*session_summary*)

the mouse meets this criterion if their last session exhibited a response bias between 10% and 90%

Args: session_summary (pd.DataFrame): Pandas dataframe with daily values for 'response_bias', ordered ascending by training day, for at least 1 day. If dataframe is not properly ordered, criterion may not be correctly calculated. This function does not sort the data to preserve prior behavior (sorting column was not required by mtrain function). The mtrain implementation created the required columns if they didn't exist, so a more informative error is raised here to assist end-users in debugging.

Returns:

bool: True if criterion is met, False otherwise

allensdk.brain_observatory.behavior.criteria.**summer_over**(*trials*)

Returns true if the maximum value of 'training_day' in the trials dataframe is >= 40, else false.

allensdk.brain_observatory.behavior.criteria.**two_out_of_three_aint_bad**(*session_summary*)

Returns true if 2 of the last 3 days showed a peak d-prime above 2.

Args:

session_summary (pd.DataFrame): Pandas dataframe with daily values for 'dprime_peak', ordered ascending by training day, for at least the past 3 days. If dataframe is not properly ordered, criterion may not be correctly calculated. This function does not sort the data to preserve prior behavior (sorting column was not required by mtrain function). The mtrain implementation created the required columns if they didn't exist, so a more informative error is raised here to assist end-users in debugging.

Returns:

bool: True if criterion is met, False otherwise

allensdk.brain_observatory.behavior.criteria.**whole_lotta_trials**(*session_summary*)

Mouse meets this criterion if the last session has more than 300 trials. Args:

session_summary (pd.DataFrame): Pandas dataframe with daily values for 'num_contingent_trials', ordered ascending by training day, for at least 1 day. If dataframe is not properly ordered, criterion may not be correctly calculated. This function does not sort the data to preserve prior behavior (sorting column was not required by mtrain function). The mtrain implementation created the required columns if they didn't exist, so a more informative error is raised here to assist end-users in debugging.

Returns:

bool: True if criterion is met, False otherwise

allensdk.brain_observatory.behavior.criteria.**yesterday_was_good**(*session_summary*)

Returns true if the last day showed a peak d-prime above 2 Args:

session_summary (pd.DataFrame): Pandas dataframe with daily values for 'dprime_peak', ordered ascending by training day, for at least 1 day. If dataframe is not properly ordered, criterion may not be correctly calculated. This function does not sort the data to preserve prior behavior (sorting column was not required by mtrain function). The mtrain implementation created the required columns if they didn't exist, so a more informative error is raised here to assist end-users in debugging.

Returns:

bool: True if criterion is met, False otherwise

allensdk.brain_observatory.behavior.dprime module

allensdk.brain_observatory.behavior.dprime.**get_catch_responses**(*correct_reject=None*, *false_alarm=None*, *aborted=None*)

allensdk.brain_observatory.behavior.dprime.**get_dprime**(*hit_rate*, *fa_rate*, *sliding_window=100*)

calculates the d-prime for a given hit rate and false alarm rate https://en.wikipedia.org/wiki/Sensitivity_index Parameters ———- hit_rate : float

rate of hits in the True class

fa_rate

[float] rate of false alarms in the False class

limits

[tuple, optional] limits on extreme values, which distort. default: (0.01,0.99)

Returns

d_prime

allensdk.brain_observatory.behavior.dprime.**get_false_alarm_rate**(*correct_reject=None*, *false_alarm=None*, *aborted=None*, *sliding_window=100*)

allensdk.brain_observatory.behavior.dprime.**get_go_responses**(*hit=None*, *miss=None*, *aborted=None*)

allensdk.brain_observatory.behavior.dprime.**get_hit_rate**(*hit=None*, *miss=None*, *aborted=None*, *sliding_window=100*)

allensdk.brain_observatory.behavior.dprime.**get_rolling_dprime**(*rolling_hit_rate*, *rolling_fa_rate*, *sliding_window=100*)

allensdk.brain_observatory.behavior.dprime.**get_trial_count_corrected_false_alarm_rate**(*correct_reject=None*, *false_alarm=None*, *aborted=None*, *sliding_window=100*)

allensdk.brain_observatory.behavior.dprime.**get_trial_count_corrected_hit_rate**(*hit=None*, *miss=None*, *aborted=None*, *sliding_window=100*)

allensdk.brain_observatory.behavior.dprime.**trial_number_limit**(*p*, *N*)

allensdk.brain_observatory.behavior.event_detection module

allensdk.brain_observatory.behavior.event_detection.**filter_events_array**(*arr: ndarray*, *scale: float = 2*, *n_time_steps: int = 20*) → ndarray

Convolve the trace array with a 1d causal half-gaussian filter to smooth it for visualization

Uses a halfnorm distribution as weights to the filter

Modified from initial implementation by Nick Ponvert

Parameters

arr: np.ndarray
Trace matrix of dimension n traces x n frames

scale: float
std deviation of halfnorm distribution in units of timesteps

n_time_steps: int
number of time steps to use for the convolution operation

Returns

np.ndarray:
Output of the convolution operation

allensdk.brain_observatory.behavior.eye_tracking_processing module

exception allensdk.brain_observatory.behavior.eye_tracking_processing.**EyeTrackingError**

Bases: Exception

allensdk.brain_observatory.behavior.eye_tracking_processing.**compute_circular_area**(*df_row: Series*) → float

Calculate the area of the pupil as a circle using the max of the height/width as radius.

Note: This calculation assumes that the pupil is a perfect circle and any eccentricity is a result of the angle at which the pupil is being viewed.

Parameters

df_row
[pd.Series] A row from an eye tracking dataframe containing only "pupil_width" and "pupil_height".

Returns

float
The circular area of the pupil in pixels^2.

allensdk.brain_observatory.behavior.eye_tracking_processing.**compute_elliptical_area**(*df_row: Series*) → float

Calculate the area of corneal reflection (cr) or eye ellipse fits using the ellipse formula.

Parameters

df_row
[pd.Series] A row from an eye tracking dataframe containing either: "cr_width", "cr_height" or "eye_width", "eye_height"

Returns

float
The elliptical area of the eye or cr in pixels^2

allensdk.brain_observatory.behavior.eye_tracking_processing.**determine_likely_blinks**(*eye_areas: Series*, *pupil_areas: Series*, *outliers: Series*, *dilation_frames: int = 2*) → Series

Determine eye tracking frames which contain likely blinks or outliers

Parameters

eye_areas
[pd.Series] A pandas series of eye areas.

pupil_areas
[pd.Series] A pandas series of pupil areas.

outliers
[pd.Series] A pandas series containing bool values of outlier rows.

dilation_frames
[int, optional] Determines the number of additional adjacent frames to mark as 'likely_blink', by default 2.

Returns

pd.Series
A pandas series of bool values that has the same length as the number of eye tracking dataframe rows (frames).

allensdk.brain_observatory.behavior.eye_tracking_processing.**determine_outliers**(*data_df: DataFrame*, *z_threshold: float*) → Series

Given a dataframe and some z-score threshold return a pandas boolean Series where each entry indicates whether a given row contains at least one outlier (where outliers are calculated along columns).

Parameters

data_df
[pd.DataFrame] A dataframe containing only columns where outlier detection is desired. (e.g. "cr_area", "eye_area", "pupil_area")

z_threshold
[float] z-score values higher than the z_threshold will be considered outliers.

Returns

pd.Series
A pandas boolean Series whose length == len(data_df.index). True denotes that a row in the data_df contains at least one outlier.

allensdk.brain_observatory.behavior.eye_tracking_processing.**filter_on_blinks**(*eye_tracking_data: DataFrame*)

Set data is specified columns where likely_blink is true to NaN.

Modify the DataFrame in place.

Parameters

eye_tracking_data
[pandas.DataFrame] Data frame containing eye tracking data.

allensdk.brain_observatory.behavior.eye_tracking_processing.**load_eye_tracking_hdf**(*eye_tracking_file: Path*) → DataFrame

Load a DeepLabCut hdf5 file containing eye tracking data into a dataframe.

Note: The eye tracking hdf5 file contains 3 separate dataframes. One for corneal reflection (cr), eye, and pupil ellipse fits. This function loads and returns this data as a single dataframe.

Parameters

eye_tracking_file
[Path] Path to an hdf5 file produced by the DeepLabCut eye tracking pipeline. The hdf5 file will contain the following keys: “cr”, “eye”, “pupil”. Each key has an associated dataframe with the following columns: “center_x”, “center_y”, “height”, “width”, “phi”.

Returns

pd.DataFrame
A dataframe containing combined corneal reflection (cr), eyelid (eye), and pupil data. Column names for each field will be renamed by prepending the field name. (e.g. center_x -> eye_center_x)

allensdk.brain_observatory.behavior.eye_tracking_processing.**process_eye_tracking_data**(*eye_data: DataFrame*, *frame_times: Series*, *z_threshold: float = 3.0*, *dilation_frames: int = 2*) → DataFrame

Processes and refines raw eye tracking data by adding additional computed feature columns.

Parameters

eye_data
[pd.DataFrame] A ‘raw’ eye tracking dataframe produced by load_eye_tracking_hdf()

frame_times
[pd.Series] A series of frame times acquired from a behavior + ophy session ‘sync file’.

z_threshold
[float] z-score values higher than the z_threshold will be considered outliers, by default 3.0.

dilation_frames
[int, optional] Determines the number of additional adjacent frames to mark as ‘likely_blink’, by default 2.

Returns

pd.DataFrame
A refined eye tracking dataframe that contains additional information about frame times, eye areas, pupil areas, and frames with likely blinks/outliers.

Raises

EyeTrackingError
If the number of sync file frame times does not match the number of eye tracking frames.

allensdk.brain_observatory.behavior.image_api module

class allensdk.brain_observatory.behavior.image_api.**Image**(*data: ndarray*, *spacing: tuple*, *unit: str = 'mm'*)

Bases: tuple

Describes a 2D Image

data
[np.ndarray] Image data points

spacing
[tuple] Spacing describes the physical size of each pixel

unit
[str] Physical unit of the spacing (currently constrained to be isotropic)

data: ndarray
Alias for field number 0

spacing: tuple
Alias for field number 1

unit: str
Alias for field number 2

class allensdk.brain_observatory.behavior.image_api.**ImageApi**

Bases: object

static deserialize(*img*)

static serialize(*data*, *spacing*, *unit*)

allensdk.brain_observatory.behavior.mtrain module

class allensdk.brain_observatory.behavior.mtrain.**ExtendedTrialSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: Schema

This schema describes the edf core trial structure

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.behavior.mtrain.**FriendlyDate**(*format: str | None = None*, ***kwargs*)

Bases: Date

class allensdk.brain_observatory.behavior.mtrain.**FriendlyDateTime**(*format: str | None = None*, ***kwargs*)

Bases: DateTime

allensdk.brain_observatory.behavior.mtrain.**annotate_change_detect**(*trials*)

adds *change* and *detect* columns to dataframe

Parameters

trials
[pandas DataFrame] dataframe of trials

inplace
[bool, optional] modify *trials* in place. if False, returns a copy. default: True

See also:

`io.load_trials`

allensdk.brain_observatory.behavior.mtrain.**annotate_trials**(*trials*)

performs multiple annotatations:

- annotate_change_detect
- fix_change_time
- explode_response_window

Parameters

trials
[pandas DataFrame] dataframe of trials

inplace
[bool, optional] modify *trials* in place. if False, returns a copy. default: True

See also:

`io.load_trials`

allensdk.brain_observatory.behavior.mtrain.**assign_session_id**(*trials*)

adds a column with a unique ID for the session defined as
a combination of the mouse ID and startdatetime

Parameters

trials
[pandas DataFrame] dataframe of trials

inplace
[bool, optional] modify *trials* in place. if False, returns a copy. default: True

See also:

`io.load_trials`

allensdk.brain_observatory.behavior.mtrain.**explode_response_window**(*trials*)

explodes the *response_window* column in lower & upper columns

Parameters

trials
[pandas DataFrame] dataframe of trials

inplace
[bool, optional] modify *trials* in place. if False, returns a copy. default: True

See also:

`io.load_trials`

allensdk.brain_observatory.behavior.mtrain.**fix_change_time**(*trials*)

forces *None* values in the *change_time* column to numpy NaN

Parameters

trials
[pandas DataFrame] dataframe of trials

inplace
[bool, optional] modify *trials* in place. if False, returns a copy. default: True

See also:

`io.load_trials`

allensdk.brain_observatory.behavior.rewards_processing module

allensdk.brain_observatory.behavior.rewards_processing.**get_rewards**(*data: Dict*, *timestamps: ndarray*) → DataFrame

Construct and return a pandas DataFrame containing reward data for this session

Parameters

data: Dict
The dict that results from reading the stimulus pickle file associated with the session

timestamps: np.ndarray[1d]
A numpy array of timestamps associated with the stimulus frames in this session. timestamps[ii] is the clock time of the iith frame.

Returns

pd.DataFrame
containing the data associated with rewards given in this session

allensdk.brain_observatory.behavior.schemas module

class allensdk.brain_observatory.behavior.schemas.**BehaviorMetadataSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

This schema contains metadata pertaining to behavior.

neurodata_doc = 'Metadata for behavior and behavior + ophys experiments'

neurodata_skip = {'date_of_acquisition'}

neurodata_type = 'BehaviorMetadata'

neurodata_type_inc = 'LabMetaData'

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.behavior.schemas.**BehaviorTaskParametersSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

This schema encompasses task parameters used for behavior or ophys + behavior.

neurodata_doc = 'Metadata for behavior or behavior + ophys task parameters'

neurodata_type = 'BehaviorTaskParameters'

neurodata_type_inc = 'LabMetaData'

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.behavior.schemas.**CompleteOphysBehaviorMetadataSchema**(*, *only: types.StrSequenceOrSe | None = None*, *exclude: types.StrSequenceOrSe = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSe = ()*, *dump_only: types.StrSequenceOrSe = ()*, *partial: bool | types.StrSequenceOrSe | None = None*, *unknown: str | None = None*)

Bases: *OphysBehaviorMetadataSchema*, *SubjectMetadataSchema*

This schema combines fields from behavior, ophys, and subject schemas. Metadata info is passed by the behavior+ophys session in a combined lump containing all the field types.

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> `Field` objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.behavior.schemas.**EyeTrackingRigGeometry**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

Eye tracking rig geometry

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.behavior.schemas.**NwbOphysMetadataSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

This schema contains fields that will be stored in pyNWB base classes pertaining to optical physiology.

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.behavior.schemas.**OphysBehaviorMetadataSchema**(**, only: types.StrSequenceOrSet | None = None, exclude: types.StrSequenceOrSet = (), many: bool = False, context: dict | None = None, load_only: types.StrSequenceOrSet = (), dump_only: types.StrSequenceOrSet = (), partial: bool | types.StrSequenceOrSet | None = None, unknown: str | None = None*)

Bases: *BehaviorMetadataSchema*, *OphysMetadataSchema*

This schema contains fields pertaining to ophys+behavior. It is used as a template for generating our custom NWB behavior + ophys extension.

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> `Field` objects

load_fields: Dict[str, ma_fields.Field]

neurodata_doc = 'Metadata for behavior + ophys experiments'

neurodata_skip = {'date_of_acquisition', 'emission_lambda', 'excitation_lambda', 'indicator', 'ophys_frame_rate', 'targeted_structure'}

neurodata_type = 'OphysBehaviorMetadata'

neurodata_type_inc = 'BehaviorMetadata'

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.behavior.schemas.**OphysEyeTrackingRigMetadataSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

This schema encompasses metadata for ophys experiment rig

neurodata_doc = 'Metadata for ophys experiment rig'

neurodata_type = 'OphysEyeTrackingRigMetadata'

neurodata_type_inc = 'NWBDataInterface'

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.behavior.schemas.**OphysMetadataSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *NwbOphysMetadataSchema*

This schema contains metadata pertaining to optical physiology (ophys).

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> Field objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.behavior.schemas.**RaisingSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: Schema

class Meta

Bases: object

unknown = 'raise'

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.behavior.schemas.**SubjectMetadataSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

This schema contains metadata pertaining to a subject in either a behavior or behavior + ophys experiment.

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> `Field` objects

load_fields: Dict[str, ma_fields.Field]

neurodata_doc = 'Metadata for an AIBS behavior or behavior + ophys subject'

neurodata_skip = {'age_in_days', 'genotype', 'sex', 'subject_id'}

neurodata_type = 'BehaviorSubject'

neurodata_type_inc = 'Subject'

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

allensdk.brain_observatory.behavior.session_metrics module

allensdk.brain_observatory.behavior.session_metrics.**num_contingent_trials**(*session_trials*)

Returns the number of "go" and "catch" trials in a training session dataframe. Args:

session_trials (pandas.DataFrame): a pandas.DataFrame describing behavior training trials, with the string column "trial_type" describing the type of trial.

Returns (int): Number of "go" and "catch" trials

allensdk.brain_observatory.behavior.session_metrics.**response_bias**(*trials*, *detect_col*, *trial_types=('go', 'catch')*)

Calculate the response bias for a subset of trial types from a behavioral training dataframe. Args:

trials (pandas.DataFrame): Dataframe containing trial-level information
from a behavioral training session. Required columns: "trial_type", *detect_col*.

detect_col (str): Name of column containing boolean
or numeric codings (0/1) for whether or not the mouse had a response.

trial_types (iterable<str>): Iterable containing string trial types
to check for the response bias. Trials of types not included in this iterable will be ignored. Default=("go", "catch")

Return:
The response bias (or average value of the *detect_col*) for trials in *trial_types*.

allensdk.brain_observatory.behavior.stimulus_processing module

allensdk.brain_observatory.behavior.trial_masks module

allensdk.brain_observatory.behavior.trial_masks.**contingent_trials**(*trials*)

GO & CATCH trials only

Parameters

trials
[pandas DataFrame] dataframe of trials

Returns

mask
[pandas Series of booleans, indexed to trials DataFrame]

allensdk.brain_observatory.behavior.trial_masks.**reward_rate**(*trials*, *thresh=2.0*)

masks trials where the reward rate (per minute) is below some threshold.

This de facto omits trials in which the animal was not licking for extended periods or periods when they were licking indiscriminantly.

Parameters

trials
[pandas DataFrame] dataframe of trials

thresh
[float, optional] threshold under which trials will not be included, default: 2.0

Returns

mask
[pandas Series of booleans, indexed to trials DataFrame]

allensdk.brain_observatory.behavior.trial_masks.**trial_types**(*trials*, *trial_types*)

only include trials of certain trial types

Parameters

trials
[pandas DataFrame] dataframe of trials

trial_types
[list or other iterator]

Returns

mask
[pandas Series of booleans, indexed to trials DataFrame]

allensdk.brain_observatory.behavior.trials_processing module

Module contents

allensdk.brain_observatory.data_release_utils package

Subpackages

allensdk.brain_observatory.data_release_utils.metadata_utils package

Submodules

allensdk.brain_observatory.data_release_utils.metadata_utils.id_generator module

class allensdk.brain_observatory.data_release_utils.metadata_utils.id_generator.**FileIDGenerator**

Bases: `object`

A class to generate a unique integer ID for each file in the data release

property dummy_value: int

Value reserved for files that are missing from the release

id_from_path(*file_path: Path*) → int

Get the unique ID for a file path. If the file has already been assigned a unique ID, return that. Otherwise, assign a unique ID to the file path and return it

allensdk.brain_observatory.data_release_utils.metadata_utils.utils module

allensdk.brain_observatory.data_release_utils.metadata_utils.utils.**add_file_paths_to_metadata_table**(*meta*
Data
id_g
File
D-
Gen
er-
a-
tor,
*file_*
Path
*file_*
str
|
Non
in-
*dex_*
str,
data
str
|
Non
on_r
str,
*file_*
str
=
'nwb
*file_*
str
|
Non
=
Non
→
Data

Add file_id and file_path columns to session dataframe.

Parameters

metadata_table: pd.DataFrame
The dataframe to which we are adding file_id and file_path

id_generator: FileIDGenerator
For maintaining a unique mapping between file_path and file_id

file_dir: pathlib.Path
directory where files will be found

file_prefix: str
Prefix of file names

index_col: str
Column in metadata_table used to index files

data_dir_col
Column in metadata_table denoting directory structure of data For example if data is stored

under each session_id

<session_id> / file_a <session_id> / file_b ...

then give the name of the session_id col here

If None, data is assumed to be stored flat

on_missing_file: str

Specifies how to handle missing files
'error' -> raise an exception 'warning' -> assign dummy file_id and warn 'skip' -> drop that row from the table and warn

file_suffix
file_stem
Explicit file stem. Overrides dynamic naming of files

Returns

metadata_table:
The same as the input dataframe but with file_id and file_path columns added

Notes

Files are assumed to be named like {file_dir}/{file_prefix}_{metadata_table.index_col}.{file_suffix}

Module contents

Module contents

allensdk.brain_observatory.ecephys package

Subpackages

allensdk.brain_observatory.ecephys.align_timestamps package

Submodules

allensdk.brain_observatory.ecephys.align_timestamps.barcode module

allensdk.brain_observatory.ecephys.align_timestamps.barcode.**extract_barcodes_from_times**(*on_times*, *off_times*, *inter_barcode_interva* *bar_duration=0.03*, *barcode_duration_ceili* *nbits=32*)

Read barcodes from timestamped rising and falling edges.

Parameters

on_times
[numpy.ndarray] Timestamps of rising edges on the barcode line

off_times
[numpy.ndarray] Timestamps of falling edges on the barcode line

inter_barcode_interval
[numeric, optional] Minimun duration of time between barcodes.

bar_duration
[numeric, optional] A value slightly shorter than the expected duration of each bar

barcode_duration_ceiling
[numeric, optional] The maximum duration of a single barcode

nbits
[int, optional] The bit-depth of each barcode

Returns

barcode_start_times
[list of numeric] For each detected barcode, the time at which that barcode started

barcodes
[list of int] For each detected barcode, the value of that barcode as an integer.

Notes

ignores first code in prod (ok, but not intended) ignores first on pulse (intended - this is needed to identify that a barcode is starting)

allensdk.brain_observatory.ecephys.align_timestamps.barcode.**find_matching_index**(*master_barcodes*, *probe_barcodes*, *alignment_type='start'*)

Given a set of barcodes for the master clock and the probe clock, find the indices of a matching set, either starting from the beginning or the end of the list.

Parameters

master_barcodes
[np.ndarray] barcode values on the master line. One per barcode

probe_barcodes
[np.ndarray] barcode values on the probe line. One per barcode

alignment_type
[string] 'start' or 'end'

Returns

master_barcode_index
[int] matching index for master barcodes (None if not found)

probe_barcode_index
[int] matching index for probe barcodes (None if not found)

allensdk.brain_observatory.ecephys.align_timestamps.barcode.**get_probe_time_offset**(*master_times*, *master_barcodes*, *probe_times*, *probe_barcodes*, *acq_start_index*, *local_probe_rate*)

Time offset between master clock and recording probes. For converting probe time to master clock.

Parameters

master_times
[np.ndarray] start times of barcodes (according to the master clock) on the master line. One per barcode.

master_barcodes
[np.ndarray] barcode values on the master line. One per barcode

probe_times
[np.ndarray] start times (according to the probe clock) of barcodes on the probe line. One per barcode

probe_barcodes
[np.ndarray] barcode values on the probe_line. One per barcode

acq_start_index
[int] sample index of probe acquisition start time

local_probe_rate
[float] the probe's apparent sampling rate

Returns

total_time_shift
[float] Time at which the probe started acquisition, assessed on the master clock. If < 0, the probe started earlier than the master line.

probe_rate
[float] The probe's sampling rate, assessed on the master clock

master_endpoints
[iterable] Defines the start and end times of the sync interval on the master clock

allensdk.brain_observatory.ecephys.align_timestamps.barcode.**linear_transform_from_intervals**(*master*, *probe*)

Find a scale and translation which aligns two 1d segments

Parameters

master
[iterable] Pair of floats defining the master interval. Order is [start, end].

probe
[iterable] Pair of floats defining the probe interval. Order is [start, end].

Returns

scale
[float] Scale factor. If > 1.0, the probe clock is running fast compared to the master clock. If < 1.0, the probe clock is running slow.

translation
[float] If > 0, the probe clock started before the master clock. If > 0, after.

Notes

solves
(master + translation) * scale = probe

for scale and translation

allensdk.brain_observatory.ecephys.align_timestamps.barcode.**match_barcodes**(*master_times*, *master_barcodes*, *probe_times*, *probe_barcodes*)

Given sequences of barcode values and (local) times on a probe line and a master line, find the time points on each clock corresponding to the first and last shared barcode.

If there's only one probe barcode, only the first matching timepoint is returned.

Parameters

master_times
[np.ndarray] start times of barcodes (according to the master clock) on the master line. One per barcode.

master_barcodes
[np.ndarray] barcode values on the master line. One per barcode

probe_times
[np.ndarray] start times (according to the probe clock) of barcodes on the probe line. One per barcode

probe_barcodes
[np.ndarray] barcode values on the probe_line. One per barcode

Returns

probe_interval
[np.ndarray] Start and end times of the matched interval according to the probe_clock.

master_interval
[np.ndarray] Start and end times of the matched interval according to the master clock

allensdk.brain_observatory.ecephys.align_timestamps.barcode_sync_dataset module

class allensdk.brain_observatory.ecephys.align_timestamps.barcode_sync_dataset.**BarcodeSyncDataset**

Bases: *EcephysSyncDataset*

property barcode_line

Obtain the index of the barcode line for this dataset.

extract_barcodes(***barcode_kwargs*)

Read barcodes and their times from this dataset's barcode line.

Parameters

****barcode_kwargs**
Will be passed to .barcode.extract_barcodes_from_times

Returns

times
[np.ndarray] The start times of each detected barcode.

codes
[np.ndarray] The values of each detected barcode

`get_barcode_table`(***barcode_kwargs*)

A convenience method for getting barcode times and codes in a dictionary.

Notes

This method is deprecated!

allensdk.brain_observatory.ecephys.align_timestamps.channel_states module

allensdk.brain_observatory.ecephys.align_timestamps.channel_states.**extract_barcodes_from_states**(*channel_s times- tamps, sam- pling_rat **bar- code_kwa*

Obtain barcodes from timestamped rising/falling edges.

Parameters

channel_states
[numpy.ndarray] Rising and falling edges, denoted 1 and -1

timestamps
[numpy.ndarray] Sample index of each event.

sampling_rate
[numeric] Samples / second

****barcode_kwargs**
Additional parameters describing the barcodes.

allensdk.brain_observatory.ecephys.align_timestamps.channel_states.**extract_splits_from_barcode_times**(*ba tol- er- an*

Determine locations of likely dropped data from barcode times Parameters ———- barcode_times : numpy.ndarray

probe barcode times

tolerance
[float] Timing tolerance (relative to median interval)

allensdk.brain_observatory.ecephys.align_timestamps.channel_states.**extract_splits_from_states**(*channel_sta times- tamps, sam- pling_rate, **bar- code_kwarg*

Obtain data split times from timestamped rising/falling edges.

Parameters

channel_states
[numpy.ndarray] Rising and falling edges, denoted 1 and -1

timestamps
[numpy.ndarray] Sample index of each event.

sampling_rate
[numeric] Samples / second

****barcode_kwargs**
Additional parameters describing the barcodes.

allensdk.brain_observatory.ecephys.align_timestamps.probe_synchronizer module

class allensdk.brain_observatory.ecephys.align_timestamps.probe_synchronizer.**ProbeSynchronizer**(*global_pro*
lo-
*cal_probe_*
to-
tal_time_si
min_time,
max_time)

Bases: `object`

classmethod compute(*master_barcode_times*, *master_barcodes*, *probe_barcode_times*, *probe_barcodes*, *min_time*, *max_time*, *probe_start_index*, *local_probe_sampling_rate*)

Compute a transform from probe samples to master times by aligning barcodes.

Parameters

master_barcode_times
[np.ndarray] start times of barcodes (according to the master clock) on the master line. One per barcode.

master_barcodes
[np.ndarray] barcode values on the master line. One per barcode

probe_barcode_times
[np.ndarray] start times (according to the probe clock) of barcodes on the probe line. One per barcode

probe_barcodes
[np.ndarray] barcode values on the probe_line. One per barcode

min_time
[Float] time (in seconds) of first barcode to align

max_time
[Float] time (in seconds) of last barcode to align

probe_start_index
[int] sample index of probe acquisition start time

local_probe_sampling_rate
[float] the probe's apparent sampling rate

Returns

ProbeSynchronizer
When called, applies the transform computed here to samples on the probe clock.

`property sampling_rate_scale`

The ratio of the probe's sampling rate assessed on the global clock to the probe's locally assessed sampling rate.

Module contents

allensdk.brain_observatory.ecephys.copy_utility package

Module contents

allensdk.brain_observatory.ecephys.current_source_density package

Module contents

allensdk.brain_observatory.ecephys.data_objects package

Submodules

allensdk.brain_observatory.ecephys.data_objects.trials module

Module contents

allensdk.brain_observatory.ecephys.ecephys_project_api package

Submodules

allensdk.brain_observatory.ecephys.ecephys_project_api.ecephys_project_api module

`class allensdk.brain_observatory.ecephys.ecephys_project_api.ecephys_project_api.EcephysProjectApi`

Bases: `object`

`get_channels`(*channel_ids: ArrayLike | None = None*, *probe_ids: ArrayLike | None = None*, *session_ids: ArrayLike | None = None*, *published_at: str | None = None*)

`get_isi_experiments`(**args*, ***kwargs*)

`get_natural_movie_template`(*number*) → Iterable

`get_natural_scene_template`(*number*) → Iterable

`get_probe_lfp_data`(*probe_id: int*) → Iterable

`get_probes`(*probe_ids: ArrayLike | None = None*, *session_ids: ArrayLike | None = None*, *published_at: str | None = None*)

get_session_data(*session_id: int*) → Iterable

get_sessions(*session_ids: ArrayLike | None = None*, *published_at: str | None = None*)

get_unit_analysis_metrics(*unit_ids: ArrayLike | None = None*, *ecephys_session_ids: ArrayLike | None = None*, *session_types: ArrayLike | None = None*) → DataFrame

get_units(*unit_ids: ArrayLike | None = None*, *channel_ids: ArrayLike | None = None*, *probe_ids: ArrayLike | None = None*, *session_ids: ArrayLike | None = None*, *published_at: str | None = None*)

allensdk.brain_observatory.ecephys.ecephys_project_api.ecephys_project_fixed_api module

allensdk.brain_observatory.ecephys.ecephys_project_api.ecephys_project_lims_api module

allensdk.brain_observatory.ecephys.ecephys_project_api.ecephys_project_warehouse_api module

allensdk.brain_observatory.ecephys.ecephys_project_api.http_engine module

allensdk.brain_observatory.ecephys.ecephys_project_api.rma_engine module

allensdk.brain_observatory.ecephys.ecephys_project_api.utilities module

Module contents

allensdk.brain_observatory.ecephys.ecephys_session_api package

Submodules

allensdk.brain_observatory.ecephys.ecephys_session_api.ecephys_nwb1_session_api module

allensdk.brain_observatory.ecephys.ecephys_session_api.ecephys_nwb_session_api module

allensdk.brain_observatory.ecephys.ecephys_session_api.ecephys_session_api module

class allensdk.brain_observatory.ecephys.ecephys_session_api.ecephys_session_api.**EcephysSessionApi**(**args*, ***kwa*

Bases: `object`

get_channels() → DataFrame

get_ecephys_session_id() → int

get_invalid_times() → DataFrame

get_lfp(*probe_id: int*) → DataArray

get_mean_waveforms() → Dict[int, ndarray]

get_metadata()

get_optogenetic_stimulation() → DataFrame

get_probes() → DataFrame

get_pupil_data() → DataFrame | None

get_rig_metadata() → dict | None

get_running_speed() → *RunningSpeed*

get_screen_gaze_data(*include_filtered_data=False*) → DataFrame | None

get_session_start_time() → datetime

get_spike_amplitudes() → Dict[int, ndarray]

get_spike_times() → Dict[int, ndarray]

get_stimulus_presentations() → DataFrame

get_units() → DataFrame

session_na = -1

test() → bool

Module contents

allensdk.brain_observatory.ecephys.file_io package

Submodules

allensdk.brain_observatory.ecephys.file_io.continuous_file module

class allensdk.brain_observatory.ecephys.file_io.continuous_file.**ContinuousFile**(*data_path*, *timestamps_path*, *total_num_channels=384*, *dtype=<class 'numpy.int16'>*)

Bases: object

Represents a continuous (.dat) file, and its associated timestamps

get_lfp_channel_order()

Returns the channel ordering for LFP data extracted from NPX files.

Parameters:

None

load(*memmap=False*, *memmap_thresh=10000000000.0*)

Reads lfp data and timestamps from the filesystem

Parameters:

memmap
[bool, optional] If True, the returned data array will be a memory map of the file on disk. Default is True.

memmap_thresh
[float, optional] Files above this size in bytes will be memory-mapped, regardless of memmap setting

allensdk.brain_observatory.ecephys.file_io.ecephys_sync_dataset module

class allensdk.brain_observatory.ecephys.file_io.ecephys_sync_dataset.**EcephysSyncDataset**

Bases: *Dataset*

extract_frame_times(*strategy*, *photodiode_cycle=60*, *frame_keys=('frames', 'stim_vsync', 'vsync_stim')*, *photodiode_keys=('photodiode', 'stim_photodiode')*, *trim_discontiguous_frame_times=True*)

extract_frame_times_from_photodiode(*photodiode_cycle=60*, *frame_keys=('frames', 'stim_vsync', 'vsync_stim')*, *photodiode_keys=('photodiode', 'stim_photodiode')*, *trim_discontiguous_frame_times=True*)

extract_frame_times_from_vsyncs(*photodiode_cycle=60*, *frame_keys=('frames', 'stim_vsync', 'vsync_stim')*, *photodiode_keys=('photodiode', 'stim_photodiode')*)

extract_led_times(*keys=('LED_sync', 'opto_trial')*, *fallback_line=18*)

classmethod factory(*path*)

Build a new SyncDataset.

Parameters

path
[str] Filesystem path to the h5 file containing sync information to be loaded.

remove_zero_frames(*frame_times*)

property sample_frequency

allensdk.brain_observatory.ecephys.file_io.stim_file module

class allensdk.brain_observatory.ecephys.file_io.stim_file.**CamStimOnePickleStimFile**(*data*, ***kwargs*)

Bases: object

property angular_wheel_rotation

Extract the total rotation of the running wheel on each frame.

property angular_wheel_velocity

Extract the mean angular velocity of the running wheel (degrees / s) for each frame.

classmethod factory(*path*, ***kwargs*)

property frames_per_second

Framerate of stimulus presentation

property pre_blank_sec

Time (s) before initial stimulus presentation

property stimuli

List of dictionaries containing information about individual stimuli

property vin

property vsig

Running speed signal voltage

Module contents

allensdk.brain_observatory.ecephys.lfp_subsampling package

Submodules

allensdk.brain_observatory.ecephys.lfp_subsampling.subsampling module

allensdk.brain_observatory.ecephys.lfp_subsampling.subsampling.**remove_lfp_noise**(*lfp*, *surface_channel*, *channel_numbers*, *channel_max=384*, *channel_limit=380*, *max_out_of_brain_channels=*

Subtract mean of channels out of brain to remove noise

Parameters:

lfp
[numpy.ndarray] 2D array of LFP values (time x channels)

surface_channel
[int] Surface channel (relative to original probe)

channel_numbers
[numpy.ndarray] Channel numbers in 'lfp' array (relative to original probe)

max_out_of_brain_channels: int
Rereferencing can sometimes fail for experiments with shallow probe insertions as the uppermost channels are in air and not agar. This places a limit on the number of channels to use for re-referencing.

Returns:

lfp_noise_removed
[numpy.ndarray] New 2D array of LFP values

allensdk.brain_observatory.ecephys.lfp_subsampling.subsampling.**remove_lfp_offset**(*lfp*, *sampling_frequency*, *cutoff_frequency*, *filter_order*)

High-pass filters LFP data to remove offset

Parameters:

lfp
[numpy.ndarray] 2D array of LFP values (time x channels)

sampling_frequency
[float] Sampling frequency in Hz

cutoff_frequency
[float] Cutoff frequency for highpass filter

filter_order
[int] Butterworth filter order

Returns:

lfp_filtered
[numpy.ndarray] New 2D array of LFP values

allensdk.brain_observatory.ecephys.lfp_subsampling.subsampling.**select_channels**(*total_channels*, *surface_channel*, *surface_padding*, *start_channel_offset*, *channel_stride*, *channel_order*, *noisy_channels=array([], dtype=float64)*, *remove_noisy_channels=False*, *reference_channels=array([], dtype=float64)*, *remove_references=False*)

Selects a subset of channels for spatial downsampling

Parameters:

total_channels
[int] Number of channels in the original data file

surface_channel
[int] Index of channel at brain surface

surface_padding
[int] Number of channels above surface to save

start_channel_offset
[int] First channel to save

channel_stride
[int] Number of channels to skip in output

channel_order
[np.ndarray] Actual order of LFP channels (needed to account for the bug in NPX extraction)

noisy_channels
[numpy.ndarray] Array indicating noisy channels

remove_noisy_channels
[bool] Flag to remove noisy channels

reference_channels
[numpy.ndarray] Array indicating refence channels

remove_references
[bool] Flag to remove reference channels

allensdk.brain_observatory.ecephys.lfp_subsampling.subsampling.**subsample_lfp**(*lfp_raw*, *selected_channels*, *subsampling_factor*)

Subsamples LFP data

Parameters:

lfp_raw
[numpy.ndarray] 2D array of LFP values (time x channels)

selected_channels
[numpy.ndarray] Indices of channels to select (spatial subsampling)

downsampling_factor
[int] Factor by which to subsample in time

Returns:

lfp_subsampled
[numpy.ndarray] New 2D array of LFP values

allensdk.brain_observatory.ecephys.lfp_subsampling.subsampling.**subsample_timestamps**(*timestamps*, *subsampling_factor*)

Subsamples an array of timestamps

Parameters:

timestamps
[numpy.ndarray] 1D array of timestamp values

downsampling_factor
[int] Factor by which to subsample the timestamps

Returns:

timestamps_sub
[numpy.ndarray] New 1D array of timestamps

Module contents

allensdk.brain_observatory.ecephys.nwb package

Submodules

allensdk.brain_observatory.ecephys.nwb.ecephys_nwb_extension_builder module

Module contents

allensdk.brain_observatory.ecephys.optotagging_table package

Module contents

allensdk.brain_observatory.ecephys.stimulus_analysis package

Submodules

allensdk.brain_observatory.ecephys.stimulus_analysis.dot_motion module

allensdk.brain_observatory.ecephys.stimulus_analysis.drifting_gratings module

allensdk.brain_observatory.ecephys.stimulus_analysis.flashes module

allensdk.brain_observatory.ecephys.stimulus_analysis.natural_movies module

allensdk.brain_observatory.ecephys.stimulus_analysis.natural_scenes module

allensdk.brain_observatory.ecephys.stimulus_analysis.receptive_field_mapping module

allensdk.brain_observatory.ecephys.stimulus_analysis.static_gratings module

allensdk.brain_observatory.ecephys.stimulus_analysis.stimulus_analysis module

Module contents

allensdk.brain_observatory.ecephys.stimulus_table package

Subpackages

allensdk.brain_observatory.ecephys.stimulus_table.visualization package

Submodules

allensdk.brain_observatory.ecephys.stimulus_table.visualization.view_blocks module

allensdk.brain_observatory.ecephys.stimulus_table.visualization.view_blocks.**build_colormap**(*table*, *existing_map={}*, *base_colors=[(0.788235294117, 0.956862745098, (1.0, 0.705882352941, 0.509803921568, (0.552941176471, 0.898039215686, 0.631372549019, (1.0, 0.623529411764, 0.607843137254, (0.815686274509, 0.733333333333, 1.0), (0.870588235294, 0.733333333333, 0.607843137254, (0.980392156863, 0.690196078431, 0.894117647059, (0.811764705882, 0.811764705882, 0.811764705882, (1.0, 0.996078431373, 0.639215686275, (0.725490196078, 0.949019607843, 0.941176470588*)

allensdk.brain_observatory.ecephys.stimulus_table.visualization.view_blocks.**get_blocks**(*table*)

allensdk.brain_observatory.ecephys.stimulus_table.visualization.view_blocks.**main**(*table_csv_path*)

allensdk.brain_observatory.ecephys.stimulus_table.visualization.view_blocks.**plot_blocks**(*blocks*, *colormap*)

Module contents

Submodules

allensdk.brain_observatory.ecephys.stimulus_table.ephys_pre_spikes module

allensdk.brain_observatory.ecephys.stimulus_table.ephys_pre_spikes.**apply_display_sequence**(*sweep_frames_ta*
frame_display_se
start_key='Start',
end_key='End',
diff_key='dif',
block_key='stimu

Adjust raw sweep frames for a stimulus based on the display sequence for that stimulus.

Parameters

sweep_frames_table
[pd.DataFrame] Each row is a sweep. Has two columns, 'start' and 'end', which describe (in frames) when that sweep began and ended.

frame_display_sequence
[np.ndarray] 2D array. Rows are display intervals. The 0th column is the start frame of that interval, the 1st the end frame.

Returns

sweep_frames_table
[pd.DataFrame] As above, but start and end frames have been adjusted based on the display sequence.

Notes

The frame values in the raw sweep_frames_table are given in 0-indexed offsets from the start of display for this stimulus. This domain only takes into account frames which are part of a display interval for that stimulus, so the frame ids need to be adjusted to lie on the global frame sequence.

allensdk.brain_observatory.ecephys.stimulus_table.ephys_pre_spikes.**apply_frame_times**(*stimulus_table*,
frame_times,
frames_per_second=No
ex-
tra_frame_time=False,
map_columns=('Start',
'End'))

Converts sweep times from frames to seconds.

Parameters

stimulus_table
[pd.DataFrame] Rows are sweeps. Columns are stimulus parameters as well as start and end frames for each sweep.

frame_times
[numpy.ndarrray] Gives the time in seconds at which each frame (indices) began.

frames_per_second
[numeric, optional] If provided, and extra_frame_time is True, will be used to calculcate the extra_frame_time.

extra_frame_time
[float, optional] If provided, an additional frame time will be appended. The time will be incremented by extra_frame_time from the previous last frame time, to denote the time at which the last frame ended. If False, no extra time will be appended. If None (default), the increment will be 1.0/fps.

map_columns
[tuple of str, optional] Which columns to replace with times. Defaults to 'Start' and 'End

Returns

stimulus_table
[pd.DataFrame] As above, but with map_columns values converted to seconds from frames.

allensdk.brain_observatory.ecephys.stimulus_table.ephys_pre_spikes.**assign_sweep_values**(*stim_table*, *sweep_table*, *on='sweep_number'*, *drop=True*, *tmp_suffix='_stimtab*

Left joins a stimulus table to a sweep table in order to associate
epochs in time with stimulus characteristics.

Parameters

stim_table
[pd.DataFrame] Each row is a stimulus epoch, with start and end times and a foreign key onto a particular sweep.

sweep_table
[pd.DataFrame] Each row is a sweep. Should have columns in common with the stim_table - the resulting table will use values from the sweep_table.

on
[str, optional] Column on which to join.

drop
[bool, optional] If True (default), the join column (argument on) will be dropped from the output.

tmp_suffix
[str, optional] Will be used to identify overlapping columns. Should not appear in the name of any column in either dataframe.

allensdk.brain_observatory.ecephys.stimulus_table.ephys_pre_spikes.**build_stimuluswise_table**(*stimulus*, *seconds_to_frame start_key='Sta end_key='End name_key='sti block_key='sti get_stimulus_n extract_const_pa drop_const_pa 'autoLog', 'autoDraw', 'win')*)

Construct a table of sweeps, including their times on the experiment-global clock and the values of each relevant parameter.

Parameters

stimulus
[dict] Describes presentation of a stimulus on a particular experiment. Has a number of fields, of which we are using:

stim_path
[str] windows file path to the stimulus data

sweep_frames
[list of lists] rows are sweeps, columns are start and end frames of that sweep (in the stimulus-specific frame domain). C-order.

sweep_order
[list of int] indices are frames, values are the sweep on that frame

display_sequence
[list of list]

rows are intervals in which the stimulus was displayed. Columns are start and end times (s, global) of the display. C-order.

dimnames
[list of str] Names of parameters for this stimulus (such as "Contrast")

sweep_table
[list of tuple] Each element is a tuple of parameter values (1 per dimname) describing a single sweep.

seconds_to_frames
[function] Converts experiment seconds to frames

start_key
[str, optional] key to use for start frame indices. Defaults to 'Start'

end_key
[str, optional] key to use for end frame indices. Defaults to 'End'

name_key
[str, optional] key to use for stimulus name annotations. Defaults to 'stimulus_name'

block_key
[str, optional] key to use for the 0-index position of this stimulus block

get_stimulus_name
[function | dict -> str, optional] extracts stimulus name from the stimulus dictionary. Default is read_stimulus_name_from_path

Returns

list of pandas.DataFrame
Each table corresponds to an entry in the display sequence. Rows are sweeps, columns are stimulus parameter values as well as "Start" and "End".

allensdk.brain_observatory.ecephys.stimulus_table.ephys_pre_spikes.**create_stim_table**(*stimuli*, *stimulus_tabler*, *spontaneous_activity_tabler*, *sort_key='Start'*, *block_key='stimulus_bl*
index_key='stimulus_inde

Build a full stimulus table

Parameters

stimuli
[list of dict] Each element is a stimulus dictionary, as provided by the stim.pkl file.

stimulus_tabler
[function] A function which takes a single stimulus dictionary as its argument and returns a stimulus table dataframe.

spontaneous_activity_tabler
[function] A function which takes a list of stimulus tables as arguments and returns a list of 0 or more tables describing spontaneous activity sweeps.

sort_key
[str, optional] Sort the final stimulus table in ascending order by this key. Defaults to 'Start'.

Returns

stim_table_full
[pandas.DataFrame] Each row is a sweep. Has columns describing (in frames) the start and end times of each sweep. Other columns describe the values of stimulus parameters on those sweeps.

allensdk.brain_observatory.ecephys.stimulus_table.ephys_pre_spikes.**make_spontaneous_activity_tables**(*stim*
start
*end_*
*duration_*

Fills in frame gaps in a set of stimulus tables. Suitable for use as the spontaneous_activity_tabler in create_stim_table.

Parameters

stimulus_tables
[list of pd.DataFrame] Input tables - should have start_key and end_key columns.

start_key
[str, optional] Column name for the start of a sweep. Defaults to 'Start'.

end_key
[str, optional] Column name for the end of a sweep. Defaults to 'End'.

duration_threshold
[numeric or None] If not None (default is 0), remove spontaneous activity sweeps whose duration is less than this threshold.

Returns

list
Either empty, or contains a single pd.DataFrame. The rows of the dataframe are spontaneous activity sweeps.

allensdk.brain_observatory.ecephys.stimulus_table.ephys_pre_spikes.**read_stimulus_name_from_path**(*stimulus*)

Obtains a human-readable stimulus name by looking at the filename of the 'stim_path' item.

Parameters

stimulus
[dict] must contain a 'stim_path' item.

Returns

str
name of stimulus

allensdk.brain_observatory.ecephys.stimulus_table.ephys_pre_spikes.**split_column**(*table*, *column*, *new_columns*, *drop_old=True*)

Divides a dataframe column into multiple columns.

Parameters

table
[pandas.DataFrame] Columns will be drawn from and assigned to this dataframe. This dataframe will NOT be modified inplace.

column
[str] This column will be split.

new_columns
[dict, mapping strings to functions] Each key will be the name of a new column, while its value (a function) will be used to build the new column's values. The functions should map from a single value of the original column to a single value of the new column.

drop_old
[bool, optional] If True, the original column will be dropped from the table.

Returns

table
[pd.DataFrame] The modified table

allensdk.brain_observatory.ecephys.stimulus_table.naming_utilities module

allensdk.brain_observatory.ecephys.stimulus_table.naming_utilities.**add_number_to_shuffled_movie**(*table*, *natural_movie_template_re=*, *stim_colname=*, *template='natural_movie_*, *tmp_colname=*

allensdk.brain_observatory.ecephys.stimulus_table.naming_utilities.**collapse_columns**(*table*)

merge, where possible, columns that describe the same parameter. This is pretty conservative - it only matches columns by capitalization and it only overrides nans.

allensdk.brain_observatory.ecephys.stimulus_table.naming_utilities.**drop_empty_columns**(*table*)

Remove from the stimulus table columns whose values are all nan

allensdk.brain_observatory.ecephys.stimulus_table.naming_utilities.**map_column_names**(*table*, *name_map=None*, *ignore_case=True*)

allensdk.brain_observatory.ecephys.stimulus_table.naming_utilities.**map_stimulus_names**(*table*, *name_map=None*, *stim_colname='stimuli*

Applies a mappting to the stimulus names in a stimulus table

Parameters

table
[pd.DataFrame] the input stimulus table

name_map
[dict, optional] rename the stimuli according to this mapping

stim_colname: str, optional
look in this column for stimulus names

allensdk.brain_observatory.ecephys.stimulus_table.naming_utilities.**standardize_movie_numbers**(*table*, *movie_re=re.* *numeral_re=re.* *digit_names={* *'one',* *'2':* *'two',* *'3':* *'three',* *'4':* *'four',* *'5':* *'five',* *'6':* *'six',* *'7':* *'seven',* *'8':* *'eight',* *'9':* *'nine'}*, *stim_colname*

Natural movie stimuli in visual coding are numbered using words, like "natural_movie_two" rather than "natural_movie_2". This function ensures that all of the natural movie stimuli in an experiment are named by that convention.

Parameters

table
[pd.DataFrame] the incoming stimulus table

movie_re
[re.Pattern, optional] regex that matches movie stimulus names

numeral_re
[re.Pattern, optional] regex that extracts movie numbers from stimulus names

digit_names
[dict, optional] map from numerals to english words

stim_colname
[str, optional] the name of the dataframe column that contains stimulus names

Returns

table
[pd.DataFrame] the stimulus table with movie numerals having been mapped to english words

allensdk.brain_observatory.ecephys.stimulus_table.output_validation module

allensdk.brain_observatory.ecephys.stimulus_table.output_validation.**validate_epoch_durations**(*table*, *start_key='St*
end_key='En
fail_on_negat

allensdk.brain_observatory.ecephys.stimulus_table.output_validation.**validate_epoch_order**(*table*, *time_keys=('Start',*
'End'))

allensdk.brain_observatory.ecephys.stimulus_table.output_validation.**validate_max_spontaneous_epoch_durat**

allensdk.brain_observatory.ecephys.stimulus_table.stimulus_parameter_extraction module

allensdk.brain_observatory.ecephys.stimulus_table.stimulus_parameter_extraction.**extract_const_params_fr**

Parameters which are not set as sweep_params in the stimulus script (usually because they are not varied during the course of the session) are not output in an easily machine-readable format. This function attempts to recover them by parsing the string repr of the stimulus.

Parameters

stim_repr
[str] The repr of the camstim stimulus object. Served up per-stimulus in the stim pickle.

repr_params_re
[re.Pattern] Extracts attributes as "="-seperated strings

array_re
[re.Pattern] Extracts list reprs from numpy array reprs.

Returns

repr_params
[dict] dictionary of paramater keys and values extracted from the stim repr. Where possible, the values are converted to native Python types.

allensdk.brain_observatory.ecephys.stimulus_table.stimulus_parameter_extraction.**extract_stim_class_from_**

allensdk.brain_observatory.ecephys.stimulus_table.stimulus_parameter_extraction.**parse_stim_repr**(*stim_repr*, *drop_par='autoLog', 'autoDraw', 'win')*, *repr_para='z0-9]+=[^=', re.IGNOR*, *array_re=re.*, *raise_on_*

Read the string representation of a psychopy stimulus and extract stimulus parameters.

Parameters

stim_repr
[str]

drop_params
[tuple]

repr_params_re
[re.Pattern]

array_re
[re.Pattern]

Returns

dict
maps extracted parameter names to values

Module contents

allensdk.brain_observatory.ecephys.visualization package

Module contents

allensdk.brain_observatory.ecephys.visualization.**plot_mean_waveforms**(*mean_waveforms*, *unit_ids*, *peak_channels*)

Utility for plotting mean waveforms on each unit's peak channel

Parameters

mean_waveforms
[dictionary] Maps unit ids to channelwise averege spike waveforms for those units

unit_ids
[array-like] unique integer identifiers for units to be included

allensdk.brain_observatory.ecephys.visualization.**plot_spike_counts**(*data_array*, *time_coords*, *cbar_label*, *title*, *xlabel='time relative to stimulus onset (s)'*, *ylabel='unit'*, *xtick_step=20*)

Utility for making a simple spike counts plot.

Parameters

data_array
[xarray.DataArray] 2D data array unitwise values per time bin. See EcephysSession.sweepwise_spike_counts

allensdk.brain_observatory.ecephys.visualization.**raster_plot**(*spike_times*, *figsize=(8, 8)*, *cmap=<matplotlib.colors.ListedColormap object>*, *title='spike raster'*, *cycle_colors=False*)

allensdk.brain_observatory.ecephys.write_nwb package

Subpackages

allensdk.brain_observatory.ecephys.write_nwb.vbn package

Module contents

Submodules

allensdk.brain_observatory.ecephys.write_nwb.nwb_writer module

allensdk.brain_observatory.ecephys.write_nwb.schemas module

class allensdk.brain_observatory.ecephys.write_nwb.schemas.**BaseBehaviorSessionDataSchema**(*, *only: types.StrSequenceO | None = None*, *exclude: types.StrSequenceO = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceO = ()*, *dump_only: types.StrSequenceO = ()*, *partial: bool | types.StrSequenceO | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> `Field` objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.ecephys.write_nwb.schemas.**BaseNeuropixelsSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: ArgSchema

Base schema for writing NWB files for projects with behavior + ecephys

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.ecephys.write_nwb.schemas.**Channel**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> Field objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

set_field_defaults(*data*, ***kwargs*)

set_impedence_default(*data*, ***kwargs*)

class allensdk.brain_observatory.ecephys.write_nwb.schemas.**InvalidEpoch**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> Field objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.ecephys.write_nwb.schemas.**Lfp**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> `Field` objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.ecephys.write_nwb.schemas.**OutputSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> `Field` objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.ecephys.write_nwb.schemas.**Probe**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> `Field` objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.ecephys.write_nwb.schemas.**ProbeOutputs**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> `Field` objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.ecephys.write_nwb.schemas.**SessionMetadata**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> `Field` objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.ecephys.write_nwb.schemas.**Unit**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> `Field` objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

class allensdk.brain_observatory.ecephys.write_nwb.schemas.**VCNInputSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *BaseNeuropixelsSchema*

Input schema for visual coding neuropixels project

class Meta

Bases: object

unknown = 'raise'

dump_fields: Dict[str, ma_fields.Field]

exclude: set[Any] | MutableSet[Any]

fields: Dict[str, ma_fields.Field]

Dictionary mapping field_names -> Field objects

load_fields: Dict[str, ma_fields.Field]

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

Module contents

Submodules

allensdk.brain_observatory.ecephys.behavior_ecephys_session module

allensdk.brain_observatory.ecephys.ecephys_project_cache module

allensdk.brain_observatory.ecephys.ecephys_session module

allensdk.brain_observatory.ecephys.nwb_util module

allensdk.brain_observatory.ecephys.nwb_util.**add_ecephys_electrodes**(*nwbfile: NWBFile*, *channels: List[dict]*, *electrode_group: EcephysElectrodeGroup*, *channel_number_whitelist: ndarray | None = None*)

Add electrode information to an ecephys nwbfile electrode table.

Parameters

nwbfile
[pynwb.NWBFile] The nwbfile to add electrodes data to

channels
[List[dict]]

A list of 'channel' dictionaries containing the following fields:
id: The unique id for a given electrode/channel probe_id: The unique id for an electrode's/channel's device valid_data: Whether the data for an electrode/channel is usable local_index: The local index of an electrode/channel on a

given device

probe_vertical_position: Length-wise position of electrode/channel
on device (microns)

probe_horizontal_position: Width-wise position of electrode/channel
on device (microns)

structure_id: The LIMS id associated with an anatomical
structure

structure_acronym: Acronym associated with an anatomical
structure

anterior_posterior_ccf_coordinate dorsal_ventral_ccf_coordinate left_right_ccf_coordinate

Optional fields which may be used in the future: impedence: The impedence of a given channel. filtering: The type of hardware filtering done a channel.

(e.g. "1000 Hz low-pass filter")

electrode_group
[EcephysElectrodeGroup] The pynwb electrode group that electrodes should be associated with

channel_number_whitelist
[Optional[np.ndarray], optional] If provided, only add electrodes (a.k.a. channels) specified by the whitelist (and in order specified), by default None

allensdk.brain_observatory.ecephys.nwb_util.**add_probe_to_nwbfile**(*nwbfile*, *probe_id*, *sampling_rate*, *lfp_sampling_rate*, *has_lfp_data*, *name*, *location='See electrode locations'*)

Creates objects required for representation of a single extracellular ephys probe within an NWB file.

Parameters

nwbfile
[pynwb.NWBFile] file to which probe information will be assigned.

probe_id
[int] unique identifier for this probe

sampling_rate: float,
sampling rate of the neuropixels probe

lfp_sampling_rate: float
sampling rate of LFP

has_lfp_data: bool
True if LFP data is available for the probe, otherwise False

name
[str, optional] human-readable name for this probe. Practically, we use tags like "probeA" or "probeB"

location
[str, optional] A required field for the *EcephysElectrodeGroup*. Because the group contains a number of electrodes/channels along the neuropixels probe, location will vary significantly. Thus by default this field is: "See electrode locations" where the nwbfile.electrodes table will provide much more detailed location information.

Returns

nwbfile
[pynwb.NWBFile] the updated file object

probe_nwb_device
[pynwb.device.Device] device object corresponding to this probe

probe_nwb_electrode_group
[pynwb.ecephys.ElectrodeGroup] electrode group object corresponding to this probe

allensdk.brain_observatory.ecephys.nwb_util.**add_ragged_data_to_dynamic_table**(*table*, *data*, *column_name*, *column_description=''*)

Builds the index and data vectors required for writing ragged array data to a pynwb dynamic table

Parameters

table
[pynwb.core.DynamicTable] table to which data will be added (as VectorData / VectorIndex)

data
[dict] each key-value pair describes some grouping of data

column_name
[str] used to set the name of this column

column_description
[str, optional] used to set the description of this column

Returns

nwbfile
[pynwb.NWBFile]

allensdk.brain_observatory.ecephys.optotagging module

class allensdk.brain_observatory.ecephys.optotagging.**OptotaggingTable**(*table: DataFrame*)

Bases: DataObject, JsonReadableInterface, NwbWritableInterface, NwbReadableInterface

Optotagging table - optotagging stimulation

classmethod from_json(*dict_repr: dict*) → *OptotaggingTable*

Populates a DataFile from a JSON compatible dict (likely parsed by argschema)

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

classmethod from_nwb(*nwbfile: NWBFile*) → *OptotaggingTable*

Populate a DataObject from a pyNWB file object.

Parameters

nwbfile:
The file object (NWBFile) of a pynwb dataset file.

Returns

DataObject:
An instantiated DataObject which has *name* and *value* properties

`to_nwb`(*nwbfile: NWBFile*) → NWBFile

Given an already populated DataObject, return an pyNWB file object that had had DataObject data added.

Parameters

nwbfile
[NWBFile] An NWB file object

Returns

NWBFile
An NWB file object that has had data from the DataObject added to it.

`property value: DataFrame`

Returns

A dataframe with columns:

- start_time: onset of stimulation
- condition: optical stimulation pattern
- level: intensity (in volts output to the LED) of stimulation
- stop_time: stop time of stimulation
- stimulus_name: stimulus name
- duration: duration of stimulation

allensdk.brain_observatory.ecephys.probes module

allensdk.brain_observatory.ecephys.stimulus_sync module

allensdk.brain_observatory.ecephys.stimulus_sync.**allocate_by_vsync**(*vs_diff*, *index*, *starts*, *ends*, *frame_duration*, *irregularity*, *cycle*)

allensdk.brain_observatory.ecephys.stimulus_sync.**assign_to_last**(*index*, *starts*, *ends*, *frame_duration*, *irregularity*, *cycle*)

allensdk.brain_observatory.ecephys.stimulus_sync.**compute_frame_times**(*photodiode_times*, *frame_duration*, *num_frames*, *cycle*, *irregular_interval_policy=<function assign_to_last>*)

allensdk.brain_observatory.ecephys.stimulus_sync.**correct_on_off_effects**(*pd_times*)

Notes

This cannot (without additional info) determine whether an assymmetric offset is odd-long or even-long.

allensdk.brain_observatory.ecephys.stimulus_sync.**estimate_frame_duration**(*pd_times*, *cycle=60*)

allensdk.brain_observatory.ecephys.stimulus_sync.**fix_unexpected_edges**(*pd_times*, *ndevs=10*, *cycle=60*, *max_frame_offset=4*)

allensdk.brain_observatory.ecephys.stimulus_sync.**flag_unexpected_edges**(*pd_times*, *ndevs=10*)

allensdk.brain_observatory.ecephys.stimulus_sync.**separate_vsyncs_and_photodiode_times**(*vs_times*, *pd_times*, *photodiode_cycle=60*)

allensdk.brain_observatory.ecephys.stimulus_sync.**trim_border_pulses**(*pd_times*, *vs_times*, *frame_interval=0.016666666666666666*, *num_frames=5*)

allensdk.brain_observatory.ecephys.stimulus_sync.**trim_discontiguous_vsyncs**(*vs_times*, *photodiode_cycle=60*)

allensdk.brain_observatory.ecephys.stimulus_sync.**trimmed_stats**(*data*, *pctiles=(10, 90)*)

allensdk.brain_observatory.ecephys.utils module

allensdk.brain_observatory.ecephys.utils.**clobbering_merge**(*to_df*, *from_df*, ***kwargs*)

allensdk.brain_observatory.ecephys.utils.**group_1d_by_unit**(*data*, *data_unit_map*, *local_to_global_unit_map=None*)

allensdk.brain_observatory.ecephys.utils.**load_and_squeeze_npy**(*path*)

allensdk.brain_observatory.ecephys.utils.**scale_amplitudes**(*spike_amplitudes*, *templates*, *spike_templates*, *scale_factor=1.0*)

allensdk.brain_observatory.ecephys.utils.**strip_substructure_acronym**(*acronym: str | list | float | None*) → str | list | None

Sanitize a structure acronym or a list of structure acronyms by removing the substructure (e.g. DG-mo becomes DG).

If acronym is a list, every element in the list will be sanitized and a list of unique acronyms will be returned. **Element order will not be preserved**.

If acronym is None, return None. If None occurs in a list of sturture acronyms, it will be omitted

Note: if acronym is NaN, it will get converted to None; any other float will provoke an error.

Module contents

allensdk.brain_observatory.ecephys.**get_unit_filter_value**(*key*, *pop=True*, *replace_none=True*, ***source*)

allensdk.brain_observatory.extract_running_speed package

Module contents

allensdk.brain_observatory.gaze_mapping package

Module contents

allensdk.brain_observatory.multi_stimulus_running_speed package

Submodules

allensdk.brain_observatory.multi_stimulus_running_speed.multi_stimulus_running_speed module

Module contents

allensdk.brain_observatory.nwb package

Subpackages

allensdk.brain_observatory.nwb.eye_tracking package

Submodules

allensdk.brain_observatory.nwb.eye_tracking.extension_builder module

allensdk.brain_observatory.nwb.eye_tracking.extension_builder.**main**()

allensdk.brain_observatory.nwb.eye_tracking.ndx_ellipse_eye_tracking module

Module contents

Submodules

allensdk.brain_observatory.nwb.behavior_ophys_nwb_extension_builder module

allensdk.brain_observatory.nwb.metadata module

allensdk.brain_observatory.nwb.metadata.**create_pynwb_extension_from_schemas**(*schema_list*, *prefix: str*, *save_dir: str*)

allensdk.brain_observatory.nwb.metadata.**extract_from_schema**(*schema*)

allensdk.brain_observatory.nwb.metadata.**load_pynwb_extension**(*schema*, *prefix: str*)

allensdk.brain_observatory.nwb.nwb_api module

allensdk.brain_observatory.nwb.nwb_utils module

allensdk.brain_observatory.nwb.schemas module

class allensdk.brain_observatory.nwb.schemas.**RunningSpeedPathsSchema**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: *RaisingSchema*

dump_fields: `Dict[str, ma_fields.Field]`

exclude: `set[Any] | MutableSet[Any]`

fields: `Dict[str, ma_fields.Field]`

Dictionary mapping field_names -> `Field` objects

load_fields: `Dict[str, ma_fields.Field]`

opts: `SchemaOpts = <marshmallow.schema.SchemaOpts object>`

Module contents

allensdk.brain_observatory.nwb.**add_average_image**(*nwbfile*, *average_image*, *image_api=None*)

allensdk.brain_observatory.nwb.**add_cell_specimen_table**(*nwbfile: NWBFile*, *cell_specimen_table: DataFrame*, *session_metadata: dict*)

This function takes the cell specimen table and writes the ROIs contained within. It writes these to a new NWB imaging plane based off the previously supplied metadata

Parameters

nwbfile: NWBFile
this is the in memory NWBFile currently being written to which ROI data is added

cell_specimen_table: pd.DataFrame
this is the DataFrame containing the cells segmented from a ophys experiment, stored in json file and loaded. example: /home/nicholasc/projects/allensdk/allensdk/test/

brain_observatory/behavior/cell_specimen_table_789359614.json

session_metadata: dict
Dictionary containing cell_specimen_table related metadata. Should include at minimum the following fields:

"emission_lambda", "excitation_lambda", "indicator", "targeted_structure", and ophys_frame_rate"

Returns

nwbfile: NWBFile
The altered in memory NWBFile object that now has a specimen table

allensdk.brain_observatory.nwb.**add_corrected_fluorescence_traces**(*nwbfile*, *corrected_fluorescence_traces*)

allensdk.brain_observatory.nwb.**add_dff_traces**(*nwbfile*, *dff_traces*, *ophys_timestamps*)

allensdk.brain_observatory.nwb.**add_eye_gaze_data_interfaces**(*pynwb_container: NWBContainer*, *pupil_areas: Series*, *eye_areas: Series*, *screen_coordinates: DataFrame*, *screen_coordinates_spherical: DataFrame*, *synced_timestamps: Series*) → NWBContainer

allensdk.brain_observatory.nwb.**add_eye_gaze_mapping_data_to_nwbfile**(*nwbfile: NWBFile*, *eye_gaze_data: dict*) → NWBFile

allensdk.brain_observatory.nwb.**add_eye_tracking_ellipse_fit_data_to_nwbfile**(*nwbfile: NWBFile*, *eye_dlc_tracking_data: dict*, *synced_timestamps: Series*) → NWBFile

allensdk.brain_observatory.nwb.**add_image**(*nwbfile*, *image_data*, *image_name*, *module_name*, *module_description*, *image_api=None*)

allensdk.brain_observatory.nwb.**add_invalid_times**(*nwbfile*, *epochs*)

Write invalid times to nwbfile if epochs are not empty Parameters ———- nwbfile: pynwb.NWBFile epochs: list of dicts

records of invalid epochs

Returns

pynwb.NWBFile

allensdk.brain_observatory.nwb.**add_licks**(*nwbfile*, *licks*)

allensdk.brain_observatory.nwb.**add_max_projection**(*nwbfile*, *max_projection*, *image_api=None*)

allensdk.brain_observatory.nwb.**add_metadata**(*nwbfile*, *metadata: dict*, *behavior_only: bool*)

allensdk.brain_observatory.nwb.**add_motion_correction**(*nwbfile*, *motion_correction*)

allensdk.brain_observatory.nwb.**add_rewards**(*nwbfile*, *rewards_df*)

allensdk.brain_observatory.nwb.**add_running_acquisition_to_nwbfile**(*nwbfile*, *running_acquisition_df: DataFrame*)

allensdk.brain_observatory.nwb.**add_running_speed_to_nwbfile**(*nwbfile*, *running_speed*, *name='speed'*, *unit='cm/s'*, *from_dataframe=False*)

Adds running speed data to an NWBFile as a timeseries in acquisition

Parameters

nwbfile
[pynwb.NWBFile] File to which running speeds will be written

running_speed
[Union[RunningSpeed, pd.DataFrame]] Either a RunningSpeed object or pandas DataFrame. Contains attributes 'values' and 'timestamps'

name
[str, optional] Used as name of timeseries object

unit
[str, optional] SI units of running speed values

from_dataframe
[bool, optional] Whether *running_speed* is a dataframe or not. Default is False.

Returns

nwbfile
[pynwb.NWBFile]

allensdk.brain_observatory.nwb.**add_segmentation_mask_image**(*nwbfile*, *segmentation_mask_image*, *image_api=None*)

allensdk.brain_observatory.nwb.**add_stimulus_timestamps**(*nwbfile*, *stimulus_timestamps*, *module_name='stimulus'*)

allensdk.brain_observatory.nwb.**add_task_parameters**(*nwbfile*, *task_parameters*)

allensdk.brain_observatory.nwb.**add_trials**(*nwbfile*, *trials*, *description_dict={}*)

allensdk.brain_observatory.nwb.**check_nwbfile_version**(*nwbfile_path: str*, *desired_minimum_version: str*, *warning_msg: str*)

allensdk.brain_observatory.nwb.**create_eye_gaze_mapping_dataframe**(*eye_gaze_data: dict*) → DataFrame

allensdk.brain_observatory.nwb.**create_eye_tracking_nwb_processing_module**(*eye_dlc_tracking_data: dict*, *synced_timestamps: Series*) → ProcessingModule

allensdk.brain_observatory.nwb.**create_gaze_mapping_nwb_processing_modules**(*eye_gaze_data: dict*)

allensdk.brain_observatory.nwb.**create_stimulus_presentation_time_interval**(*name: str*, *description: str*, *columns_to_add: Iterable*) → TimeIntervals

allensdk.brain_observatory.nwb.**eye_tracking_data_is_valid**(*eye_dlc_tracking_data: dict*, *synced_timestamps: Series*) → bool

allensdk.brain_observatory.nwb.**read_eye_dlc_tracking_ellipses**(*input_path: Path*) → dict

Reads eye tracking ellipse fit data from an h5 file.

Args:

input_path (Path): Path to eye tracking ellipse fit h5 file

Returns:

dict: Loaded h5 data. Each 'params' field contains dataframes with]
ellipse fit parameters. Dataframes contain 5 columns each consisting of: "center_x", "center_y", "height", "phi", "width"

allensdk.brain_observatory.nwb.**read_eye_gaze_mappings**(*input_path: Path*) → dict

Reads eye gaze mapping data from an h5 file.

Args:

input_path (Path): Path to eye gaze mapping h5 data file produced by
'allensdk.brain_observatory.gaze_mapping' module.

Returns:

dict: Loaded h5 data.
*_eye_areas: Area of eye (in pixels^2) over time *_pupil_areas: Area of pupil (in pixels^2) over time *_screen_coordinates: y, x screen coordinates (in cm) over time *_screen_coordinates_spherical: y, x screen coordinates (in deg)

over time

synced_frame_timestamps: synced timestamps for video frames
(in sec)

allensdk.brain_observatory.nwb.**setup_table_for_epochs**(*table*, *timeseries*, *tag*)

allensdk.brain_observatory.nwb.**setup_table_for_invalid_times**(*invalid_epochs*)

Create table with invalid times if invalid_epochs are present

Parameters

invalid_epochs: list of dicts
of invalid epoch records

Returns

pd.DataFrame of invalid times if epochs are not empty,
otherwise return None

allensdk.brain_observatory.ophys package

Subpackages

allensdk.brain_observatory.ophys.trace_extraction package

Module contents

Submodules

allensdk.brain_observatory.ophys.project_constants module

Collection of specific project metadata not readily available on LIMS.

Module contents

allensdk.brain_observatory.receptive_field_analysis package

Submodules

allensdk.brain_observatory.receptive_field_analysis.chisquarerf module

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**NLL_to_pvalue**(*NLLs*, *log_base=10.0*)

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**build_trial_matrix**(*LSN_template*, *num_trials*, *on_off_luminance=(255, 0)*)

Construct indicator arrays for on/off pixels across trials.

Parameters

LSN_template
[np.ndarray] Dimensions are (nTrials, nYPixels, nXPixels). Luminance values per pixel and trial. The size of the first dimension may be larger than the num_trials argument (in which case only the first num_trials slices will be used) but may not be smaller.

num_trials
[int] The number of trials (left-justified) to build indicators for.

on_off_luminance
[array-like, optional] The zeroth element is the luminance value of a pixel when on, the first when off. Defaults are [255, 0].

Returns

trial_mat
[np.ndarray] Dimensions are (nYPixels, nXPixels, {on, off}, nTrials). Boolean values indicate that a pixel was on/off on a particular trial.

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**chi_square_binary**(*events*, *LSN_template*)

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**chi_square_within_mask**(*exclusion_mask*, *events_per_pixel*, *trials_per_pixel*)

Determine if cells respond preferentially to on/off pixels in a mask using a chi2 test.

Parameters

exclusion_mask
[np.ndarray] Dimensions are (nYPixels, nXPixels, {on, off}). Integer indicator for INCLUSION (!) of a pixel within the testing region.

events_per_pixel
[np.ndarray] Dimensions are (nCells, nYPixels, nXPixels, {on, off}). Integer values are response counts by cell to on/off luminance at each pixel.

trials_per_pixel
[np.ndarray] Dimensions are (nYPixels, nXPixels, {on, off}). Integer values are counts of trials where a pixel is on/off.

Returns

p_vals
[np.ndarray] One-dimensional, of length nCells. Float values are p-values for the hypothesis that a given cell has a receptive field within the exclusion mask.

chi
[np.ndarray] Dimensions are (nCells, nYPixels, nXPixels, {on, off}). Values (float) are squared residual event counts divided by expected event counts.

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**deinterpolate_RF**(*rf_map*, *x_pnts*, *y_pnts*, *deg_per_pnt*)

Downsample an image

Parameters

rf_map
[np.ndarray] Input image

x_pnts
[np.ndarray] Count of sample points along the first (column) axis

y_pnts
[np.ndarray] Count of sample points along the zeroth (row) axis

deg_per_pnt
[numeric] scale factor

Returns

sampled_yx
[np.ndarray] Downsampled image

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**get_disc_masks**(*LSN_template*, *radius=3*, *on_luminance=255*, *off_luminance=0*)

Obtain an indicator mask surrounding each pixel. The mask is a square, excluding pixels which are coactive on any trial with the main pixel.

Parameters

LSN_template
[np.ndarray] Dimensions are (nTrials, nYPixels, nXPixels). Luminance values per pixel and trial.

radius
[int] The base mask will be a box whose sides are 2 * radius + 1 in length.

on_luminance
[int, optional] The value of the luminance for on trials. Default is 255

off_luminance
[int, optional] The value of the luminance for off trials. Default is 0

Returns

masks
[np.ndarray] Dimensions are (nYPixels, nXPixels, nYPixels, nXPixels). The first 2 dimensions describe the pixel from which the mask was computed. The last 2 serve as the dimensions of the mask images themselves. Masks are binary arrays of type float, with 1 indicating inside, 0 outside.

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**get_events_per_pixel**(*responses_np*, *trial_matrix*)

Obtain a matrix linking cellular responses to pixel activity.

Parameters

responses_np
[np.ndarray] Dimensions are (nTrials, nCells). Boolean values indicate presence/absence of a response on a given trial.

trial_matrix
[np.ndarray] Dimensions are (nYPixels, nXPixels, {on, off}, nTrials). Boolean values indicate that a pixel was on/off on a particular trial.

Returns

events_per_pixel
[np.ndarray] Dimensions are (nCells, nYPixels, nXPixels, {on, off}). Values for each cell, pixel, and on/off state are the sum of events for that cell across all trials where the pixel was in the on/off state.

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**get_expected_events_by_pixel**(*exclusion_ma*
events_per_p
tri-
als_per_pixel

Calculate expected number of events per pixel

Parameters

exclusion_mask
[np.ndarray] Dimensions are (nYPixels, nXPixels, {on, off}). Integer indicator for INCLUSION (!) of a pixel within the testing region.

events_per_pixel
[np.ndarray] Dimensions are (nCells, nYPixels, nXPixels, {on, off}). Integer values are response counts by cell to on/off luminance at each pixel.

trials_per_pixel
[np.ndarray] Dimensions are (nYPixels, nXPixels, {on, off}). Integer values are counts of trials where a pixel is on/off.

Returns

np.ndarray
Dimensions (nCells, nYPixels, nXPixels, {on, off}). Float values are pixelwise counts of events expected if events are evenly distributed in mask across trials.

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**get_peak_significance**(*chi_square_grid_NLL*, *LSN_template*, *alpha=0.05*)

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**interpolate_RF**(*rf_map*, *deg_per_pnt*)

Upsample an image

Parameters

rf_map
[np.ndarray] Input image

deg_per_pnt
[numeric] scale factor

Returns

interpolated
[np.ndarray] Upsampled image

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**locate_median**(*y*, *x*)

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**pvalue_to_NLL**(*p_values*, *max_NLL=10.0*)

allensdk.brain_observatory.receptive_field_analysis.chisquarerf.**smooth_STA**(*STA*, *gauss_std=0.75*, *total_degrees=64*)

Smooth an image by convolution with a gaussian kernel

Parameters

STA
[np.ndarray] Input image

gauss_std
[numeric, optional] Standard deviation of the gaussian kernel. Will be applied to the upsampled image, so units are visual degrees. Default is 0.75

total_degrees
[int, optional] Size in visual degrees of the input image along its zeroth (row) axis. Used to set the scale factor for up/downsampling.

Returns

STA_smoothed
[np.ndarray] Smoothed image

allensdk.brain_observatory.receptive_field_analysis.eventdetection module

allensdk.brain_observatory.receptive_field_analysis.eventdetection.**detect_events**(*data*, *cell_index*, *stimulus*, *debug_plots=False*)

allensdk.brain_observatory.receptive_field_analysis.fit_parameters module

allensdk.brain_observatory.receptive_field_analysis.fit_parameters.**add_to_fit_parameters_dict_single**(*fit_p*)

allensdk.brain_observatory.receptive_field_analysis.fit_parameters.**compute_distance**(*center_on*, *center_off*)

allensdk.brain_observatory.receptive_field_analysis.fit_parameters.**compute_overlap**(*data_fitted_on*, *data_fitted_off*)

allensdk.brain_observatory.receptive_field_analysis.fit_parameters.**get_gaussian_fit_single_channel**(*rf*, *fit_pa*

allensdk.brain_observatory.receptive_field_analysis.fitgaussian2D module

exception
allensdk.brain_observatory.receptive_field_analysis.fitgaussian2D.**GaussianFitError**

Bases: RuntimeError

allensdk.brain_observatory.receptive_field_analysis.fitgaussian2D.**fitgaussian2D**(*data*)

Fit a 2D gaussian to an image

Parameters

data
[np.ndarray] input image

Returns

p2
[list] height row mean column mean row standard deviation column standard deviation rotation

Notes

see gaussian2D for details about output values

allensdk.brain_observatory.receptive_field_analysis.fitgaussian2D.**gaussian2D**(*height*, *center_x*, *center_y*, *width_x*, *width_y*, *rotation*)

Build a function which evaluates a scaled 2d gaussian pdf

Parameters

height
[float] scale factor

center_x
[float] first coordinate of mean

center_y
[float] second coordinate of mean

width_x
[float] standard deviation along x axis

width_y
[float] standard deviation along y axis

rotation
[float] degrees clockwise by which to rotate the gaussian

Returns

rotgauss: fn
parameters are x and y positions (row/column semantics are set by your inputs to this function). Return value is the scaled gaussian pdf evaluated at the argued point.

allensdk.brain_observatory.receptive_field_analysis.fitgaussian2D.**moments2**(*data*)

Treating input image data as an independent multivariate gaussian, estimate mean and standard deviations

Parameters

data
[np.ndarray] 2d numpy array.

Returns

height
[float] The maximum observed value in the data

y
[float] Mean row index

x
[float] Mean column index

width_y
[float] The standard deviation along the mean row

width_x
[float] The standard deviation along the mean column

None
This function returns an instance of None.

Notes

uses original method from website for finding center

allensdk.brain_observatory.receptive_field_analysis.postprocessing module

allensdk.brain_observatory.receptive_field_analysis.postprocessing.**get_gaussian_fit**(*rf*)

allensdk.brain_observatory.receptive_field_analysis.postprocessing.**run_postprocessing**(*data*, *rf*)

allensdk.brain_observatory.receptive_field_analysis.receptive_field module

allensdk.brain_observatory.receptive_field_analysis.receptive_field.**compute_receptive_field**(*data*, *cell_index*, *stimulus*, ***kwargs*)

allensdk.brain_observatory.receptive_field_analysis.receptive_field.**compute_receptive_field_with_postpr**

allensdk.brain_observatory.receptive_field_analysis.receptive_field.**events_to_pvalues_no_fdr_correction**

allensdk.brain_observatory.receptive_field_analysis.receptive_field.**get_attribute_dict**(*rf*)

allensdk.brain_observatory.receptive_field_analysis.receptive_field.**print_summary**(*rf*)

allensdk.brain_observatory.receptive_field_analysis.receptive_field.**read_h5_group**(*g*)

allensdk.brain_observatory.receptive_field_analysis.receptive_field.**read_receptive_field_from_h5**(*file_nam*
path=N

allensdk.brain_observatory.receptive_field_analysis.receptive_field.**write_receptive_field_to_h5**(*rf*, *file_name*
prefix='')

allensdk.brain_observatory.receptive_field_analysis.tools module

allensdk.brain_observatory.receptive_field_analysis.tools.**dict_generator**(*indict*, *pre=None*)

allensdk.brain_observatory.receptive_field_analysis.tools.**list_of_dicts_to_dict_of_lists**(*list_of_dicts*)

allensdk.brain_observatory.receptive_field_analysis.tools.**read_h5_group**(*g*)

allensdk.brain_observatory.receptive_field_analysis.utilities module

allensdk.brain_observatory.receptive_field_analysis.utilities.**convolve**(*img*, *sigma=4*)

2D Gaussian convolution

allensdk.brain_observatory.receptive_field_analysis.utilities.**get_A**(*data*, *stimulus*)

allensdk.brain_observatory.receptive_field_analysis.utilities.**get_A_blur**(*data*, *stimulus*)

allensdk.brain_observatory.receptive_field_analysis.utilities.**get_attribute_dict**(*rf*)

allensdk.brain_observatory.receptive_field_analysis.utilities.**get_components**(*receptive_field_data*)

allensdk.brain_observatory.receptive_field_analysis.utilities.**get_shuffle_matrix**(*data*, *event_vector*, *A*, *number_of_shuffles=5000*, *response_detection_error_std_*

allensdk.brain_observatory.receptive_field_analysis.utilities.**get_sparse_noise_epoch_mask_list**(*st*, *number_of_acq*, *threshold=7*)

allensdk.brain_observatory.receptive_field_analysis.utilities.**smooth**(*x*, *window_len=11*, *window='hanning'*, *mode='valid'*)

smooth the data using a window with requested size.

This method is based on the convolution of a scaled window with the signal. The signal is prepared by introducing reflected copies of the signal (with the window size) in both ends so that transient parts are minimized in the begining and end part of the output signal.

input:

x: the input signal window_len: the dimension of the smoothing window; should be an odd

integer

window: the type of window from 'flat', 'hanning', 'hamming',

'bartlett', 'blackman' flat window will produce a moving average smoothing.

output:

the smoothed signal

example:

t=linspace(-2,2,0.1) x=sin(t)+randn(len(t))*0.1 y=smooth(x)

see also:

numpy.hanning, numpy.hamming, numpy.bartlett, numpy.blackman, numpy.convolve scipy.signal.lfilter

TODO: the window parameter could be the window itself if an array instead
of a string

NOTE: length(output) != length(input), to correct this:
return y[(window_len/2-1):-(window_len/2)] instead of just y.

allensdk.brain_observatory.receptive_field_analysis.utilities.**upsample_image_to_degrees**(*img*)

allensdk.brain_observatory.receptive_field_analysis.visualization module

allensdk.brain_observatory.receptive_field_analysis.visualization.**plot_chi_square_summary**(*rf_data*, *ax=None*, *cax=None*, *cmap='magma'*)

allensdk.brain_observatory.receptive_field_analysis.visualization.**plot_ellipses**(*gaussian_fit_dict*, *ax=None*, *show=True*, *close=True*, *save_file_name=None*, *color='b'*)

Example Usage: oeid, cell_index, stimulus = 512176430, 12, 'locally_sparse_noise' brain_observatory_cache = BrainObservatoryCache() data_set = brain_observatory_cache.get_ophys_experiment_data(oeid) lsn = LocallySparseNoise(data_set, stimulus) result = compute_receptive_field_with_postprocessing(data_set, cell_index, stimulus, alpha=.05, number_of_shuffles=5000) plot_ellipses(result['off']['gaussian_fit'], color='r')

allensdk.brain_observatory.receptive_field_analysis.visualization.**plot_fields**(*on_data*, *off_data*, *on_axes*, *off_axes*, *cbar_axes=None*, *clim=None*, *cmap='magma'*)

allensdk.brain_observatory.receptive_field_analysis.visualization.**plot_gaussian_fit**(*rf_data*, *ax_on*, *ax_off*, *ax_cbar=None*, *cmap='magma'*)

allensdk.brain_observatory.receptive_field_analysis.visualization.**plot_mask**(*rf_data*, *ax_on*, *ax_off*, *ax_cbar=None*, *cmap='magma'*)

allensdk.brain_observatory.receptive_field_analysis.visualization.**plot_msr_summary**(*lsn*, *cell_index*, *ax_on*, *ax_off*, *ax_cbar=None*, *cmap=None*)

allensdk.brain_observatory.receptive_field_analysis.visualization.**plot_p_values**(*rf_data*, *ax_on*, *ax_off*, *ax_cbar=None*, *cmap='magma'*)

allensdk.brain_observatory.receptive_field_analysis.visualization.**plot_receptive_field_data**(*rf*, *lsn*, *show=True*, *save_file_name*, *close=True*, *cmap='magma*

allensdk.brain_observatory.receptive_field_analysis.visualization.**plot_rts_blur_summary**(*rf_data*, *ax_on*, *ax_off*, *ax_cbar=None*, *cmap='magma'*)

allensdk.brain_observatory.receptive_field_analysis.visualization.**plot_rts_summary**(*rf_data*, *ax_on*, *ax_off*, *ax_cbar=None*, *cmap='magma'*)

allensdk.brain_observatory.receptive_field_analysis.visualization.**pvalue_to_NLL**(*p_values*, *max_NLL=10.0*)

Module contents

allensdk.brain_observatory.sync_utilities package

Module contents

allensdk.brain_observatory.sync_utilities.**get_synchronized_frame_times**(*session_sync_file: Path*, *sync_line_label_keys: Tuple[str, ...]*, *drop_frames: List[int] | None = None*, *trim_after_spike: bool = True*) → Series

Get experimental frame times from an experiment session sync file.

1. Get rising edges from the sync dataset
2. **Occasionally an extra set of frame times are acquired after the rest of** the signals. These are manifested by a discontiguous time sequence. We detect and remove these.

3. Remove dropped frames

Parameters

session_sync_file
[Path] Path to an ephys session sync file. The sync file contains rising/falling edges from a daq system which indicates when certain events occur (so they can be related to each other).

sync_line_label_keys
[Tuple[str, ...]] Line label keys to get times for. See class attributes of allensdk.brain_observatory.sync_dataset.Dataset for a listing of possible keys.

drop_frames
[List] frame indices to be removed from frame times

trim_after_spike
[bool = True] If True, will call trim_discontiguous_times on the frame times before returning them, which will detect any spikes in the data and remove all elements for the list which come after the spike.

Returns

pd.Series
An array of times when eye tracking frames were acquired.

allensdk.brain_observatory.sync_utilities.**trim_discontiguous_times**(*times: ndarray*, *threshold=100*) → ndarray

If the time sequence is discontigous, detect the first instance occurance and trim off the tail of the sequence

Parameters

times
[frame times]

Returns

trimmed frame times

allensdk.brain_observatory.vbn_2022 package

Subpackages

allensdk.brain_observatory.vbn_2022.input_json_writer package

Submodules

allensdk.brain_observatory.vbn_2022.input_json_writer.input_json_writer module

allensdk.brain_observatory.vbn_2022.input_json_writer.schemas module

class allensdk.brain_observatory.vbn_2022.input_json_writer.schemas.**VBN2022InputJsonWriterSchema**(*, *only: types.St | None = None*, *exclude: types.St = ()*, *many: bool = False*, *context: dict | None = None*, *load_on types.St = ()*, *dump_o types.St = ()*, *partial: bool | types.St | None = None*, *unknown: str | None = None*)

Bases: ArgSchema

create_path_lookup(*data*, ***kwargs*)

Construct lookups mapping ecephys_session_id to the input_json_path and the nwb_file_path

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

allensdk.brain_observatory.vbn_2022.input_json_writer.utils module

Module contents

allensdk.brain_observatory.vbn_2022.metadata_writer package

Submodules

allensdk.brain_observatory.vbn_2022.metadata_writer.dataframe_manipulations module

allensdk.brain_observatory.vbn_2022.metadata_writer.lims_queries module

allensdk.brain_observatory.vbn_2022.metadata_writer.metadata_writer module

allensdk.brain_observatory.vbn_2022.metadata_writer.schemas module

Module contents

allensdk.brain_observatory.vbn_2022.utils package

Submodules

allensdk.brain_observatory.vbn_2022.utils.schemas module

class allensdk.brain_observatory.vbn_2022.utils.schemas.**ProbeToSkip**(*, *only: types.StrSequenceOrSet | None = None*, *exclude: types.StrSequenceOrSet = ()*, *many: bool = False*, *context: dict | None = None*, *load_only: types.StrSequenceOrSet = ()*, *dump_only: types.StrSequenceOrSet = ()*, *partial: bool | types.StrSequenceOrSet | None = None*, *unknown: str | None = None*)

Bases: ArgSchema

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

validate_probe_names(*data*, ***kwargs*)

Module contents

Module contents

allensdk.brain_observatory.visualization package

Module contents

allensdk.brain_observatory.visualization.**plot_running_speed**(*timestamps*, *values*, *start_index=0*, *stop_index=None*, *step=1*, *ylabel='running speed (cm/s)'*, *xlabel='time (s)'*, *title=None*)

Make a simple plot of a running speed trace

Parameters

timestamps
[numpy.ndarray] Times at which running speed samples were collected

values
[numpy.ndarray] Running speed values (by default: linear cm / s with negative values indicating backwards movement)

Submodules

allensdk.brain_observatory.argschema_utilities module

class allensdk.brain_observatory.argschema_utilities.**ArgSchemaParserPlus**(**args*, ***kwargs*)

Bases: ArgSchemaParser

class allensdk.brain_observatory.argschema_utilities.**InputFile**(**, load_default: typing.Any = <marshmallow.missing>, missing: typing.Any = <marshmallow.missing>, dump_default: typing.Any = <marshmallow.missing>, default: typing.Any = <marshmallow.missing>, data_key: str | None = None, attribute: str | None = None, validate: None | typing.Callable[[typing.Any], typing.Any] | typing.Iterable[typing.Callable[[typing.Any], typing.Any]] = None, required: bool = False, allow_none: bool | None = None, load_only: bool = False, dump_only: bool = False, error_messages: dict[str, str] | None = None, metadata: typing.Mapping[str, typing.Any] | None = None, **additional_metadata*)

Bases: String

A marshmallow String field subclass which deserializes json str fields that represent a desired input path to pathlib.Path. Also performs read access checking.

class allensdk.brain_observatory.argschema_utilities.**OutputFile**(**, load_default: typing.Any = <marshmallow.missing>, missing: typing.Any = <marshmallow.missing>, dump_default: typing.Any = <marshmallow.missing>, default: typing.Any = <marshmallow.missing>, data_key: str | None = None, attribute: str | None = None, validate: None | typing.Callable[[typing.Any], typing.Any] | typing.Iterable[typing.Callable[[typing.Any], typing.Any]] = None, required: bool = False, allow_none: bool | None = None, load_only: bool = False, dump_only: bool = False, error_messages: dict[str, str] | None = None, metadata: typing.Mapping[str, typing.Any] | None = None, **additional_metadata*)

Bases: String

A marshmallow String field subclass which deserializes json str fields that represent a desired output file path to a pathlib.Path. Also performs write access checking.

class allensdk.brain_observatory.argschema_utilities.**RaisingSchema**(**, only: types.StrSequenceOrSet | None = None, exclude: types.StrSequenceOrSet = (), many: bool = False, context: dict | None = None, load_only: types.StrSequenceOrSet = (), dump_only: types.StrSequenceOrSet = (), partial: bool | types.StrSequenceOrSet | None = None, unknown: str | None = None*)

Bases: DefaultSchema

class **Meta**

Bases: object

unknown = 'raise'

opts: SchemaOpts = <marshmallow.schema.SchemaOpts object>

allensdk.brain_observatory.argschema_utilities.**check_read_access**(*path*)

allensdk.brain_observatory.argschema_utilities.**check_write_access**(*filepath*, *allow_exists=False*)

allensdk.brain_observatory.argschema_utilities.**check_write_access_dir**(*dirpath*)

allensdk.brain_observatory.argschema_utilities.**check_write_access_overwrite**(*path*)

allensdk.brain_observatory.argschema_utilities.**optional_lims_inputs**(*argv*, *input_schema*, *output_schema*, *lims_input_getter*)

allensdk.brain_observatory.argschema_utilities.**write_or_print_outputs**(*data*, *parser*)

allensdk.brain_observatory.brain_observatory_exceptions module

exception
allensdk.brain_observatory.brain_observatory_exceptions.**BrainObservatoryAnalysisException**

Bases: Exception

exception allensdk.brain_observatory.brain_observatory_exceptions.**EpochSeparationException**(**args*, ***kwargs*)

Bases: Exception

exception
allensdk.brain_observatory.brain_observatory_exceptions.**MissingStimulusException**

Bases: Exception

exception allensdk.brain_observatory.brain_observatory_exceptions.**NoEyeTrackingException**

Bases: Exception

allensdk.brain_observatory.brain_observatory_plotting module

allensdk.brain_observatory.brain_observatory_plotting.**plot_drifting_grating_traces**(*dg*, *save_dir*)

saves figures with a Ori X TF grid of mean resposes

allensdk.brain_observatory.brain_observatory_plotting.**plot_lsn_traces**(*lsn*, *save_dir*, *suffix=''*)

allensdk.brain_observatory.brain_observatory_plotting.**plot_ns_traces**(*nsa*, *save_dir*)

allensdk.brain_observatory.brain_observatory_plotting.**plot_running_a**(*dg*, *nm1*, *nm3*, *save_dir*)

allensdk.brain_observatory.brain_observatory_plotting.**plot_sg_traces**(*sg*, *save_dir*)

allensdk.brain_observatory.chisquare_categorical module

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@author: dan

allensdk.brain_observatory.chisquare_categorical.**advance_combination**(*curr_combination*, *options_per_column*)

allensdk.brain_observatory.chisquare_categorical.**chisq_from_stim_table**(*stim_table*, *columns*, *mean_sweep_events*, *num_shuffles=1000*, *verbose=False*)

allensdk.brain_observatory.chisquare_categorical.**compute_chi**(*observed*, *expected*)

allensdk.brain_observatory.chisquare_categorical.**compute_chi_shuffle**(*mean_sweep_events*, *sweep_categories*, *num_shuffles=1000*)

allensdk.brain_observatory.chisquare_categorical.**compute_expected**(*mean_sweep_events*, *sweep_conditions*)

allensdk.brain_observatory.chisquare_categorical.**compute_observed**(*mean_sweep_events*, *sweep_conditions*)

allensdk.brain_observatory.chisquare_categorical.**make_category_dummy**(*sweep_categories*)

allensdk.brain_observatory.chisquare_categorical.**stim_table_to_categories**(*stim_table*, *columns*, *verbose=False*)

allensdk.brain_observatory.circle_plots module

class allensdk.brain_observatory.circle_plots.**CoronaPlotter**(*angle_start=270*, *plot_scale=1.2*, *inner_radius=0.3*, **args*, ***kwargs*)

Bases: *PolarPlotter*

infer_dims(*category_data*)

plot(*category_data*, *data=None*, *clim=None*, *cmap=<matplotlib.colors.LinearSegmentedColormap object>*)

set_dims(*categories*)

show_arrow(*color=None*)

show_circle(*color=None*)

class allensdk.brain_observatory.circle_plots.**FanPlotter**(*group_scale=0.9*, **args*, ***kwargs*)

Bases: *PolarPlotter*

static for_drifting_gratings()

static for_static_gratings()

infer_dims(*r_data*, *angle_data*, *group_data*)

plot(*r_data*, *angle_data*, *group_data=None*, *data=None*, *cmap=<matplotlib.colors.LinearSegmentedColormap object>*, *clim=None*, *rmap=None*, *rlim=None*, *axis_color=None*, *label_color=None*)

set_dims(*rs*, *angles*, *groups*)

show_angle_labels(*angles=None*, *labels=None*, *color=None*, *offset=0.05*, *fontdict=None*)

show_axes(*angles=None*, *radii=None*, *closed=False*, *color=None*)

show_group_labels(*groups=None*, *color=None*, *fontdict=None*)

show_r_labels(*radii=None*, *labels=None*, *color=None*, *offset=0.1*, *fontdict=None*)

class allensdk.brain_observatory.circle_plots.**PolarPlotter**(*direction=-1*, *angle_start=0*, *circle_scale=1.1*, *inner_radius=None*, *plot_center=(0.0, 0.0)*, *plot_scale=0.9*)

Bases: `object`

DIR_CCW = 1

DIR_CW = -1

finalize()

class allensdk.brain_observatory.circle_plots.**TrackPlotter**(*direction=-1*, *angle_start=270.0*, *inner_radius=0.45*, *ring_length=None*, **args*, ***kwargs*)

Bases: *PolarPlotter*

plot(*data*, *clim=None*, *cmap=<matplotlib.colors.LinearSegmentedColormap object>*, *mean_cmap=<matplotlib.colors.LinearSegmentedColormap object>*, *norm=None*)

show_arrow(*color=None*)

allensdk.brain_observatory.circle_plots.**add_angle_labels**(*ax*, *angles*, *labels*, *radius*, *color=None*, *fontdict=None*, *offset=0.05*)

allensdk.brain_observatory.circle_plots.**add_arrow**(*ax*, *radius*, *start_angle*, *end_angle*, *color=None*, *width=18.0*)

allensdk.brain_observatory.circle_plots.**angle_lines**(*angles*, *inner_radius*, *outer_radius*)

allensdk.brain_observatory.circle_plots.**build_hex_pack**(*n*)

allensdk.brain_observatory.circle_plots.**hex_pack**(*radius*, *n*)

allensdk.brain_observatory.circle_plots.**make_pincushion_plot**(*data*, *trials*, *on*, *nrows*, *ncols*, *clim=None*, *color_map=None*, *radius=None*)

allensdk.brain_observatory.circle_plots.**polar_line_circles**(*radii*, *theta*, *start_r=0*)

allensdk.brain_observatory.circle_plots.**polar_linspace**(*radius*, *start_angle*, *stop_angle*, *num*, *endpoint=False*, *degrees=True*)

Evenly distributed list of x,y coordinates from an input range of angles and a radius in polar coordinates.

allensdk.brain_observatory.circle_plots.**polar_to_xy**(*angles*, *radius*)

Convert an array of angles (in radians) and a radius in polar coordinates to an array of x,y coordinates.

allensdk.brain_observatory.circle_plots.**radial_arcs**(*rs*, *start_theta*, *end_theta*)

allensdk.brain_observatory.circle_plots.**radial_circles**(*rs*)

allensdk.brain_observatory.circle_plots.**reset_hex_pack**()

allensdk.brain_observatory.circle_plots.**rings_in_hex_pack**(*ct*)

allensdk.brain_observatory.circle_plots.**spiral_trials**(*radii*, *x=0.0*, *y=0.0*)

allensdk.brain_observatory.circle_plots.**spiral_trials_polar**(*r*, *theta*, *radii*, *offset=None*)

allensdk.brain_observatory.circle_plots.**wedge_ring**(*N*, *inner_radius*, *outer_radius*, *start=0*, *stop=360*)

allensdk.brain_observatory.comparison_utils module

allensdk.brain_observatory.comparison_utils.**compare_fields**(*x1: Any*, *x2: Any*, *err_msg=''*, *ignore_keys: Set[str] | None = None*)

Helper function to compare if two fields (attributes) are equal to one another.

Parameters

x1
[Any] The first field

x2
[Any] The other field

err_msg
[str, optional] The error message to display if two compared fields do not equal one another, by default "" (an empty string)

ignore_keys
For dictionary comparison, ignore these keys

allensdk.brain_observatory.demixer module

allensdk.brain_observatory.demixer.**demix_time_dep_masks**(*raw_traces: ndarray*, *stack: ndarray*, *masks: ndarray*, *max_block_size: int = 1000*) → Tuple[ndarray, list]

Demix traces of potentially overlapping masks extraced from a single 2p recording.

Parameters

- **raw_traces** – 2d array of traces for each mask, of dimensions (n, t), where *t* is the number of time points and *n* is the number of masks.
- **stack** – 3d array representing a 1p recording movie, of dimensions (t, H, W) or corresponding hdf5 dataset.
- **masks** – 3d array of binary roi masks, of shape (n, H, W), where *n* is the number of masks, and HW are the dimensions of an individual frame in the movie *stack*.

Max_block_size
int representing maximum number of movie frames to read at a time (-1 for full length *t* of *stack*) (the default is 1000)

Returns
Tuple of demixed traces and whether each frame was skipped in the demixing calculation.

allensdk.brain_observatory.demixer.**find_negative_baselines**(*trace*)

allensdk.brain_observatory.demixer.**find_negative_transients_threshold**(*trace*, *window=500*, *length=10*, *std_devs=3*)

allensdk.brain_observatory.demixer.**find_zero_baselines**(*traces*)

allensdk.brain_observatory.demixer.**identify_valid_masks**(*mask_array*)

allensdk.brain_observatory.demixer.**plot_negative_baselines**(*raw_traces*, *demix_traces*, *mask_array*, *roi_ids_mask*, *plot_dir*, *ext='png'*)

allensdk.brain_observatory.demixer.**plot_negative_transients**(*raw_traces*, *demix_traces*, *valid_roi*, *mask_array*, *roi_ids_mask*, *plot_dir*, *ext='png'*)

allensdk.brain_observatory.demixer.**plot_overlap_masks_lengthOne**(*roi_ind*, *masks*, *savefile=None*, *weighted=False*)

allensdk.brain_observatory.demixer.**plot_traces**(*raw_trace*, *demix_trace*, *roi_id*, *roi_ind*, *save_file*)

allensdk.brain_observatory.demixer.**plot_transients**(*roi_ind*, *t_trans*, *masks*, *traces*, *demix_traces*, *savefile*)

allensdk.brain_observatory.demixer.**rolling_window**(*trace*, *window=500*)

Parameters

- **trace** –
- **window** –

Returns

allensdk.brain_observatory.dff module

allensdk.brain_observatory.dff.**calculate_dff**(*traces*, *dff_computation_cb=None*, *save_plot_dir=None*)

Apply dF/F computation to a set of traces.

The default computation method is *compute_dff_windowed_median()* using default window parameters.

Parameters

traces
[np.ndarray] 2D array of traces to be analyzed.

dff_computation_cb
[function] Function that takes traces as an argument and returns an array of the same shape that is the calculated dF/F.

save_plot_dir
[str] Directory to save dF/F plots to. By default no plots are saved.

Returns

dff
[np.ndarray] 2D array of dF/F traces.

allensdk.brain_observatory.dff.**compute_dff_windowed_median**(*traces*, *median_kernel_long=5401*, *median_kernel_short=101*, *noise_stds=None*, *n_small_baseline_frames=None*, ***kwargs*)

Compute dF/F of a set of traces with median filter detrending.

The operation is basically:

T_long = windowed_median(T) # long timescale kernel

T_dff1 = (T - T_long) / elementwise_max(T_long, noise_std(T))

T_short = windowed_median(T_dff1) # short timescale kernel

T_dff = T_dff1 - elementwise_min(T_short, 2.5*noise_std(T_dff1))

Parameters

traces
[np.ndarray] 2D array of traces to be analyzed.

median_kernel_long
[int] Window size to use for long timescale median detrending.

median_kernel_short
[int] Window size to use for short timescale median detrending.

noise_stds
[list] List that will contain noise_std(T_dff1) for each trace. The value for each trace will be appended to the list if provided.

n_small_baseline_frames
[list] List that will contain the number of frames for each trace where the long-timescale median window is less than noise_std(T). The value for each trace will be appended to the list if provided.

kwargs:
Additional keyword arguments are passed to *noise_std()* .

Returns

dff
[np.ndarray] 2D array of dF/F traces.

allensdk.brain_observatory.dff.**compute_dff_windowed_mode**(*traces*, *mode_kernelsize=5400*, *mean_kernelsize=3000*)

Compute dF/F of a set of traces using a low-pass windowed-mode operator.

The operation is basically:

T_mm = windowed_mean(windowed_mode(T))

T_dff = (T - T_mm) / T_mm

Parameters

traces
[np.ndarray] 2D array of traces to be analyzed.

mode_kernelsize
[int] Window size to use for windowed_mode.

mean_kernelsize
[int] Window size to use for windowed_mean.

Returns

dff
[np.ndarray] 2D array of dF/F traces.

allensdk.brain_observatory.dff.**main**()

allensdk.brain_observatory.dff.**movingaverage**(*x*, *kernelsize*, *y*)

Compute the windowed average of an array.

Parameters

x
[np.ndarray] Array to be analyzed

kernelsize
[int] Size of the moving window

y
[np.ndarray] Output array to store the results

allensdk.brain_observatory.dff.**movingmode_fast**(*x*, *kernelsize*, *y*)

Compute the windowed mode of an array. A running mode is initialized with a histogram of values over the initial kernelsize/2 values. The mode is then updated as the kernel moves by adding and subtracting values from the histogram.

Parameters

x
[np.ndarray] Array to be analyzed

kernelsize
[int] Size of the moving window

y
[np.ndarray] Output array to store the results

allensdk.brain_observatory.dff.**noise_std**(*x*, *noise_kernel_length=31*, *positive_peak_scale=1.5*, *outlier_std_scale=2.5*)

Robust estimate of the standard deviation of the trace noise.

allensdk.brain_observatory.dff.**plot_onetrace**(*dff*, *fc*)

Debug plotting function

allensdk.brain_observatory.dff.**robust_std**(*x*)

Robust estimate of standard deviation.

Estimate of the standard deviation using the median absolute deviation of x.

allensdk.brain_observatory.drifting_gratings module

class allensdk.brain_observatory.drifting_gratings.**DriftingGratings**(*data_set*, ***kwargs*)

Bases: *StimulusAnalysis*

Perform tuning analysis specific to drifting gratings stimulus.

Parameters

data_set: BrainObservatoryNwbDataSet object

static from_analysis_file(*data_set*, *analysis_file*)

get_noise_correlation(*corr='spearman'*)

get_peak()

Computes metrics related to each cell's peak response condition.

Returns

Pandas data frame containing the following columns (_dg suffix is for drifting grating):

- ori_dg (orientation)
- tf_dg (temporal frequency)
- reliability_dg
- osi_dg (orientation selectivity index)
- dsi_dg (direction selectivity index)
- peak_dff_dg (peak dF/F)
- ptest_dg
- p_run_dg
- run_modulation_dg
- cv_dg (circular variance)

get_representational_similarity(*corr='spearman'*)

get_response()

Computes the mean response for each cell to each stimulus condition. Return is a (# orientations, # temporal frequencies, # cells, 3) np.ndarray. The final dimension contains the mean response to the condition (index 0), standard error of the mean of the response to the condition (index 1), and the number of trials with a significant response ($p < 0.05$) to that condition (index 2).

Returns

Numpy array storing mean responses.

get_signal_correlation(*corr='spearman'*)

property number_ori

property number_tf

open_star_plot(*cell_specimen_id=None*, *include_labels=False*, *cell_index=None*)

property orivals

plot_direction_selectivity(*si_range=[0, 1.5]*, *n_hist_bins=50*, *color='#ccccdd'*, *p_value_max=0.05*, *peak_dff_min=3*)

plot_orientation_selectivity(*si_range=[0, 1.5]*, *n_hist_bins=50*, *color='#ccccdd'*, *p_value_max=0.05*, *peak_dff_min=3*)

plot_preferred_direction(*include_labels=False*, *si_range=[0, 1.5]*, *color='#ccccdd'*, *p_value_max=0.05*, *peak_dff_min=3*)

plot_preferred_temporal_frequency(*si_range=[0, 1.5]*, *color='#ccccdd'*, *p_value_max=0.05*, *peak_dff_min=3*)

populate_stimulus_table()

Implemented by subclasses.

reshape_response_array()

Returns
response array in cells x stim x repetition for noise

correlations

property tfvals

allensdk.brain_observatory.findlevel module

allensdk.brain_observatory.findlevel.**findlevel**(*inwave*, *threshold*, *direction='both'*)

allensdk.brain_observatory.locally_sparse_noise module

class allensdk.brain_observatory.locally_sparse_noise.**LocallySparseNoise**(*data_set*, *stimulus=None*, ***kwargs*)

Bases: *StimulusAnalysis*

Perform tuning analysis specific to the locally sparse noise stimulus.

Parameters

data_set: BrainObservatoryNwbDataSet object
stimulus: string
Name of locally sparse noise stimulus. See brain_observatory.stimulus_info.

nrows: int
Number of rows in the stimulus template

ncol: int
Number of columns in the stimulus template

property LSN

LSN_GREY = 127

LSN_OFF = 0

LSN_OFF_SCREEN = 64

LSN_ON = 255

property LSN_mask

property cell_index_receptive_field_analysis_data

property extralength

static from_analysis_file(*data_set*, *analysis_file*, *stimulus*)

get_mean_response()

get_peak()

Implemented by subclasses.

get_receptive_field()

Calculates receptive fields for each cell

get_receptive_field_analysis_data()

Calculates receptive fields for each cell

get_receptive_field_attribute_df()

property interlength

property mean_response

static merge_mean_response(*rc1*, *rc2*)

Move out of this class, to session analysis

open_pincushion_plot(*on*, *cell_specimen_id=None*, *color_map=None*, *cell_index=None*)

plot_cell_receptive_field(*on*, *cell_specimen_id=None*, *color_map=None*, *clim=None*, *mask=None*, *cell_index=None*, *scalebar=True*)

plot_population_receptive_field(*color_map='RdPu'*, *clim=None*, *mask=None*, *scalebar=True*)

plot_receptive_field_analysis_data(*cell_index*, ***kwargs*)

populate_stimulus_table()

Implemented by subclasses.

static read_cell_index_receptive_field_analysis(*file_handle*, *prefix*, *path=None*)

property receptive_field

static save_cell_index_receptive_field_analysis(*cell_index_receptive_field_analysis_data*, *new_nwb*, *prefix*)

sort_trials()

property sweeplength

allensdk.brain_observatory.natural_movie module

class allensdk.brain_observatory.natural_movie.**NaturalMovie**(*data_set*, *movie_name*, ***kwargs*)

Bases: *StimulusAnalysis*

Perform tuning analysis specific to natural movie stimulus.

Parameters

data_set: BrainObservatoryNwbDataSet object
movie_name: string

one of [stimulus_info.NATURAL_MOVIE_ONE,
stimulus_info.NATURAL_MOVIE_TWO, stimulus_info.NATURAL_MOVIE_THREE
]

static from_analysis_file(*data_set*, *analysis_file*, *movie_name*)

get_peak()

Computes properties of the peak response condition for each cell.

Returns

Pandas data frame with the below fields. A suffix of "nm1", "nm2" or "nm3" is appended to the field name depending on which of three movie clips was presented.

- peak_nm1 (frame with peak response)
- response_variability_nm1

get_sweep_response()

Returns the dF/F response for each cell

Returns

Numpy array

open_track_plot(*cell_specimen_id=None*, *cell_index=None*)

populate_stimulus_table()

Implemented by subclasses.

property sweep_response

property sweeplength

allensdk.brain_observatory.natural_scenes module

class allensdk.brain_observatory.natural_scenes.**NaturalScenes**(*data_set*, ***kwargs*)

Bases: *StimulusAnalysis*

Perform tuning analysis specific to natural scenes stimulus.

Parameters

data_set: BrainObservatoryNwbDataSet object

property extralength

static from_analysis_file(*data_set*, *analysis_file*)

get_noise_correlation(*corr='spearman'*)

get_peak()

Computes metrics about peak response condition for each cell.

Returns

Pandas data frame with the following fields ('_ns' suffix is for natural scene):

- scene_ns (scene number)
- reliability_ns
- peak_dff_ns (peak dF/F)
- ptest_ns
- p_run_ns
- run_modulation_ns
- time_to_peak_ns

get_representational_similarity(*corr='spearman'*)

get_response()

Computes the mean response for each cell to each stimulus condition. Return is a (# scenes, # cells, 3) np.ndarray. The final dimension contains the mean response to the condition (index 0), standard error of the mean of the response to the condition (index 1), and the number of trials with a significant ($p < 0.05$) response to that condition (index 2).

Returns

Numpy array storing mean responses.

get_signal_correlation(*corr='spearman'*)

property interlength

property number_scenes

open_corona_plot(*cell_specimen_id=None*, *cell_index=None*)

plot_time_to_peak(*p_value_max=0.05*, *color_map=<matplotlib.colors.LinearSegmentedColormap object>*)

populate_stimulus_table()

Implemented by subclasses.

reshape_response_array()

Returns

response array in cells x stim x repetition for noise

correlations

property sweeplength

allensdk.brain_observatory.observatory_plots module

class allensdk.brain_observatory.observatory_plots.**DimensionPatchHandler**(*vals*, *start_color*, *end_color*, **args*, ***kwargs*)

Bases: object

dim_color(*index*)

legend_artist(*legend*, *orig_handle*, *fontsize*, *handlebox*)

allensdk.brain_observatory.observatory_plots.**figure_in_px**(*w*, *h*, *file_name*, *dpi=96.0*, *transparent=False*)

allensdk.brain_observatory.observatory_plots.**finalize_no_axes**(*pad=0.0*)

allensdk.brain_observatory.observatory_plots.**finalize_no_labels**(*pad=0.3*, *legend=False*)

allensdk.brain_observatory.observatory_plots.**finalize_with_axes**(*pad=0.3*)

allensdk.brain_observatory.observatory_plots.**float_label**(*n*)

allensdk.brain_observatory.observatory_plots.**plot_cell_correlation**(*sig_corrs*, *labels*, *colors*, *scale=15*)

allensdk.brain_observatory.observatory_plots.**plot_combined_speed**(*binned_resp_vis*, *binned_dx_vis*, *binned_resp_sp*, *binned_dx_sp*, *evoked_color*, *spont_color*)

allensdk.brain_observatory.observatory_plots.**plot_condition_histogram**(*vals*, *bins*, *color='#ccccdd'*)

allensdk.brain_observatory.observatory_plots.**plot_mask_outline**(*mask*, *ax*, *color='k'*)

allensdk.brain_observatory.observatory_plots.**plot_pupil_location**(*xy_deg*, *s=1*, *c=None*, *cmap=<matplotlib.colors.LinearSegmentedColor object>*, *edgecolor=''*, *include_labels=True*)

allensdk.brain_observatory.observatory_plots.**plot_radial_histogram**(*angles*, *counts*, *all_angles=None*, *include_labels=False*, *offset=180.0*, *direction=-1*, *closed=False*, *color='#ccccdd'*)

allensdk.brain_observatory.observatory_plots.**plot_receptive_field**(*rf*, *color_map=None*, *clim=None*, *mask=None*, *outline_color='#cccccc'*, *scalebar=True*)

allensdk.brain_observatory.observatory_plots.**plot_representational_similarity**(*rs*, *dims=None*, *dim_labels=None*, *colors=None*, *dim_order=None*, *labels=True*)

allensdk.brain_observatory.observatory_plots.**plot_selectivity_cumulative_histogram**(*sis*, *xlabel*, *si_range=[0, 1.5]*, *n_hist_bins=50*, *color='#ccccdd'*)

allensdk.brain_observatory.observatory_plots.**plot_speed**(*binned_resp*, *binned_dx*, *num_bins*, *color*)

allensdk.brain_observatory.observatory_plots.**plot_time_to_peak**(*msrs*, *ttps*, *t_start*, *t_end*, *stim_start*, *stim_end*, *cmap*)

allensdk.brain_observatory.observatory_plots.**population_correlation_scatter**(*sig_corrs*, *noise_corrs*, *labels*, *colors*, *scale=15*)

allensdk.brain_observatory.r_neuropil module

class allensdk.brain_observatory.r_neuropil.**NeuropilSubtract**(*lam=0.05*, *dt=1.0*, *folds=4*)

Bases: object

TODO: docs

estimate_error(*r*)

Estimate error values for a given r for each fold and return the mean.

fit(*r_range=[0.0, 2.0]*, *iterations=3*, *dr=0.1*, *dr_factor=0.1*)

Estimate error values for a range of r values. Identify a new r range around the minimum error values and repeat multiple times. TODO: docs

fit_block_coordinate_desc(*r_init=5.0*, *min_delta_r=1e-08*)

set_F(*F_M*, *F_N*)

Break the F_M and F_N traces into the number of folds specified in the class constructor and normalize each fold of F_M and R_N relative to F_N.

allensdk.brain_observatory.r_neuropil.**ab_from_T**(*T*, *lam*, *dt*)

allensdk.brain_observatory.r_neuropil.**ab_from_diagonals**(*mat_dict*)

Constructs value for scipy.linalg.solve_banded

Parameters

mat_dict: dictionary of diagonals keyed by offsets

Returns

ab: value for scipy.linalg.solve_banded

allensdk.brain_observatory.r_neuropil.**alpha_filter**(*A=1.0*, *alpha=0.05*, *beta=0.25*, *T=100*)

allensdk.brain_observatory.r_neuropil.**error_calc**(*F_M*, *F_N*, *F_C*, *r*)

allensdk.brain_observatory.r_neuropil.**error_calc_outlier**(*F_M*, *F_N*, *F_C*, *r*)

allensdk.brain_observatory.r_neuropil.**estimate_contamination_ratios**(*F_M*, *F_N*, *lam=0.05*, *folds=4*, *iterations=3*, *r_range=[0.0, 2.0]*, *dr=0.1*, *dr_factor=0.1*)

Calculates neuropil contamination of ROI

Parameters

F_M: ROI trace
F_N: Neuropil trace

Returns

dictionary: key-value pairs

- 'r': the contamination ratio – corrected trace = M - r*N
- 'err': RMS error
- 'min_error': minimum error
- 'bounds_error': boolean. True if error or R are outside tolerance

allensdk.brain_observatory.r_neuropil.**get_diagonals_from_sparse**(*mat*)

Returns a dictionary of diagonals keyed by offsets

Parameters

mat: scipy.sparse matrix

Returns

dictionary: diagonals keyed by offsets

allensdk.brain_observatory.r_neuropil.**normalize_F**(*F_M*, *F_N*)

allensdk.brain_observatory.r_neuropil.**synthesize_F**(*T*, *af1*, *af2*, *p1=0.05*, *p2=0.1*)

Build a synthetic F_C, F_M, F_N, and r of length T TODO: docs

allensdk.brain_observatory.r_neuropil.**validate_with_synthetic_F**(*T*, *N*)

Compute N synthetic traces of length T with known values of r, then estimate r. TODO: docs

allensdk.brain_observatory.roi_masks module

class allensdk.brain_observatory.roi_masks.**Mask**(*image_w*, *image_h*, *label*, *mask_group*)

Bases: `object`

Abstract class to represent image segmentation mask. Its two main subclasses are RoiMask and NeuropilMask. The former represents the mask of a region of interest (ROI), such as a cell observed in 2-photon imaging. The latter represents the neuropil around that cell, and is useful when subtracting the neuropil signal from the measured ROI signal.

This class should not be instantiated directly.

Parameters

image_w: integer
Width of image that ROI resides in

image_h: integer
Height of image that ROI resides in

label: text
User-defined text label to identify mask

mask_group: integer
User-defined number to help put masks into different categories

`get_mask_plane()`

Returns mask content on full-size image plane

Returns

numpy 2D array [img_rows][img_cols]

`init_by_pixels(border, pix_list)`

Initialize mask using a list of mask pixels

Parameters

border: float[4]
Coordinates defining useable area of image. See create_roi_mask()

pix_list: integer[][2]
List of pixel coordinates (x,y) that define the mask

`property overlaps_motion_border`

`class allensdk.brain_observatory.roi_masks.NeuropilMask(w, h, label, mask_group)`

Bases: *Mask*

`init_by_mask(border, array)`

Initialize mask using spatial mask

Parameters

border: float[4]
Border widths on the [right, left, down, up] sides. The resulting neuropil mask will not include pixels falling into a border.

array: integer[image height][image width]
Image-sized array that describes the mask. Active parts of the mask should have values >0. Background pixels must be zero

`class allensdk.brain_observatory.roi_masks.RoiMask(image_w, image_h, label, mask_group)`

Bases: *Mask*

`init_by_mask(border, array)`

Initialize mask using spatial mask

Parameters

border: float[4]
Coordinates defining useable area of image. See create_roi_mask().

roi_mask: integer[image height][image width]
Image-sized array that describes the mask. Active parts of the mask should have values >0. Background pixels must be zero

`allensdk.brain_observatory.roi_masks.calculate_roi_and_neuropil_traces(movie_h5, roi_mask_list, motion_border)`

get roi and neuropil masks

allensdk.brain_observatory.roi_masks.**calculate_traces**(*stack*, *mask_list*, *block_size=1000*)

Calculates the average response of the specified masks in the image stack

Parameters

stack: float[image height][image width]
Image stack that masks are applied to

mask_list: list<Mask>
List of masks

Returns

float[number masks][number frames]
This is the average response for each Mask in each image frame

allensdk.brain_observatory.roi_masks.**create_neuropil_mask**(*roi*, *border*, *combined_binary_mask*, *label=None*)

Conveninece function to create and initializes a Neuropil mask. Neuropil masks are defined as the region around an ROI, up to 13 pixels out, that does not include other ROIs

Parameters

roi: RoiMask object
The ROI that the neuropil masks will be based on

border: float[4]
Border widths on the [right, left, down, up] sides. The resulting neuropil mask will not include pixels falling into a border.

combined_binary_mask
List of pixel coordinates (x,y) that define the mask

combined_binary_mask: integer[image_h][image_w]
Image-sized array that shows the position of all ROIs in the image. ROI masks should have a value of one. Background pixels must be zero. In other words, ithe combined_binary_mask is a bitmap union of all ROI masks

label: text
User-defined text label to identify the mask

Returns

NeuropilMask object

allensdk.brain_observatory.roi_masks.**create_roi_mask**(*image_w*, *image_h*, *border*, *pix_list=None*, *roi_mask=None*, *label=None*, *mask_group=-1*)

Conveninece function to create and initializes an RoiMask

Parameters

image_w: integer
Width of image that ROI resides in

image_h: integer
Height of image that ROI resides in

border: float[4]
Coordinates defining useable area of image. If the entire image is usable, and masks are valid anywhere in the image, this should be [0, 0, 0, 0]. The following constants help describe the array order:

RIGHT_SHIFT = 0

LEFT_SHIFT = 1

DOWN_SHIFT = 2

UP_SHIFT = 3

When parts of the image are unusable, for example due motion correction shifting of different image frames, the border array should store the usable image area

pix_list: integer[][2]
List of pixel coordinates (x,y) that define the mask

roi_mask: integer[image_h][image_w]
Image-sized array that describes the mask. Active parts of the mask should have values >0. Background pixels must be zero

label: text
User-defined text label to identify mask

mask_group: integer
User-defined number to help put masks into different categories

Returns

RoiMask object

allensdk.brain_observatory.roi_masks.**create_roi_mask_array**(*rois*)

Create full image mask array from list of RoiMasks.

Parameters

rois: list<RoiMask>
List of roi masks.

Returns

np.ndarray: NxWxH array
Boolean array of of len(rois) image masks.

allensdk.brain_observatory.roi_masks.**validate_mask**(*mask*)

Check a given roi or neuropil mask for (a subset of) disqualifying problems.

allensdk.brain_observatory.running_speed module

class allensdk.brain_observatory.running_speed.**RunningSpeed**(*timestamps: ndarray*, *values: ndarray*)

Bases: `tuple`

Describes the rate at which an experimental subject ran during a session.

values
[np.ndarray] running speed (cm/s) at each sample point

timestamps
[np.ndarray] The time at which each sample was collected (s).

timestamps: ndarray
Alias for field number 0

values: ndarray
Alias for field number 1

allensdk.brain_observatory.session_analysis module

class allensdk.brain_observatory.session_analysis.**SessionAnalysis**(*nwb_path*, *save_path*)

Bases: object

Run all of the stimulus-specific analyses associated with a single experiment session.

Parameters

nwb_path: string, path to NWB file
save_path: string, path to HDF5 file to store outputs. Recommended NOT to modify the NWB file.

append_experiment_metrics(*metrics*)

Extract stimulus-agnostic metrics from an experiment into a dictionary

append_metadata(*df*)

Append the metadata fields from the NWB file as columns to a pd.DataFrame

append_metrics_drifting_grating(*metrics*, *dg*)

Extract metrics from the DriftingGratings peak response table into a dictionary.

append_metrics_locally_sparse_noise(*metrics*, *lsn*)

Extract metrics from the LocallySparseNoise peak response table into a dictionary.

append_metrics_natural_movie_one(*metrics*, *nma*)

Extract metrics from the NaturalMovie(stimulus_info.NATURAL_MOVIE_ONE) peak response table into a dictionary.

append_metrics_natural_movie_three(*metrics*, *nma*)

Extract metrics from the NaturalMovie(stimulus_info.NATURAL_MOVIE_THREE) peak response table into a dictionary.

append_metrics_natural_movie_two(*metrics*, *nma*)

Extract metrics from the NaturalMovie(stimulus_info.NATURAL_MOVIE_TWO) peak response table into a dictionary.

append_metrics_natural_scene(*metrics*, *ns*)

Extract metrics from the NaturalScenes peak response table into a dictionary.

append_metrics_static_grating(*metrics*, *sg*)

Extract metrics from the StaticGratings peak response table into a dictionary.

save_session_a(*dg*, *nm1*, *nm3*, *peak*)

Save the output of session A analysis to self.save_path.

Parameters

dg: DriftingGratings instance
nm1: NaturalMovie instance
This NaturalMovie instance should have been created with movie_name = stimulus_info.NATURAL_MOVIE_ONE

nm3: NaturalMovie instance
This NaturalMovie instance should have been created with movie_name = stimulus_info.NATURAL_MOVIE_THREE

peak: pd.DataFrame
The combined peak response property table created in self.session_a().

save_session_b(*sg*, *nm1*, *ns*, *peak*)

Save the output of session B analysis to self.save_path.

Parameters

sg: StaticGratings instance
nm1: NaturalMovie instance
This NaturalMovie instance should have been created with movie_name = stimulus_info.NATURAL_MOVIE_ONE

ns: NaturalScenes instance
peak: pd.DataFrame
The combined peak response property table created in self.session_b().

save_session_c(*lsn*, *nm1*, *nm2*, *peak*)

Save the output of session C analysis to self.save_path.

Parameters

lsn: LocallySparseNoise instance
nm1: NaturalMovie instance
This NaturalMovie instance should have been created with movie_name = stimulus_info.NATURAL_MOVIE_ONE

nm2: NaturalMovie instance
This NaturalMovie instance should have been created with movie_name = stimulus_info.NATURAL_MOVIE_TWO

peak: pd.DataFrame
The combined peak response property table created in self.session_c().

save_session_c2(*lsn4*, *lsn8*, *nm1*, *nm2*, *peak*)

Save the output of session C2 analysis to self.save_path.

Parameters

lsn4: LocallySparseNoise instance
This LocallySparseNoise instance should have been created with self.stimulus = stimulus_info.LOCALLY_SPARSE_NOISE_4DEG.

lsn8: LocallySparseNoise instance
This LocallySparseNoise instance should have been created with self.stimulus = stimulus_info.LOCALLY_SPARSE_NOISE_8DEG.

nm1: NaturalMovie instance
This NaturalMovie instance should have been created with movie_name = stimulus_info.NATURAL_MOVIE_ONE

nm2: NaturalMovie instance
This NaturalMovie instance should have been created with movie_name = stimulus_info.NATURAL_MOVIE_TWO

peak: pd.DataFrame
The combined peak response property table created in self.session_c2().

session_a(*plot_flag=False*, *save_flag=True*)

Run stimulus-specific analysis for natural movie one, natural movie three, and drifting gratings. The input NWB be for a stimulus_info.THREE_SESSION_A experiment.

Parameters

plot_flag: bool
Whether to generate brain_observatory_plotting work plots after running analysis.

save_flag: bool
Whether to save the output of analysis to self.save_path upon completion.

session_b(*plot_flag=False*, *save_flag=True*)

Run stimulus-specific analysis for natural scenes, static gratings, and natural movie one. The input NWB be for a stimulus_info.THREE_SESSION_B experiment.

Parameters

plot_flag: bool
Whether to generate brain_observatory_plotting work plots after running analysis.

save_flag: bool
Whether to save the output of analysis to self.save_path upon completion.

session_c(*plot_flag=False*, *save_flag=True*)

Run stimulus-specific analysis for natural movie one, natural movie two, and locally sparse noise. The input NWB be for a stimulus_info.THREE_SESSION_C experiment.

Parameters

plot_flag: bool
Whether to generate brain_observatory_plotting work plots after running analysis.

save_flag: bool
Whether to save the output of analysis to self.save_path upon completion.

session_c2(*plot_flag=False*, *save_flag=True*)

Run stimulus-specific analysis for locally sparse noise (4 deg.), locally sparse noise (8 deg.), natural movie one, and natural movie two. The input NWB be for a stimulus_info.THREE_SESSION_C2 experiment.

Parameters

plot_flag: bool
Whether to generate brain_observatory_plotting work plots after running analysis.

save_flag: bool
Whether to save the output of analysis to self.save_path upon completion.

verify_roi_lists_equal(*roi1*, *roi2*)

TODO: replace this with simpler numpy comparisons

allensdk.brain_observatory.session_analysis.**main**()

allensdk.brain_observatory.session_analysis.**multi_dataframe_merge**(*dfs*)

merge a number of pd.DataFrames into a single dataframe on their index columns. If any columns are duplicated, prefer the first occuring instance of the column

allensdk.brain_observatory.session_analysis.**run_session_analysis**(*nwb_path*, *save_path*, *plot_flag=False*, *save_flag=True*)

Inspect an NWB file to determine which experiment session was run and compute all stimulus-specific analyses.

Parameters

nwb_path: string
Path to NWB file.

save_path: string
path to save results. Recommended NOT to use NWB file.

plot_flag: bool
Whether to save brain_observatory_plotting work plots.

save_flag: bool
Whether to save results to save_path.

allensdk.brain_observatory.session_api_utils module

class allensdk.brain_observatory.session_api_utils.**ParamsMixin**(*ignore: set = {'api'}*)

Bases: object

This mixin adds parameter management functionality to the class it is mixed into.

This mixin expects that the class it is mixed into will have an __init__ with type annotated parameters. It also expects for the class to have semi-private attributes of the __init__ type annotated parameters.

Example:

SomeClassWhereParamManagementIsDesired(ParamsMixin):

Managed params should be typed (with simple types if possible)! def __init__(self, param_to_ignore, a_param_1: int, a_param_2: float,

b_param_1: list):

Parameters can be ignored by the mixin super().__init__(ignore={'param_to_ignore'})

Pay attention to the naming scheme! self._a_param_1 = a_param_1 self._a_param_2 = a_param_2 self._b_param_1 = b_param_1

...

After being mixed in, methods like 'get_params', 'set_params', 'needs_data_refresh', and 'clear_updated_params' will be available.

clear_updated_params(*data_params: set*)

This method clears 'updated params' whose data have been updated

get_params() → Dict[str, Any]

Get managed params and their values

needs_data_refresh(*data_params: set*) → bool

Check if specific params have been updated via *set_params()*

set_params(**params*)

Set managed params

allensdk.brain_observatory.session_api_utils.**is_equal**(*a: Any*, *b: Any*) → bool

Function to deal with checking if two variables of possibly mixed types have the same value.

allensdk.brain_observatory.session_api_utils.**sessions_are_equal**(*A*, *B*, *reraise=False*, *ignore_keys: Dict[str, Set[str]] | None = None*, *skip_fields: Iterable | None = None*, *test_methods=False*) → bool

Check if two Session objects are equal (have same property and get method values).

Parameters

A
[Session A] The first session to compare

B
[Session B] The second session to compare

reraise
[bool, optional] Whether to reraise when encountering an Assertion or AttributeError, by default False

ignore_keys
Set of keys to ignore for property/method. Should be given as {property/method name: {field_to_ignore, ...}, ...}

test_methods
Whether to test get methods

skip_fields
Do not compare these fields

Returns

bool
Whether the two sessions are equal to one another.

allensdk.brain_observatory.static_gratings module

class allensdk.brain_observatory.static_gratings.**StaticGratings**(*data_set*, ***kwargs*)

Bases: *StimulusAnalysis*

Perform tuning analysis specific to static gratings stimulus.

Parameters

data_set: BrainObservatoryNwbDataSet object

property extralength

static from_analysis_file(*data_set*, *analysis_file*)

get_noise_correlation(*corr='spearman'*)

get_peak()

Computes metrics related to each cell's peak response condition.

Returns

Panda data frame with the following fields (_sg suffix is for static grating):

- ori_sg (orientation)
- sf_sg (spatial frequency)
- phase_sg
- response_variability_sg
- osi_sg (orientation selectivity index)
- peak_dff_sg (peak dF/F)
- ptest_sg

- time_to_peak_sg

get_representational_similarity(*corr='spearman'*)

get_response()

Computes the mean response for each cell to each stimulus condition. Return is a (# orientations, # spatial frequencies, # phases, # cells, 3) np.ndarray. The final dimension contains the mean response to the condition (index 0), standard error of the mean of the response to the condition (index 1), and the number of trials with a significant response ($p < 0.05$) to that condition (index 2).

Returns

Numpy array storing mean responses.

get_signal_correlation(*corr='spearman'*)

property interlength

property number_ori

property number_phase

property number_sf

open_fan_plot(*cell_specimen_id=None*, *include_labels=False*, *cell_index=None*)

property orivals

property phasevals

plot_orientation_selectivity(*si_range=[0, 1.5]*, *n_hist_bins=50*, *color='#ccccdd'*, *p_value_max=0.05*, *peak_dff_min=3*)

plot_preferred_orientation(*include_labels=False*, *si_range=[0, 1.5]*, *color='#ccccdd'*, *p_value_max=0.05*, *peak_dff_min=3*)

plot_preferred_spatial_frequency(*si_range=[0, 1.5]*, *color='#ccccdd'*, *p_value_max=0.05*, *peak_dff_min=3*)

plot_time_to_peak(*p_value_max=0.05*, *color_map=<matplotlib.colors.LinearSegmentedColormap object>*)

populate_stimulus_table()

Implemented by subclasses.

reshape_response_array()

Returns

response array in cells x stim conditions x repetition for

noise correlations this is a re-organization of the mean sweep response table

property sfvals

property sweeplength

allensdk.brain_observatory.stimulus_analysis module

class allensdk.brain_observatory.stimulus_analysis.**StimulusAnalysis**(*data_set*)

Bases: object

Base class for all response analysis code. Subclasses are responsible for computing metrics and traces relevant to a particular stimulus. The base class contains methods for organizing sweep responses row of a stimulus stable (get_sweep_response). Subclasses implement the get_response method, computes the mean sweep response to all sweeps for a each stimulus condition.

Parameters

data_set: BrainObservatoryNwbDataSet instance
speed_tuning: boolean, deprecated
Whether or not to compute speed tuning histograms

property acquisition_rate

property binned_cells_sp

property binned_cells_vis

property binned_dx_sp

property binned_dx_vis

property cell_id

property celltraces

property dfftraces

property dxcm

property dxtime

get_fluorescence()

get_peak()

Implemented by subclasses.

get_response()

Implemented by subclasses.

get_speed_tuning(*binsize*)

Calculates speed tuning, spontaneous versus visually driven. The return is a 5-tuple of speed and dF/F histograms.

binned_dx_sp: (bins,2) np.ndarray of running speeds binned during spontaneous activity stimulus. The first bin contains all speeds below 1 cm/s. Dimension 0 is mean running speed in the bin. Dimension 1 is the standard error of the mean.

binned_cells_sp: (bins,2) np.ndarray of fluorescence during spontaneous activity stimulus. First bin contains all data for speeds below 1 cm/s. Dimension 0 is mean fluorescence in the bin. Dimension 1 is the standard error of the mean.

binned_dx_vis: (bins,2) np.ndarray of running speeds outside of spontaneous activity stimulus. The first bin contains all speeds below 1 cm/s. Dimension 0 is mean running speed in the bin. Dimension 1 is the standard error of the mean.

binned_cells_vis: np.ndarray of fluorescence outside of spontaneous activity stimulu. First bin contains all data for speeds below 1 cm/s. Dimension 0 is mean fluorescence in the bin. Dimension 1 is the standard error of the mean.

peak_run: pd.DataFrame of speed-related properties of a cell.

Returns

tuple: binned_dx_sp, binned_cells_sp, binned_dx_vis, binned_cells_vis, peak_run

`get_sweep_response()`

Calculates the response to each sweep in the stimulus table for each cell and the mean response. The return is a 3-tuple of:

- sweep_response: pd.DataFrame of response dF/F traces organized

by cell (column) and sweep (row)

- mean_sweep_response: mean values of the traces returned in

sweep_response

- pval: p value from 1-way ANOVA comparing response during sweep

to response prior to sweep

Returns

3-tuple: sweep_response, mean_sweep_response, pval

`property mean_sweep_response`

`property numbercells`

`property peak`

`property peak_run`

`plot_representational_similarity(repsim, stimulus=False)`

`plot_running_speed_histogram(xlim=None, nbins=None)`

`plot_speed_tuning(cell_specimen_id=None, cell_index=None, evoked_color='#b30000', spontaneous_color='#0000b3')`

`populate_stimulus_table()`

Implemented by subclasses.

`property pval`

`property response`

`property roi_id`

`row_from_cell_id(csid=None, idx=None)`

`property stim_table`

`property sweep_response`

property timestamps

allensdk.brain_observatory.stimulus_analysis.**nonraising_ks_2samp**(*data1*, *data2*, ***kwargs*)

scipy.stats.ks_2samp now raises a ValueError if one of the input arrays is of length 0. Previously it signaled this case by returning nans. This function restores the prior behavior.

allensdk.brain_observatory.stimulus_info module

class allensdk.brain_observatory.stimulus_info.**BinaryIntervalSearchTree**(*search_list*)

Bases: object

add(*input_list*, *tmp=None*)

static from_df(*input_df*)

search(*fi*, *tmp=None*)

class allensdk.brain_observatory.stimulus_info.**BrainObservatoryMonitor**(*experiment_geometry=None*)

Bases: *Monitor*

http://help.brain-map.org/display/observatory/Documentation?preview=/10616846/10813485/VisualCoding_VisualStimuli.pdf # noqa: E501 https://www.cnet.com/products/asus-pa248q/specs/

grating_to_screen(*phase*, *spatial_frequency*, *orientation*, ***kwargs*)

lsn_image_to_screen(*img*, ***kwargs*)

pixels_to_visual_degrees(*n*, ***kwargs*)

visual_degrees_to_pixels(*vd*, ***kwargs*)

warp_image(*img*, ***kwargs*)

class allensdk.brain_observatory.stimulus_info.**ExperimentGeometry**(*distance*, *mon_height_cm*, *mon_width_cm*, *mon_res*, *eyepoint*)

Bases: object

generate_warp_coordinates()

property warp_coordinates

class allensdk.brain_observatory.stimulus_info.**Monitor**(*n_pixels_r*, *n_pixels_c*, *panel_size*, *spatial_unit*)

Bases: object

property aspect_ratio

get_mask()

grating_to_screen(*phase*, *spatial_frequency*, *orientation*, *distance_from_monitor*, *p2p_amp=256*, *baseline=127*, *translation=(0, 0)*)

property height

lsn_image_to_screen(*img*, *stimulus_type*, *origin='lower'*, *background_color=127*, *translation=(0, 0)*)

map_stimulus(*source_stimulus_coordinate*, *source_stimulus_type*, *target_stimulus_type*)

property mask

natural_movie_image_to_screen(*img*, *origin='lower'*, *translation=(0, 0)*)

natural_scene_image_to_screen(*img*, *origin='lower'*, *translation=(0, 0)*)

property panel_size

property pixel_size

pixels_to_visual_degrees(*n*, *distance_from_monitor*, *small_angle_approximation=True*)

set_spatial_unit(*new_unit*)

show_image(*img*, *ax=None*, *show=True*, *mask=False*, *warp=False*, *origin='lower'*)

spatial_frequency_to_pix_per_cycle(*spatial_frequency*, *distance_from_monitor*)

visual_degrees_to_pixels(*vd*, *distance_from_monitor*, *small_angle_approximation=True*)

property width

class allensdk.brain_observatory.stimulus_info.**StimulusSearch**(*nwb_dataset*)

Bases: object

search(*fi*)

allensdk.brain_observatory.stimulus_info.**all_stimuli**()

Return a list of all stimuli in the data set

allensdk.brain_observatory.stimulus_info.**get_spatial_grating**(*height=None*, *aspect_ratio=None*, *ori=None*, *pix_per_cycle=None*, *phase=None*, *p2p_amp=2*, *baseline=0*)

allensdk.brain_observatory.stimulus_info.**get_spatio_temporal_grating**(*t*, *temporal_frequency=None*, ***kwargs*)

allensdk.brain_observatory.stimulus_info.**lsn_coordinate_to_monitor_coordinate**(*lsn_coordinate*, *monitor_shape*, *stimulus_type*)

allensdk.brain_observatory.stimulus_info.**make_display_mask**(*display_shape=(1920, 1200)*)

Build a display-shaped mask that indicates which pixels are on screen after warping the stimulus.

allensdk.brain_observatory.stimulus_info.**map_monitor_coordinate_to_stimulus_coordinate**(*monitor_coordinate*, *monitor_shape*, *stimulus_type*)

allensdk.brain_observatory.stimulus_info.**map_monitor_coordinate_to_template_coordinate**(*monitor_coord*, *monitor_shape*, *template_shape*)

allensdk.brain_observatory.stimulus_info.**map_stimulus**(*source_stimulus_coordinate*, *source_stimulus_type*, *target_stimulus_type*, *monitor_shape*)

allensdk.brain_observatory.stimulus_info.**map_stimulus_coordinate_to_monitor_coordinate**(*template_coordinate*, *monitor_shape*, *stimulus_type*)

allensdk.brain_observatory.stimulus_info.**map_template_coordinate_to_monitor_coordinate**(*template_coord*, *monitor_shape*, *template_shape*)

allensdk.brain_observatory.stimulus_info.**mask_stimulus_template**(*template_display_coords*, *template_shape*, *display_mask=None*, *threshold=1.0*)

Build a mask for a stimulus template of a given shape and display coordinates that indicates which part of the template is on screen after warping.

Parameters

template_display_coords: list
list of (x,y) display coordinates

template_shape: tuple
(width,height) of the display template

display_mask: np.ndarray
boolean 2D mask indicating which display coordinates are on screen after warping.

threshold: float
Fraction of pixels associated with a template display coordinate that should remain on screen to count as belonging to the mask.

Returns

tuple: (template mask, pixel fraction)

allensdk.brain_observatory.stimulus_info.**monitor_coordinate_to_lsn_coordinate**(*monitor_coordinate*, *monitor_shape*, *stimulus_type*)

allensdk.brain_observatory.stimulus_info.**monitor_coordinate_to_natural_movie_coordinate**(*monitor_coordinate*, *monitor_shape*)

allensdk.brain_observatory.stimulus_info.**natural_movie_coordinate_to_monitor_coordinate**(*natural_movie_coor*, *monitor_shape*)

allensdk.brain_observatory.stimulus_info.**natural_scene_coordinate_to_monitor_coordinate**(*natural_scene_coor*, *monitor_shape*)

allensdk.brain_observatory.stimulus_info.**rotate**(*X*, *Y*, *theta*)

allensdk.brain_observatory.stimulus_info.**sessions_with_stimulus**(*stimulus*)

Return the names of the sessions that contain a given stimulus.

allensdk.brain_observatory.stimulus_info.**stimuli_in_session**(*session*, *allow_unknown=True*)

Return a list what stimuli are available in a given session.

Parameters

session: string

Must be one of: [
stimulus_info.THREE_SESSION_A, stimulus_info.THREE_SESSION_B, stimulus_info.THREE_SESSION_C, stimulus_info.THREE_SESSION_C2]

allensdk.brain_observatory.stimulus_info.**translate_image_and_fill**(*img*, *translation=(0, 0)*)

allensdk.brain_observatory.stimulus_info.**warp_stimulus_coords**(*vertices*, *distance=15.0*, *mon_height_cm=32.5*, *mon_width_cm=51.0*, *mon_res=(1920, 1200)*, *eyepoint=(0.5, 0.5)*)

For a list of screen vertices, provides a corresponding list of texture coordinates.

Parameters

vertices: numpy.ndarray
[[x0,y0], [x1,y1], . . .] A set of vertices to convert to texture positions.

distance: float
distance from the monitor in cm.

mon_height_cm: float
monitor height in cm

mon_width_cm: float
monitor width in cm

mon_res: tuple
monitor resolution (x,y)

eyepoint: tuple

Returns

np.ndarray
x,y coordinates shaped like the input that describe what pixel coordinates are displayed an the input coordinates after warping the stimulus.

allensdk.brain_observatory.sync_dataset module

dataset.py

Dataset object for loading and unpacking an HDF5 dataset generated by sync.py

@author: derricw

Allen Institute for Brain Science

Dependencies

numpy http://www.numpy.org/ h5py http://www.h5py.org/

class allensdk.brain_observatory.sync_dataset.**Dataset**(*path*)

Bases: object

A sync dataset. Contains methods for loading and parsing the binary data.

Parameters

path
[str] Path to HDF5 file.

Examples

```
>>> dset = Dataset('my_h5_file.h5')
>>> logger.info(dset.meta_data)
>>> dset.stats()
>>> dset.close()
```

```
>>> with Dataset('my_h5_file.h5') as d:
...     logger.info(dset.meta_data)
...     dset.stats()
```

The sync file documentation from MPE can be found at sharepoint > Instrumentation > Shared Documents > Sync_line_labels_discussion_2020-01-27-.xlsx # NOQA E501 Direct link: https://alleninstitute.sharepoint.com/:x:/s/Instrumentation/ES2bi1xJ3E9NupX-zQeXTlYBS2mVVySycfbCQhsD_jPMUw?e=Z9jCwH

BEHAVIOR_TRACKING_KEYS = ('beh_frame_received', 'cam1_exposure', 'behavior_monitoring')

DEPRECATED_KEYS = {}

EYE_TRACKING_KEYS = ('eye_frame_received', 'cam2_exposure', 'eyetracking', 'eye_cam_exposing', 'eye_tracking')

FRAME_KEYS = ('frames', 'stim_vsync', 'vsync_stim')

OPTOGENETIC_STIMULATION_KEYS = ('LED_sync', 'opto_trial')

PHOTODIODE_KEYS = ('photodiode', 'stim_photodiode')

property analog_meta_data

close()

Closes the dataset.

duty_cycle(*line*)

Doesn't work right now. Freezes python for some reason.

Returns the duty cycle of a line.

frequency(*line*, *edge='rising'*)

Returns the average frequency of a line.

get_all_bits()

Returns the data for all bits.

get_all_events()

Returns all counter values and their cooresponding IO state.

get_all_times(*units='samples'*)

Returns all counter values.

Parameters

units
[str] Return times in 'samples' or 'seconds'

get_analog_channel(*channel*, *start_time=0.0*, *stop_time=None*, *downsample=1*)

Returns the data from the specified analog channel between the
timepoints.

Args:
channel (int, str): desired channel index or label start_time (Optional[float]): start time in seconds stop_time (Optional[float]): stop time in seconds downsample (Optional[int]): downsample factor

Returns:
ndarray: slice of data for specified channel

Raises:
KeyError: no analog data present

get_analog_meta()

Returns the metadata for the analog data.

get_bit(*bit*)

Returns the values for a specific bit.

Parameters

bit
[int] Bit to return.

get_bit_changes(*bit*)

Returns the first derivative of a specific bit.
Data points are 1 on rising edges and 255 on falling edges.

Parameters

bit
[int] Bit for which to return changes.

`get_edges`(*kind: str*, *keys: str | Sequence[str]*, *units: str = 'seconds'*, *permissive: bool = False*) → ndarray | None

Utility function for extracting edge times from a line

Parameters

kind
[One of "rising", "falling", or "all". Should this method return] timestamps for rising, falling or both edges on the appropriate line

keys
[These will be checked in sequence. Timestamps will be returned] for the first which is present in the line labels

units
[one of "seconds", "samples", or "indices". The returned] "time"stamps will be given in these units.

raise_missing
[If True and no matching line is found, a KeyError will] be raised

Returns

An array of edge times. If raise_missing is False and none of the keys
were found, returns None.

Raises

KeyError
[none of the provided keys were found among this dataset's] line labels

`get_events_by_bit`(*bit*, *units='samples'*)

Returns all counter values for transitions (both rising and falling)
for a specific bit.

Parameters

bit
[int] Bit for which to return events.

`get_events_by_line`(*line*, *units='samples'*)

Returns all counter values for transitions (both rising and falling)
for a specific line.

Parameters

line
[str] Line for which to return events.

`get_falling_edges`(*line*, *units='samples'*)

Returns the counter values for the falling edges for a specific bit
or line.

Parameters

line
[str] Line for which to return edges.

`get_line`(*line*)

Returns the values for a specific line.

Parameters

line
[str] Line to return.

`get_line_changes`(*line*)

Returns the first derivative of a specific line.
Data points are 1 on rising edges and 255 on falling edges.

Parameters

line
[(str)] Line name for which to return changes.

`get_nearest`(*source*, *target*, *source_edge='rising'*, *target_edge='rising'*, *direction='previous'*, *units='indices'*)

For all values of the source line, finds the nearest edge from the
target line.

By default, returns the indices of the target edges.

Args:
source (str, int): desired source line target (str, int): desired target line source_edge [Optional(str)]: "rising" or "falling" source edges target_edge [Optional(str): "rising" or "falling" target edges direction (str): "previous" or "next". Whether to prefer the

previous edge or the following edge.

units (str): "indices"

`get_rising_edges`(*line*, *units='samples'*)

Returns the counter values for the rizing edges for a specific bit or
line.

Parameters

line
[str] Line for which to return edges.

`line_stats`(*line*, *print_results=True*)

Quick-and-dirty analysis of a bit.

##TODO: Split this up into smaller functions.

`load`(*path*)

Loads an hdf5 sync dataset.

Parameters

path
[str] Path to hdf5 file.

period(*line*, *edge='rising'*)

Returns a dictionary with avg, min, max, and st of period for a line.

plot_all(*start_time*, *stop_time*, *auto_show=True*)

Plot all active bits.

Yikes. Come up with a better way to show this.

plot_bit(*bit*, *start_time=0.0*, *end_time=None*, *auto_show=True*, *axes=None*, *name=''*)

Plots a specific bit at a specific time period.

plot_bits(*bits*, *start_time=0.0*, *end_time=None*, *auto_show=True*)

Plots a list of bits.

plot_line(*line*, *start_time=0.0*, *end_time=None*, *auto_show=True*)

Plots a specific line at a specific time period.

plot_lines(*lines*, *start_time=0.0*, *end_time=None*, *auto_show=True*)

Plots specific lines at a specific time period.

property sample_freq

stats()

Quick-and-dirty analysis of all bits. Prints a few things about each
bit where events are found.

allensdk.brain_observatory.sync_dataset.**get_bit**(*uint_array*, *bit*)

Returns a bool array for a specific bit in a uint ndarray.

Parameters

uint_array
[(numpy.ndarray)] The array to extract bits from.

bit
[(int)] The bit to extract.

allensdk.brain_observatory.sync_dataset.**unpack_uint32**(*uint32_array*, *endian='L'*)

Unpacks an array of 32-bit unsigned integers into bits.

Default is least significant bit first.

***Not currently used by sync dataset because get_bit is better and does**
basically the same thing. I'm just leaving it in because it could potentially account for endianness and possibly have other uses in the future.

allensdk.brain_observatory.sync_stim_aligner module

Module contents

class allensdk.brain_observatory.**JSONEncoder**(*, *skipkeys=False*, *ensure_ascii=True*, *check_circular=True*, *allow_nan=True*, *sort_keys=False*, *indent=None*, *separators=None*, *default=None*)

Bases: JSONEncoder

default(*o*)

Implement this method in a subclass such that it returns a serializable object for `o`, or calls the base implementation (to raise a `TypeError`).

For example, to support arbitrary iterators, you could implement default like this:

```
def default(self, o):
    try:
        iterable = iter(o)
    except TypeError:
        pass
    else:
        return list(iterable)
    # Let the base class default method raise the TypeError
    return JSONEncoder.default(self, o)
```

allensdk.brain_observatory.**dict_to_indexed_array**(*dc*, *order=None*)

Given a dictionary and an ordered arr, build a concatenation of the dictionary's values and an index describing how that concatenation can be unpacked

allensdk.brain_observatory.**hook**(*json_dict*)

6.1.3 allensdk.config package

Subpackages

allensdk.config.app package

Submodules

allensdk.config.app.application_config module

class allensdk.config.app.application_config.**ApplicationConfig**(*defaults*, *name='app'*, *halp='Run application.'*, *default_log_config=None*)

Bases: `object`

Convenience class that handles of application configuration from environment variables, .conf files and the command line using Python standard libraries and formats.

apply_configuration_from_command_line(*parsed_args*)

Read application configuration variables from the command line.

Unassigned variables are left unchanged if previously assigned, set to their default values, or None if no default is specified at init time. Assigned variables will overwrite the previous value.

see: https://docs.python.org/2/howto/argparse.html

Parameters

parsed_args
[dict] the arguments as parsed from the command line.

apply_configuration_from_environment()

Read application configuration variables from the environment.

The variable names are upper case and have a prefix defined by the application.

See: https://docs.python.org/2/library/os.html

apply_configuration_from_file(*config_file_path*)

Read application configuration variables from a .conf file.

Unassigned variables are set to their default values or None if no default is specified at init time. The variables are found in a section named by the application.

Parameters

config_file_path
[string] path to to an INI (.conf) or JSON format application config file.

Returns

see: https://docs.python.org/2/library/configparser.html

create_argparser()

Initialization for the command-line parsing stage.

An application specific prefix is applied to argument names.

Parameters

prog
[string] Application specific prefix for argument names.

description
[string] A brief 'help' description of the application.

Returns

argParse.ArgumentParser
The initialized argument parser object.

Notes

Defaults are set at the first environment reading. Command line args only override them when present

from_json_file(*json_path*)

Read an application configuration from a JSON format file.

Parameters

json_path
[string] Path to the JSON file.

Returns

string
An application configuration in INI format

from_json_string(*json_string*)

Read a configuration from a JSON format string.

Parameters

json_string
[string] A JSON-formatted string containing an application configuration.

Returns

string
An application configuration in INI format

`load`(*command_line_args*, *disable_existing_loggers=True*)

Load application configuration options, first from the environment, then from the configuration file, then from the command line.

Each stage of loading can override the previous stage.

Parameters

command_line_args
[dict] Parameters passed to the application.

disable_existing_loggers
[boolean] Reset the logging system or not.

Returns

fileConfig
Configuration object with all levels applied

`parse_command_line_args`(*args*)

Simply call the internal argparser object.

Parameters

args
[array] Parameters passed to the application.

Returns

Namespace
Parsed paramenters.

`to_config_string`(*description*)

Create a configuration string from a dict.

Parameters

description
[dict] Configuration options for an application.

Returns

string
Equivalent configuration as an INI format string

Notes

The Python configparser library natively supports this functionality in Python 3.

Module contents

allensdk.config.app is a package that assists in configuring application software, as opposed to domain-specific configuration.

allensdk.config.model package

Subpackages

allensdk.config.model.formats package

Submodules

allensdk.config.model.formats.hdf5_util module

class allensdk.config.model.formats.hdf5_util.**Hdf5Util**

Bases: object

read(*file_path*)

write(*file_path*, *m*)

allensdk.config.model.formats.json_description_parser module

class allensdk.config.model.formats.json_description_parser.**JsonDescriptionParser**

Bases: *DescriptionParser*

log = <Logger allensdk.config.model.formats.json_description_parser (WARNING)>

read(*file_path*, *description=None*, *section=None*, ***kwargs*)

Parse a complete or partial configuration.

Parameters

json_string
[string] Input to parse.

description
[Description, optional] Where to put the parsed configuration. If None a new one is created.

section
[string, optional] Where to put the parsed configuration within the description.

Returns

Description
The input description with parsed configuration added.

Section is only specified for "bare" objects that are to be added to a section array.

read_string(*json_string*, *description=None*, *section=None*, ***kwargs*)

Parse a complete or partial configuration.

Parameters

json_string
[string] Input to parse.

description
[Description, optional] Where to put the parsed configuration. If None a new one is created.

section
[string, optional] Where to put the parsed configuration within the description.

Returns

Description
The input description with parsed configuration added.

Section is only specified for "bare" objects that are to be added to a section array.

`write`(*filename*, *description*)

Write the description to a JSON file.

Parameters

description
[Description] Object to write.

`write_string`(*description*)

Write the description to a JSON string.

Parameters

description
[Description] Object to write.

Returns

string
JSON serialization of the input.

allensdk.config.model.formats.pycfg_description_parser module

`class` allensdk.config.model.formats.pycfg_description_parser.**PycfgDescriptionParser**

Bases: *DescriptionParser*

log = <Logger allensdk.config.model.formats.pycfg_description_parser (WARNING)>

`read`(*pycfg_file_path*, *description=None*, *section=None*, ***kwargs*)

Read a serialized description from a Python (.pycfg) file.

Parameters

filename
[string] Name of the .pycfg file.

Returns

Description
Configuration object.

`read_string`(*python_string*, *description=None*, *section=None*, ***kwargs*)

Read a serialized description from a Python (.pycfg) string.

Parameters

python_string
[string] Python string with a serialized description.

Returns

Description
Configuration object.

`write`(*filename*, *description*)

Write the description to a Python (.pycfg) file.

Parameters

filename
[string] Name of the file to write.

`write_string`(*description*)

Write the description to a pretty-printed Python string.

Parameters

description
[Description] Configuration object to write.

Module contents

Submodules

allensdk.config.model.description module

`class allensdk.config.model.description.Description`

Bases: `object`

`fix_unary_sections`(*section_names=None*)

Wrap section contents that don't have the proper array surrounding them in an array.

Parameters

section_names
[list of strings, optional] Keys of sections that might not be in array form.

`is_empty`()

Check if anything is in the object.

Returns

boolean
true if self.data is missing or empty

`unpack`(*data*, *section=None*)

Read the manifest and other stand-alone configuration structure, or insert a configuration object into a section of an existing configuration.

Parameters

data
[dict] A configuration object including top level sections, or an configuration object to be placed within a section.

section
[string, optional.] If this is present, place data within an existing section array.

`unpack_manifest`(*data*)

Pull the manifest configuration section into a separate place.

Parameters

data
[dict] A configuration structure that still has a manifest section.

`update_data`(*data*, *section=None*)

Merge configuration data possibly from multiple files.

Parameters

data
[dict] Configuration structure to add.

section
[string, optional] What configuration section to read it into if the file does not specify.

allensdk.config.model.description_parser module

`class` allensdk.config.model.description_parser.`DescriptionParser`

Bases: `object`

`log = <Logger allensdk.config.model.description_parser (WARNING)>`

`parser_for_extension`(*filename*)

Choose a subclass that can read the format.

Parameters

filename
[string] For the extension.

Returns

DescriptionParser
Appropriate subclass.

`read`(*file_path*, *description=None*, *section=None*, ***kwargs*)

Parse data needed for a simulation.

Parameters

description
[dict] Configuration from parsing previous files.

section
[string, optional] What configuration section to read it into if the file does not specify.

`read_string`(*data_string*, *description=None*, *section=None*, *header=None*)

Parse data needed for a simulation from a string.

`write`(*filename*, *description*)

Save the configuration.

Parameters

filename
[string] Name of the file to write.

Module contents

Submodules

allensdk.config.manifest module

class allensdk.config.manifest.**Manifest**(*config=None*, *relative_base_dir='.'*, *version=None*)

Bases: object

Manages the location of external files referenced in an Allen SDK configuration

DIR = 'dir'

DIRNAME = 'dir_name'

FILE = 'file'

VERSION = 'manifest_version'

add_file(*file_key*, *file_name*, *dir_key=None*, *path_format=None*)

Insert a new file entry.

Parameters

file_key
[string] Reference to the entry.

file_name
[string] Subtitutions of the %s, %d style allowed.

dir_key
[string] Reference to the parent directory entry.

path_format
[string, optional] File type for further parsing.

add_path(*key*, *path*, *path_type='dir'*, *absolute=True*, *path_format=None*, *parent_key=None*)

Insert a new entry.

Parameters

key
[string] Identifier for referencing the entry.

path
[string] Specification for a path using %s, %d style substitution.

path_type
[string enumeration] 'dir' (default) or 'file'

absolute
[boolean] Is the spec relative to the process current directory.

path_format
[string, optional] Indicate a known file type for further parsing.

parent_key
[string] Refer to another entry.

`add_paths`(*path_info*)

add information about paths stored in the manifest.

Parameters

path_info
[dict] Information about the new paths

`as_dataframe`()

`check_dir`(*path_key*, *do_exit=False*)

Verify a directories existence or optionally exit.

Parameters

path_key
[string] Reference to the entry.

do_exit
[boolean] What to do if the directory is not present.

`create_dir`(*path_key*)

Make a directory for an entry.

Parameters

path_key
[string] Reference to the entry.

`get_format`(*path_key*)

Retrieve the type of a path entry.

Parameters

path_key
[string] reference to the entry

Returns

string
File type.

`get_path`(*path_key*, **args*)

Retrieve an entry with substitutions.

Parameters

path_key
[string] Refer to the entry to retrieve.

args
[any types, optional] arguments to be substituted into the path spec for %s, %d, etc.

Returns

string
Path with parent structure and substitutions applied.

`load_config`(*config*, *version=None*)

Load paths into the manifest from an Allen SDK config section.

Parameters

config
[Config] Manifest section of an Allen SDK config.

`log = <Logger allensdk.config.manifest (WARNING)>`

`resolve_paths`(*description_dict*, *suffix='_key'*)
Walk input items and expand those that refer to a manifest entry.

Parameters

description_dict
[dict] Any entries with key names ending in suffix will be expanded.

suffix
[string] Indicates the entries to be expanded.

`classmethod safe_make_parent_dirs`(*file_name*)
Create a parent directories for file.

Parameters

file_name
[string]

Returns

leftmost
[string] most rootward directory created

`classmethod safe_mkdir`(*directory*)
Create path if not already there.

Parameters

directory
[string] create it if it doesn't exist

Returns

leftmost
[string] most rootward directory created

`exception allensdk.config.manifest.ManifestVersionError`(*message*, *version*, *found_version*)
Bases: `Exception`

`property outdated`

allensdk.config.manifest_builder module

`class allensdk.config.manifest_builder.ManifestBuilder`
Bases: `object`

`add_path`(*key*, *spec*, *typename='dir'*, *parent_key=None*, *format=None*)

`add_section`(*name*, *contents*)

`as_dataframe`()

`df_columns = ['key', 'parent_key', 'spec', 'type', 'format']`

from_dataframe(*df*)

get_config()

get_manifest()

set_version(*value*)

write_json_file(*path*, *overwrite=False*)

write_json_string()

Module contents

allensdk.config.**enable_console_log**(*level=None*)

configure allensdk logging to output to the console.

Parameters

level

[int] logging level 0-50 (logging.INFO, logging.DEBUG, etc.)

Notes

See: Logging Cookbook

6.1.4 allensdk.core package

Subpackages

allensdk.core.lazy_property package

Submodules

allensdk.core.lazy_property.lazy_property module

class allensdk.core.lazy_property.lazy_property.**LazyProperty**(*api_method: Callable*, *wrappers: Iterable = ()*, *settable: bool = False*, **args*, ***kwargs*)

Bases: object

calculate()

allensdk.core.lazy_property.lazy_property_mixin module

class allensdk.core.lazy_property.lazy_property_mixin.**LazyPropertyMixin**

Bases: object

property LazyProperty

Module contents

Submodules

allensdk.core.auth_config module

allensdk.core.authentication module

class allensdk.core.authentication.**CredentialProvider**

Bases: ABC

METHOD = 'custom'

abstract provide(*credential*)

class allensdk.core.authentication.**DbCredentials**(*dbname*, *user*, *host*, *port*, *password*)

Bases: tuple

dbname

Alias for field number 0

host

Alias for field number 2

password

Alias for field number 4

port

Alias for field number 3

user

Alias for field number 1

class allensdk.core.authentication.**EnvCredentialProvider**(*environ: Dict[str, Any] | None = None*)

Bases: *CredentialProvider*

Provides credentials from environment variables for variables listed in CREDENTIAL_KEYS.

METHOD = 'env'

provide(*credential*)

allensdk.core.authentication.**credential_injector**(*credential_map: Dict[str, Any]*, *provider:* CredentialProvider | *None = None*)

Decorator used to inject credentials from another source if not explicitly provided in the function call. This function will only supply values for keyword arguments. All keys defined in *credential_map* must correspond to keyword arguments in the function signature.

Parameters

credential_map: Dict[Str: Any]
Dictionary where the keys are the keyword of a credential kwarg passed to the decorated function, and the values are the name of the credential in the credential provider (see CREDENTIAL_KEYS).

Example of credential_map for PostgresQueryMixin connecting to LIMS database:

{
"dbname": "LIMS_DBNAME", "user": "LIMS_USER", "host": "LIMS_HOST", "password": "LIMS_PASSWORD", "port": "LIMS_PORT"
}

provider: Optional[CredentialProvider]
Subclass of CredentialProvider to provide credentials to the wrapped function. If left unspecified, will default to EnvCredentialProvider, which provides credentials from environment variables.

allensdk.core.authentication.**get_credential_provider**()

allensdk.core.authentication.**set_credential_provider**(*provider*)

allensdk.core.brain_observatory_cache module

allensdk.core.brain_observatory_nwb_data_set module

class allensdk.core.brain_observatory_nwb_data_set.**BrainObservatoryNwbDataSet**(*nwb_file*)

Bases: object

FILE_METADATA_MAPPING = {'age': 'general/subject/age', 'device_string': 'general/devices/2-photon microscope', 'excitation_lambda': 'general/optophysiology/imaging_plane_1/excitation_lambda', 'experiment_container_id': 'general/experiment_container_id', 'fov': 'general/fov', 'generated_by': 'general/generated_by', 'genotype': 'general/subject/genotype', 'imaging_depth': 'general/optophysiology/imaging_plane_1/imaging depth', 'indicator': 'general/optophysiology/imaging_plane_1/indicator', 'ophys_experiment_id': 'general/session_id', 'session_start_time': 'session_start_time', 'session_type': 'general/session_type', 'sex': 'general/subject/sex', 'specimen_name': 'general/specimen_name', 'targeted_structure': 'general/optophysiology/imaging_plane_1/location'}

MOTION_CORRECTION_DATASETS = ['MotionCorrection/2p_image_series/xy_translations', 'MotionCorrection/2p_image_series/xy_translation']

PIPELINE_DATASET = 'brain_observatory_pipeline'

STIMULUS_TABLE_TYPES = {'abstract_feature_series': ['drifting_gratings', 'static_gratings'], 'indexed_time_series': ['natural_scenes', 'locally_sparse_noise', 'locally_sparse_noise_4deg', 'locally_sparse_noise_8deg'], 'repeated_indexed_time_series': ['natural_movie_one', 'natural_movie_two', 'natural_movie_three']}

SUPPORTED_PIPELINE_VERSION = '3.0'

`get_cell_specimen_ids()`

Returns an array of cell IDs for all cells in the file

Returns

cell specimen IDs: list

`get_cell_specimen_indices(cell_specimen_ids)`

Given a list of cell specimen ids, return their index based on their order in this file.

Parameters

cell_specimen_ids: list of cell specimen ids

`get_corrected_fluorescence_traces(cell_specimen_ids=None)`

Returns an array of demixed and neuropil-corrected fluorescence traces for all ROIs and the timestamps for each datapoint

Parameters

cell_specimen_ids: list or array (optional)
List of cell IDs to return traces for. If this is None (default) then all are returned

Returns

timestamps: 2D numpy array
Timestamp for each fluorescence sample

traces: 2D numpy array
Corrected fluorescence traces for each cell

`get_demixed_traces(cell_specimen_ids=None)`

Returns an array of demixed fluorescence traces for all ROIs and the timestamps for each datapoint

Parameters

cell_specimen_ids: list or array (optional)
List of cell IDs to return traces for. If this is None (default) then all are returned

Returns

timestamps: 2D numpy array
Timestamp for each fluorescence sample

traces: 2D numpy array
Demixed fluorescence traces for each cell

`get_dff_traces(cell_specimen_ids=None)`

Returns an array of dF/F traces for all ROIs and the timestamps for each datapoint

Parameters

cell_specimen_ids: list or array (optional)
List of cell IDs to return data for. If this is None (default) then all are returned

Returns

timestamps: 2D numpy array
Timestamp for each fluorescence sample

dF/F: 2D numpy array
dF/F values for each cell

`get_fluorescence_timestamps()`

Returns an array of timestamps in seconds for the fluorescence traces

`get_fluorescence_traces(cell_specimen_ids=None)`

Returns an array of fluorescence traces for all ROI and the timestamps for each datapoint

Parameters

cell_specimen_ids: list or array (optional)
List of cell IDs to return traces for. If this is None (default) then all are returned

Returns

timestamps: 2D numpy array
Timestamp for each fluorescence sample

traces: 2D numpy array
Fluorescence traces for each cell

`get_locally_sparse_noise_stimulus_template(stimulus, mask_off_screen=True)`

Return an array of the stimulus template for the specified stimulus.

Parameters

stimulus: string

Which locally sparse noise stimulus to retrieve. Must be one of:
stimulus_info.LOCALLY_SPARSE_NOISE stimulus_info.LOCALLY_SPARSE_NOISE_4DEG stimulus_info.LOCALLY_SPARSE_NOISE_8DEG

mask_off_screen: boolean
Set off-screen regions of the stimulus to LocallySparseNoise.LSN_OFF_SCREEN.

Returns

tuple: (template, off-screen mask)

`get_max_projection()`

Returns the maximum projection image for the 2P movie.

Returns

max projection: np.ndarray

`get_metadata()`

Returns a dictionary of meta data associated with each experiment, including Cre line, specimen number, visual area imaged, imaging depth

Returns

metadata: dictionary

`get_motion_correction()`

Returns a Panda DataFrame containing the x- and y- translation of each image used for image alignment

`get_neuropil_r(cell_specimen_ids=None)`

Returns a scalar value of r for neuropil correction of flourescence traces

Parameters

cell_specimen_ids: list or array (optional)
List of cell IDs to return traces for. If this is None (default) then results for all are returned

Returns

r: 1D numpy array, len(r)=len(cell_specimen_ids)
Scalar for neuropil subtraction for each cell

`get_neuropil_traces`(*cell_specimen_ids=None*)

Returns an array of neuropil fluorescence traces for all ROIs and the timestamps for each datapoint

Parameters

cell_specimen_ids: list or array (optional)
List of cell IDs to return traces for. If this is None (default) then all are returned

Returns

timestamps: 2D numpy array
Timestamp for each fluorescence sample

traces: 2D numpy array
Neuropil fluorescence traces for each cell

`get_pupil_location`(*as_spherical=True*)

Returns the x, y pupil location.

Parameters

as_spherical
[bool] Whether to return the location as spherical (default) or not. If true, the result is altitude and azimuth in degrees, otherwise it is x, y in centimeters. (0,0) is the center of the monitor.

Returns

(timestamps, location)
Timestamps is an (Nx1) array of timestamps in seconds. Location is an (Nx2) array of spatial location.

`get_pupil_size`()

Returns the pupil area in pixels.

Returns

(timestamps, areas)
Timestamps is an (Nx1) array of timestamps in seconds. Areas is an (Nx1) array of pupil areas in pixels.

`get_roi_ids`()

Returns an array of IDs for all ROIs in the file

Returns

ROI IDs: list

`get_roi_mask`(*cell_specimen_ids=None*)

Returns an array of all the ROI masks

Parameters

cell specimen IDs: list or array (optional)
List of cell IDs to return traces for. If this is None (default) then all are returned

Returns

List of ROI_Mask objects

`get_roi_mask_array`(*cell_specimen_ids=None*)

Return a numpy array containing all of the ROI masks for requested cells. If cell_specimen_ids is omitted, return all masks.

Parameters

cell_specimen_ids: list
List of cell specimen ids. Default None.

Returns

np.ndarray: NxWxH array, where N is number of cells

`get_running_speed`()

Returns the mouse running speed in cm/s

`get_session_type`()

Returns the type of experimental session, presently one of the following: three_session_A, three_session_B, three_session_C

Returns

session type: string

`get_stimulus`(*frame_ind*)

`get_stimulus_epoch_table`()

Returns a pandas dataframe that summarizes the stimulus epoch duration for each acquisition time index in the experiment

Parameters

None

Returns

timestamps: 2D numpy array
Timestamp for each fluorescence sample

traces: 2D numpy array
Fluorescence traces for each cell

`get_stimulus_table`(*stimulus_name*)

Return a stimulus table given a stimulus name

Notes

For more information, see: http://help.brain-map.org/display/observatory/Documentation?preview=/10616846/10813485/VisualCoding_VisualStimuli.pdf

`get_stimulus_template`(*stimulus_name*)

Return an array of the stimulus template for the specified stimulus.

Parameters

stimulus_name: string
Must be one of the strings returned by list_stimuli().

Returns

stimulus table: pd.DataFrame

list_stimuli()

Return a list of the stimuli presented in the experiment.

Returns

stimuli: list of strings

property number_of_cells

Number of cells in the experiment

save_analysis_arrays(**datasets*)

save_analysis_dataframes(**tables*)

property stimulus_search

allensdk.core.brain_observatory_nwb_data_set.**align_running_speed**(*dxcm*, *dxtime*, *timestamps*)

If running speed timestamps differ from fluorescence timestamps, adjust by inserting NaNs to running speed.

Returns

tuple: dxcm, dxtime

allensdk.core.brain_observatory_nwb_data_set.**get_epoch_mask_list**(*st*, *threshold*, *max_cuts=2*)

Convenience function to cut a stim table into multiple epochs

Parameters

- **st** – input stimtable
- **threshold** – threshold on the max duration of a subepoch
- **max_cuts** – maximum number of allowed epochs to cut into

Returns

epoch_mask_list, a list of indices that define the start and end of sub-epochs

allensdk.core.cache_method_utilities module

class allensdk.core.cache_method_utilities.**CachedInstanceMethodMixin**

Bases: object

cache_clear()

Calls *cache_clear* method on all bound methods in this instance (where valid). Intended to clear calls cached with the *memoize* decorator. Note that this will also clear functions decorated with *lru_cache* and *lfu_cache* in this class (or any other function with *cache_clear* attribute).

allensdk.core.cell_types_cache module

class allensdk.core.cell_types_cache.**CellTypesCache**(*cache=True*, *manifest_file=None*, *base_uri=None*)

Bases: *Cache*

Cache class for storing and accessing data from the Cell Types Database. By default, this class will cache any downloaded metadata or files in well known locations defined in a manifest file. This behavior can be disabled.

Parameters

cache: boolean
Whether the class should save results of API queries to locations specified in the manifest file. Queries for files (as opposed to metadata) must have a file location. If caching is disabled, those locations must be specified in the function call (e.g. get_ephys_data(file_name='file.nwb')).

manifest_file: string
File name of the manifest to be read. Default is "cell_types_manifest.json".

Attributes

api: CellTypesApi instance
The object used for making API queries related to the Cell Types Database

`CELLS_KEY = 'CELLS'`

`EPHYS_DATA_KEY = 'EPHYS_DATA'`

`EPHYS_FEATURES_KEY = 'EPHYS_FEATURES'`

`EPHYS_SWEEPS_KEY = 'EPHYS_SWEEPS'`

`MANIFEST_VERSION = '1.1'`

`MARKER_KEY = 'MARKER'`

`MORPHOLOGY_FEATURES_KEY = 'MORPHOLOGY_FEATURES'`

`RECONSTRUCTION_KEY = 'RECONSTRUCTION'`

`build_manifest`(*file_name*)

Construct a manifest for this Cache class and save it in a file.

Parameters

file_name: string
File location to save the manifest.

`get_all_features`(*dataframe=False*, *require_reconstruction=True*)

Download morphology and electrophysiology features for all cells and merge them into a single table.

Parameters

dataframe: boolean
Return the output as a Pandas DataFrame. If False, return a list of dictionaries.

require_reconstruction: boolean
Only return ephys and morphology features for cells that have reconstructions. Default True.

`get_cells`(*file_name=None*, *require_morphology=False*, *require_reconstruction=False*, *reporter_status=None*, *species=None*, *simple=True*)

Download metadata for all cells in the database and optionally return a subset filtered by whether or not they have a morphology or reconstruction.

Parameters

file_name: string
File name to save/read the cell metadata as JSON. If file_name is None, the file_name will be pulled out of the manifest. If caching is disabled, no file will be saved. Default is None.

require_morphology: boolean
Filter out cells that have no morphological images.

require_reconstruction: boolean
Filter out cells that have no morphological reconstructions.

reporter_status: list
Filter for cells that have one or more cell reporter statuses.

species: list
Filter for cells that belong to one or more species. If None, return all. Must be one of [CellTypesApi.MOUSE, CellTypesApi.HUMAN].

`get_ephys_data`(*specimen_id*, *file_name=None*)

Download electrophysiology traces for a single cell in the database.

Parameters

specimen_id: int
The ID of a cell specimen to download.

file_name: string
File name to save/read the ephys features metadata as CSV. If file_name is None, the file_name will be pulled out of the manifest. If caching is disabled, no file will be saved. Default is None.

Returns

NwbDataSet
A class instance with helper methods for retrieving stimulus and response traces out of an NWB file.

`get_ephys_features`(*dataframe=False*, *file_name=None*)

Download electrophysiology features for all cells in the database.

Parameters

file_name: string
File name to save/read the ephys features metadata as CSV. If file_name is None, the file_name will be pulled out of the manifest. If caching is disabled, no file will be saved. Default is None.

dataframe: boolean
Return the output as a Pandas DataFrame. If False, return a list of dictionaries.

`get_ephys_sweeps`(*specimen_id*, *file_name=None*)

Download sweep metadata for a single cell specimen.

Parameters

specimen_id: int
ID of a cell.

`get_morphology_features`(*dataframe=False*, *file_name=None*)

Download morphology features for all cells with reconstructions in the database.

Parameters

file_name: string
File name to save/read the ephys features metadata as CSV. If file_name is None, the file_name will be pulled out of the manifest. If caching is disabled, no file will be saved. Default is None.

dataframe: boolean
Return the output as a Pandas DataFrame. If False, return a list of dictionaries.

`get_reconstruction`(*specimen_id*, *file_name=None*)

Download and open a reconstruction for a single cell in the database.

Parameters

specimen_id: int
The ID of a cell specimen to download.

file_name: string
File name to save/read the reconstruction SWC. If file_name is None, the file_name will be pulled out of the manifest. If caching is disabled, no file will be saved. Default is None.

Returns

Morphology
A class instance with methods for accessing morphology compartments.

`get_reconstruction_markers`(*specimen_id*, *file_name=None*)

Download and open a reconstruction marker file for a single cell in the database.

Parameters

specimen_id: int
The ID of a cell specimen to download.

file_name: string
File name to save/read the reconstruction marker. If file_name is None, the file_name will be pulled out of the manifest. If caching is disabled, no file will be saved. Default is None.

Returns

Morphology
A class instance with methods for accessing morphology compartments.

`class allensdk.core.cell_types_cache.ReporterStatus`

Bases: `object`

Valid strings for filtering by cell reporter status.

`INDETERMINATE = None`

`NA = None`

`NEGATIVE = 'negative'`

`POSITIVE = 'positive'`

allensdk.core.dat_utilities module

class allensdk.core.dat_utilities.**DatUtilities**

Bases: object

classmethod save_voltage(*output_path*, *v*, *t*)

Save a single voltage output result into a simple text format.

The output file is one t v pair per line.

Parameters

output_path
[string] file name for output

v
[numpy array] voltage

t
[numpy array] time

allensdk.core.dataframe_utils module

allensdk.core.dataframe_utils.**INT_NULL = -99**

A collection of utilities to manipulate pandas DataFrames.

allensdk.core.dataframe_utils.**enforce_df_column_order**(*input_df: DataFrame*, *column_order: List[str]*) → DataFrame

Return the data frame but with columns ordered.

Parameters

input_df
[pandas.DataFrame] Data frame with columns to be ordered.

column_order
[list of str] Ordering of column names to enforce. Columns not specified are shifted to the end of the order but retain their order amongst others not specified. If a specified column is not in the DataFrame it is ignored.

Returns

output_df
[pandas.DataFrame] DataFrame the same as the input but with columns reordered.

allensdk.core.dataframe_utils.**enforce_df_int_typing**(*input_df: DataFrame*, *int_columns: List[str]*, *use_pandas_type: object = False*) → DataFrame

Enforce integer typing for columns that may have lost int typing when combined into the final DataFrame.

Parameters

input_df
[pandas.DataFrame] DataFrame with typing to enforce.

int_columns
[list of str] Columns to enforce int typing and fill any NaN/None values with the value set in INT_NULL in this file. Requested columns not in the dataframe are ignored.

use_pandas_type
[bool] Instead of filling with the value INT_NULL to enforce integer typing, use the pandas type Int64. This type can have issues converting to numpy/array type values.

Returns

output_df
[pandas.DataFrame] DataFrame specific columns hard typed to Int64 to allow NA values without resorting to float type.

allensdk.core.dataframe_utils.**patch_df_from_other**(*target_df: DataFrame*, *source_df: DataFrame*, *columns_to_patch: List[str]*, *index_column: str*) → DataFrame

Overwrite column values in target_df from column values in source_df in rows where the two dataframes share a value of index_column.

Parameters

target_df: pd.DataFrame
The dataframe whose columns will get overwritten

source_df: pd.DataFrame
The dataframe from which correct values are to be read

columns_to_patch: List[str]
The columns to be overwritten

index_column: str
The column to join the dataframes on

Returns

patched_df: pd.DataFrame
target_df except with the specified columns and rows overwritten.

Notes

If any of the columns_to_patch are not in target_df, they will be added.

This function starts by creating a copy of target_df, so it will not alter the argument in-place.

allensdk.core.dataframe_utils.**return_one_dataframe_row_only**(*input_table: DataFrame*, *index_value: int*, *table_name: str*) → Series

Lookup and return one and only one row from the DataFrame returning an informative error if no or multiple rows are returned for a given index.

This method is used mainly to return a more informative error when attempting to retrieve metadata from the values behavior cache metadata tables.

Parameters

input_table
[pandas.DataFrame] Input dataframe to retrieve row from.

index_value
[int]

Index of the row to return. Must match an index in the input
dataframe/table. i.e. in the case of ecephys_session_table or

behavior_session_table.

table_name
[str] Name of the table being returned. Used to output the table name in case of error.

Returns

row
[pandas.Series] Row corresponding to the input index.

allensdk.core.exceptions module

exception allensdk.core.exceptions.**DataFrameIndexError**(*msg*, *caught_exception=None*)

Bases: LookupError

More verbose method for accessing invalid rows or columns in a dataframe. Should be used when an index error is thrown on a dataframe.

exception allensdk.core.exceptions.**DataFrameKeyError**(*msg*, *caught_exception=None*)

Bases: LookupError

More verbose method for accessing invalid rows or columns in a dataframe. Should be used when a keyerror is thrown on a dataframe.

exception allensdk.core.exceptions.**MissingDataError**

Bases: ValueError

allensdk.core.h5_utilities module

allensdk.core.h5_utilities.**decode_bytes**(*bytes_dataset*, *encoding='UTF-8'*)

Convert the elements of a dataset of bytes to str

allensdk.core.h5_utilities.**h5_object_matcher_relname_in**(*relnames*, *h5_object_name*, *h5_object*)

Asks if an h5 object's relative name (the final section of its absolute name) is contained within a provided array

Parameters

relnames
[array-like] Relative names against which to match

h5_object_name
[str] Full name (path from origin) of h5 object

h5_object
[h5py.Group, h5py.Dataset] Check this object's relative name

Returns

bool
whether the match succeeded

h5_object
[h5py.group, h5py.Dataset] the argued object

allensdk.core.h5_utilities.**keyed_locate_h5_objects**(*matcher_cbs*, *h5_file*, *start_node=None*)

Traverse an h5 file and build up a dictionary mapping supplied keys to located objects

allensdk.core.h5_utilities.**load_datasets_by_relnames**(*relnames*, *h5_file*, *start_node*)

A convenience function for finding and loading into memory one or more datasets from an h5 file

allensdk.core.h5_utilities.**locate_h5_objects**(*matcher_cb*, *h5_file*, *start_node=None*)

Traverse an h5 file and return objects matching supplied criteria

allensdk.core.h5_utilities.**traverse_h5_file**(*callback*, *h5_file*, *start_node=None*)

Traverse an h5 file and apply a callback to each node

allensdk.core.json_utilities module

class allensdk.core.json_utilities.**JsonComments**

Bases: object

classmethod read_file(*file_name*)

classmethod read_string(*json_string*)

classmethod remove_comments(*json_string*)

Strip single and multiline javascript-style comments.

Parameters

json
[string] Json string with javascript-style comments.

Returns

string
Copy of the input with comments removed.

Note: A JSON decoder MAY accept and ignore comments.

classmethod remove_multiline_comments(*json_string*)

Rebuild input without substrings matching /*...*/.

Parameters

json_string
[string] may or may not contain multiline comments.

Returns

string
Copy of the input without the comments.

allensdk.core.json_utilities.**json_handler**(*obj*)

Used by write_json convert a few non-standard types to things that the json package can handle.

allensdk.core.json_utilities.**read**(*file_name*)

Shortcut reading JSON from a file.

allensdk.core.json_utilities.**read_url**(*url*, *method='POST'*)

allensdk.core.json_utilities.**read_url_get**(*url*)

Transform a JSON contained in a file into an equivalent nested python dict.

Parameters

url
[string] where to get the json.

Returns

dict
Python version of the input

Note: if the input is a bare array or literal, for example, the output will be of the corresponding type.

allensdk.core.json_utilities.**read_url_post**(*url*)

Transform a JSON contained in a file into an equivalent nested python dict.

Parameters

url
[string] where to get the json.

Returns

dict
Python version of the input

Note: if the input is a bare array or literal, for example, the output will be of the corresponding type.

allensdk.core.json_utilities.**write**(*file_name*, *obj*)

Shortcut for writing JSON to a file. This also takes care of serializing numpy and data types.

allensdk.core.json_utilities.**write_string**(*obj*)

Shortcut for writing JSON to a string. This also takes care of serializing numpy and data types.

allensdk.core.mouse_connectivity_cache module

class allensdk.core.mouse_connectivity_cache.**MouseConnectivityCache**(*resolution=None*, *cache=True*, *manifest_file=None*, *ccf_version=None*, *base_uri=None*, *version=None*)

Bases: *ReferenceSpaceCache*

Cache class for storing and accessing data related to the adult mouse Connectivity Atlas. By default, this class will cache any downloaded metadata or files in well known locations defined in a manifest file. This behavior can be disabled.

Parameters

resolution: int
Resolution of grid data to be downloaded when accessing projection volume, the annotation volume, and the annotation volume. Must be one of (10, 25, 50, 100). Default is 25.

ccf_version: string
Desired version of the Common Coordinate Framework. This affects the annotation volume (get_annotation_volume) and structure masks (get_structure_mask). Must be one of (MouseConnectivityApi.CCF_2015, MouseConnectivityApi.CCF_2016). Default: MouseConnectivityApi.CCF_2016

cache: boolean
Whether the class should save results of API queries to locations specified in the manifest file. Queries for files (as opposed to metadata) must have a file location.

If caching is disabled, those locations must be specified in the function call (e.g. get_projection_density(file_name='file.nrrd')).

manifest_file: string
File name of the manifest to be read. Default is "mouse_connectivity_manifest.json".

Attributes

resolution: int
Resolution of grid data to be downloaded when accessing projection volume, the annotation volume, and the annotation volume. Must be one of (10, 25, 50, 100). Default is 25.

api: MouseConnectivityApi instance
Used internally to make API queries.

`ALIGNMENT3D_KEY = 'ALIGNMENT3D'`

`DATA_MASK_KEY = 'DATA_MASK'`

`DEFAULT_STRUCTURE_SET_IDS = (167587189,)`

`DEFORMATION_FIELD_HEADER_KEY = 'DEFORMATION_FIELD_HEADER'`

`DEFORMATION_FIELD_VOXEL_KEY = 'DEFORMATION_FIELD_VOXELS'`

`DFMFLD_RESOLUTIONS = (25,)`

`EXPERIMENTS_KEY = 'EXPERIMENTS'`

`INJECTION_DENSITY_KEY = 'INJECTION_DENSITY'`

`INJECTION_FRACTION_KEY = 'INJECTION_FRACTION'`

`MANIFEST_VERSION = 1.3`

`PROJECTION_DENSITY_KEY = 'PROJECTION_DENSITY'`

`STRUCTURE_UNIONIZES_KEY = 'STRUCTURE_UNIONIZES'`

`SUMMARY_STRUCTURE_SET_ID = 167587189`

`add_manifest_paths`(*manifest_builder*)

Construct a manifest for this Cache class and save it in a file.

Parameters

file_name: string
File location to save the manifest.

`property default_structure_ids`

`filter_experiments`(*experiments*, *cre=None*, *injection_structure_ids=None*)

Take a list of experiments and filter them by cre status and injection structure.

Parameters

cre: boolean or list
If True, return only cre-positive experiments. If False, return only cre-negative experiments. If None, return all experients. If list, return all experiments with cre line names in the supplied list. Default None.

injection_structure_ids: list
Only return experiments that were injected in the structures provided here. If None, return all experiments. Default None.

`filter_structure_unionizes`(*unionizes*, *is_injection=None*, *structure_ids=None*, *include_descendants=False*, *hemisphere_ids=None*)

Take a list of unionzes and return a subset of records filtered by injection status, structure, and hemisphere.

Parameters

is_injection: boolean
If True, only return unionize records that disregard non-injection pixels. If False, only return unionize records that disregard injection pixels. If None, return all records. Default None.

structure_ids: list
Only return unionize records for a set of structures. If None, return all records. Default None.

include_descendants: boolean
Include all descendant records for specified structures. Default False.

hemisphere_ids: list
Only return unionize records that disregard pixels outside of a hemisphere. or set of hemispheres. Left = 1, Right = 2, Both = 3. If None, include all records [1, 2, 3]. Default None.

`get_affine_parameters`(*section_data_set_id*, *direction='trv'*, *file_name=None*)

Extract the parameters of the 3D affine tranformation mapping this section data set's image-space stack to CCF-space (or vice-versa).

Parameters

section_data_set_id
[int] download the parameters for this data set.

direction
[str, optional]

Valid options are:

trv
["transform from reference to volume". Maps CCF points] to image space points. If you are resampling data into CCF, this is the direction you want.

tvr
["transform from volume to reference". Maps image] space points to CCF points.

file_name
[str] If provided, store the downloaded file here.

Returns

alignment
[numpy.ndarray]

4 X 3 matrix. In order to transform a point [X_1, X_2, X_3] run
np.dot([X_1, X_2, X_3, 1], alignment). In

to build a SimpleITK affine transform run:
transform = sitk.AffineTransform(3) transform.SetParameters(alignment.flatten())

`get_data_mask`(*experiment_id*, *file_name=None*)

Read a data mask volume for a single experiment. Download it first if it doesn't exist. Data mask is a binary mask of voxels that have valid data. Only use valid data in analysis!

Parameters

experiment_id: int
ID of the experiment to download/read. This corresponds to section_data_set_id in the API.

file_name: string
File name to store the template volume. If it already exists, it will be read from this file. If file_name is None, the file_name will be pulled out of the manifest. Default is None.

`get_deformation_field`(*section_data_set_id*, *header_path=None*, *voxel_path=None*)

Extract the local alignment parameters for this dataset. This a 3D vector image (3 components) describing a deformable local mapping from CCF voxels to this section data set's affine-aligned image stack.

Parameters

section_data_set_id
[int] Download the deformation field for this data set

header_path
[str, optional] If supplied, the deformation field header will be downloaded to this path.

voxel_path
[str, optiona] If supplied, the deformation field voxels will be downloaded to this path.

Returns

numpy.ndarray
3D X 3 component vector array (origin 0, 0, 0; 25-micron isometric resolution) defining a deformable transformation from CCF-space to affine-transformed image space.

`get_experiment_structure_unionizes`(*experiment_id*, *file_name=None*, *is_injection=None*, *structure_ids=None*, *include_descendants=False*, *hemisphere_ids=None*)

Retrieve the structure unionize data for a specific experiment. Filter by structure, injection status, and hemisphere.

Parameters

experiment_id: int
ID of the experiment of interest. Corresponds to section_data_set_id in the API.

file_name: string
File name to save/read the experiments list. If file_name is None, the file_name will be pulled out of the manifest. If caching is disabled, no file will be saved. Default is None.

is_injection: boolean
If True, only return unionize records that disregard non-injection pixels. If False, only return unionize records that disregard injection pixels. If None, return all records. Default None.

structure_ids: list
Only return unionize records for a specific set of structures. If None, return all records. Default None.

include_descendants: boolean
Include all descendant records for specified structures. Default False.

hemisphere_ids: list
Only return unionize records that disregard pixels outside of a hemisphere. or set of hemispheres. Left = 1, Right = 2, Both = 3. If None, include all records [1, 2, 3]. Default None.

`get_experiments`(*dataframe=False*, *file_name=None*, *cre=None*, *injection_structure_ids=None*)

Read a list of experiments that match certain criteria. If caching is enabled, this will save the whole (unfiltered) list of experiments to a file.

Parameters

dataframe: boolean
Return the list of experiments as a Pandas DataFrame. If False, return a list of dictionaries. Default False.

file_name: string
File name to save/read the structures table. If file_name is None, the file_name will be pulled out of the manifest. If caching is disabled, no file will be saved. Default is None.

cre: boolean or list
If True, return only cre-positive experiments. If False, return only cre-negative experiments. If None, return all experients. If list, return all experiments with cre line names in the supplied list. Default None.

injection_structure_ids: list
Only return experiments that were injected in the structures provided here. If None, return all experiments. Default None.

`get_injection_density`(*experiment_id*, *file_name=None*)

Read an injection density volume for a single experiment. Download it first if it doesn't exist. Injection density is the proportion of projecting pixels in a grid voxel only including pixels that are part of the injection site in [0,1].

Parameters

experiment_id: int
ID of the experiment to download/read. This corresponds to section_data_set_id in the API.

file_name: string
File name to store the template volume. If it already exists, it will be read from this file. If file_name is None, the file_name will be pulled out of the manifest. Default is None.

`get_injection_fraction`(*experiment_id*, *file_name=None*)

Read an injection fraction volume for a single experiment. Download it first if it doesn't exist. Injection fraction is the proportion of pixels in the injection site in a grid voxel in [0,1].

Parameters

experiment_id: int
ID of the experiment to download/read. This corresponds to section_data_set_id in the API.

file_name: string
File name to store the template volume. If it already exists, it will be read from this

file. If file_name is None, the file_name will be pulled out of the manifest. Default is None.

`get_projection_density`(*experiment_id*, *file_name=None*)

Read a projection density volume for a single experiment. Download it first if it doesn't exist. Projection density is the proportion of of projecting pixels in a grid voxel in [0,1].

Parameters

experiment_id: int
ID of the experiment to download/read. This corresponds to section_data_set_id in the API.

file_name: string
File name to store the template volume. If it already exists, it will be read from this file. If file_name is None, the file_name will be pulled out of the manifest. Default is None.

`get_projection_matrix`(*experiment_ids*, *projection_structure_ids=None*, *hemisphere_ids=None*, *parameter='projection_volume'*, *dataframe=False*)

`get_structure_unionizes`(*experiment_ids*, *is_injection=None*, *structure_ids=None*, *include_descendants=False*, *hemisphere_ids=None*)

Get structure unionizes for a set of experiment IDs. Filter the results by injection status, structure, and hemisphere.

Parameters

experiment_ids: list
List of experiment IDs. Corresponds to section_data_set_id in the API.

is_injection: boolean
If True, only return unionize records that disregard non-injection pixels. If False, only return unionize records that disregard injection pixels. If None, return all records. Default None.

structure_ids: list
Only return unionize records for a specific set of structures. If None, return all records. Default None.

include_descendants: boolean
Include all descendant records for specified structures. Default False.

hemisphere_ids: list
Only return unionize records that disregard pixels outside of a hemisphere. or set of hemispheres. Left = 1, Right = 2, Both = 3. If None, include all records [1, 2, 3]. Default None.

`rank_structures`(*experiment_ids*, *is_injection*, *structure_ids=None*, *hemisphere_ids=None*, *rank_on='normalized_projection_volume'*, *n=5*, *threshold=0.01*)

Produces one or more (per experiment) ranked lists of brain structures, using a specified data field.

Parameters

experiment_ids
[list of int] Obtain injection_structures for these experiments.

is_injection
[boolean] Use data from only injection (or non-injection) unionizes.

structure_ids
[list of int, optional] Consider only these structures. It is a good idea to make sure that these structures are not spatially overlapping; otherwise your results will contain redundant information. Defaults to the summary structures - a brain-wide list of nonoverlapping mid-level structures.

hemisphere_ids
[list of int, optional] Consider only these hemispheres (1: left, 2: right, 3: both). Like with structures, you might get redundant results if you select overlapping options. Defaults to [1, 2].

rank_on
[str, optional] Rank unionize data using this field (descending). Defaults to normalized_projection_volume.

n
[int, optional] Return only the top n structures.

threshold
[float, optional] Consider only records whose data value - specified by the rank_on parameter - exceeds this value.

Returns

list
Each element (1 for each input experiment) is a list of dictionaries. The dictionaries describe the top injection structures in descending order. They are specified by their structure and hemisphere id fields and additionally report the value specified by the rank_on parameter.

allensdk.core.nwb_data_set module

class allensdk.core.nwb_data_set.**NwbDataSet**(*file_name*, *spike_time_key=None*)

Bases: object

A very simple interface for exracting electrophysiology data from an NWB file.

DEPRECATED_SPIKE_TIMES = 'aibs_spike_times'

SPIKE_TIMES = 'spike_times'

fill_sweep_responses(*fill_value=0.0*, *sweep_numbers=None*, *extend_experiment=False*)

Fill sweep response arrays with a single value.

Parameters

fill_value: float
Value used to fill sweep response array

sweep_numbers: list
List of integer sweep numbers to be filled (default all sweeps)

extend_experiment: bool
If True, extend experiment epoch length to the end of the sweep (undo any truncation)

get_experiment_sweep_numbers()

Get all of the sweep numbers for experiment epochs in the file, not including test sweeps.

`get_pipeline_version()`

Returns the AI pipeline version number, stored in the metadata field 'generated_by'. If that field is missing, version 0.0 is returned.

Returns

int tuple: (major, minor)

`get_spike_times(sweep_number, key=None)`

Return any spike times stored in the NWB file for a sweep.

Parameters

sweep_number: int
index to access

key
[string] label where the spike times are stored (default NwbDataSet.SPIKE_TIMES)

Returns

list
list of spike times in seconds relative to the start of the sweep

`get_sweep(sweep_number)`

Retrieve the stimulus, response, index_range, and sampling rate for a particular sweep. This method hides the NWB file's distinction between a "Sweep" and an "Experiment". An experiment is a subset of of a sweep that excludes the initial test pulse. It also excludes any erroneous response data at the end of the sweep (usually for ramp sweeps, where recording was terminated mid-stimulus).

Some sweeps do not have an experiment, so full data arrays are returned. Sweeps that have an experiment return full data arrays (include the test pulse) with any erroneous data trimmed from the back of the sweep.

Parameters

sweep_number: int

Returns

dict
A dictionary with 'stimulus', 'response', 'index_range', and 'sampling_rate' elements. The index range is a 2-tuple where the first element indicates the end of the test pulse and the second index is the end of valid response data.

`get_sweep_metadata(sweep_number)`

Retrieve the sweep level metadata associated with each sweep. Includes information on stimulus parameters like its name and amplitude as well as recording quality metadata, like access resistance and seal quality.

Parameters

sweep_number: int

Returns

dict
A dictionary with 'aibs_stimulus_amplitude_pa', 'aibs_stimulus_name', 'gain', 'initial_access_resistance', 'seal' elements. These specific fields are ones encoded in the original AIBS in vitro .nwb files.

`get_sweep_numbers()`

Get all of the sweep numbers in the file, including test sweeps.

set_spike_times(*sweep_number*, *spike_times*, *key=None*)

Set or overwrite the spikes times for a sweep.

Parameters

sweep_number
[int] index to access

key
[string] where the times are stored (default NwbDataSet.SPIKE_TIME)

spike_times: np.array
array of spike times in seconds

set_sweep(*sweep_number*, *stimulus*, *response*)

Overwrite the stimulus or response of an NWB file. If the supplied arrays are shorter than stored arrays, they are padded with zeros to match the original data size.

Parameters

sweep_number: int
stimulus: np.array
Overwrite the stimulus with this array. If None, stimulus is unchanged.

response: np.array
Overwrite the response with this array. If None, response is unchanged.

allensdk.core.obj_utilities module

allensdk.core.obj_utilities.**parse_obj**(*lines*)

Parse a wavefront obj file into a triplet of vertices, normals, and faces. This parser is specific to obj files generated from our annotation volumes

Parameters

lines
[list of str] Lines of input obj file

Returns

vertices
[np.ndarray] Dimensions are (nSamples, nCoordinates=3). Locations in the reference space of vertices

vertex_normals
[np.ndarray] Dimensions are (nSample, nElements=3). Vectors normal to vertices.

face_vertices
[np.ndarray] Dimensions are (sample, nVertices=3). References are given in indices (0-indexed here, but 1-indexed in the file) of vertices that make up each face.

face_normals
[np.ndarray] Dimensions are (sample, nNormals=3). References are given in indices (0-indexed here, but 1-indexed in the file) of vertex normals that make up each face.

Notes

This parser is specialized to the obj files that the Allen Institute for Brain Science generates from our own structure annotations.

allensdk.core.obj_utilities.**read_obj**(*path*)

allensdk.core.ontology module

class allensdk.core.ontology.**Ontology**(*df*)

Bases: object

Note: Deprecated from 0.12.5 *Ontology* has been replaced by *StructureTree*.

get_child_ids(*structure_ids*)

Find the set of ids that are immediate children of one or more structures.

Parameters

structure_ids: iterable
Any iterable type that contains structure ids that can be cast to integers.

Returns

set
Set of child structure ids

get_children(*structure_ids*)

Find the set of structures that are immediate children of one or more structures.

Parameters

structure_ids: iterable
Any iterable type that contains structure ids that can be cast to integers.

Returns

pandas.DataFrame
Set of child structures

get_descendant_ids(*structure_ids*)

Find the set of the ids of structures that are descendants of one or more structures. The returned set will include the input structure ids.

Parameters

structure_ids: iterable
Any iterable type that contains structure ids that can be cast to integers.

Returns

set
Set of descendant structure ids.

get_descendants(*structure_ids*)

Find the set of structures that are descendants of one or more structures. The returned set will include the input structures.

Parameters

structure_ids: iterable
Any iterable type that contains structure ids that can be cast to integers.

Returns

pandas.DataFrame
Set of descendant structures.

`structure_descends_from`(*child_id*, *parent_id*)

Return whether one structure id is a descendant of another structure id.

allensdk.core.ophys_experiment_session_id_mapping module

allensdk.core.pickle_utils module

`allensdk.core.pickle_utils.load_and_sanitize_pickle`(*pickle_path: str | Path*) → Any

Load the data from a pickle file and pass it through sanitize_pickle_data, so that all bytes in the data are cast to strings.

Parameters

pickle_path: Union[str, pathlib.Path]
Path to the pickle file

Returns

pickle_data: Any
The data that was in the pickle file

Notes

Because sanitize_pickle_data alters the data in-place, this method encapsulates loading and sanitizing so that users do not think they have a pre-sanitization copy of the data available.

allensdk.core.reference_space module

`class allensdk.core.reference_space.ReferenceSpace`(*structure_tree*, *annotation*, *resolution*)

Bases: `object`

`static check_and_write`(*base_dir*, *structure_id*, *fn*)

A many_structure_masks callback that writes the mask to a nrrd file if the file does not already exist.

`check_coverage`(*structure_ids*, *domain_mask*)

Determines whether a spatial domain is completely covered by structures in a set.

Parameters

structure_ids
[list of int] Specifies the set of structures to check.

domain_mask
[numpy ndarray] Same shape as annotation. 1 inside the mask, 0 out. Specifies spatial domain.

Returns

numpy ndarray
1 where voxels are missing from the candidate, 0 where the candidate exceeds the domain

`direct_voxel_counts()`

Determines the number of voxels directly assigned to one or more structures.

Returns

dict
Keys are structure ids, values are the number of voxels directly assigned to those structures.

`property direct_voxel_map`

`downsample(target_resolution)`

Obtain a smaller reference space by downsampling

Parameters

target_resolution
[tuple of numeric] Resolution in microns of the output space.

interpolator
[string] Method used to interpolate the volume. Currently only 'nearest' is supported

Returns

ReferenceSpace
A new ReferenceSpace with the same structure tree and a downsampled annotation.

`export_itksnap_labels(id_type=<class 'numpy.uint16'>, label_description_kwargs=None)`

Produces itksnap labels, remapping large ids if needed.

Parameters

id_type
[np.integer, optional] Used to determine the type of the output annotation and whether ids need to be remapped to smaller values.

label_description_kwargs
[dict, optional] Keyword arguments passed to StructureTree.export_label_description

Returns

np.ndarray
Annotation volume, remapped if needed

pd.DataFrame
label_description dataframe

`get_slice_image(axis, position, cmap=None)`

Produce a AxBx3 RGB image from a slice in the annotation

Parameters

axis
[int] Along which to slice the annotation volume. 0 is coronal, 1 is horizontal, and 2 is sagittal.

position
[int] In microns. Take the slice from this far along the specified axis.

cmap
[dict, optional] Keys are structure ids, values are rgb triplets. Defaults to structure rgb_triplets.

Returns

np.ndarray
RGB image array.

Notes

If you assign a custom colormap, make sure that you take care of the background in addition to the structures.

`make_structure_mask`(*structure_ids*, *direct_only=False*)

Return an indicator array for one or more structures

Parameters

structure_ids
[list of int] Make a mask that indicates the union of these structures' voxels

direct_only
[bool, optional] If True, only include voxels directly assigned to a structure in the mask. Otherwise include voxels assigned to descendants.

Returns

numpy ndarray
Same shape as annotation. 1 inside mask, 0 outside.

`many_structure_masks`(*structure_ids*, *output_cb=None*, *direct_only=False*)

Build one or more structure masks and do something with them

Parameters

structure_ids
[list of int] Specify structures to be masked

output_cb
[function, optional] Must have the following signature: output_cb(structure_id, fn). On each requested id, fn will be curried to make a mask for that id. Defaults to returning the structure id and mask.

direct_only
[bool, optional] If True, only include voxels directly assigned to a structure in the mask. Otherwise include voxels assigned to descendants.

Yields

Return values of output_cb called on each structure_id, structure_mask pair.

Notes

output_cb is called on every yield, so any side-effects (such as writing to a file) will be carried out regardless of what you do with the return values. You do actually have to iterate through the output, though.

`remove_unassigned`(*update_self=True*)

Obtains a structure tree consisting only of structures that have at least one voxel in the annotation.

Parameters

update_self
[bool, optional] If True, the contained structure tree will be replaced,

Returns

list of dict
elements are filtered structures

`static return_mask_cb`(*structure_id*, *fn*)

A basic callback for many_structure_masks

`total_voxel_counts`()

Determines the number of voxels assigned to a structure or its descendants

Returns

dict
Keys are structure ids, values are the number of voxels assigned to structures' descendants.

`property total_voxel_map`

`validate_structures`(*structure_ids*, *domain_mask*)

Determines whether a set of structures produces an exact and nonoverlapping tiling of a spatial domain

Parameters

structure_ids
[list of int] Specifies the set of structures to check.

domain_mask
[numpy ndarray] Same shape as annotation. 1 inside the mask, 0 out. Specifies spatial domain.

Returns

set
Ids of structures that are the ancestors of other structures in the supplied set.

numpy ndarray
Indicator for missing voxels.

`write_itksnap_labels`(*annotation_path*, *label_path*, ***kwargs*)

Generate a label file (nrrd) and a label_description file (csv) for use with ITKSnap

Parameters

annotation_path
[str] write generated label file here

label_path
[str] write generated label_description file here

****kwargs**
will be passed to self.export_itksnap_labels

allensdk.core.reference_space_cache module

class allensdk.core.reference_space_cache.**ReferenceSpaceCache**(*resolution*, *reference_space_key*, ***kwargs*)

Bases: *Cache*

ANNOTATION_KEY = 'ANNOTATION'

MANIFEST_VERSION = 1.2

REFERENCE_SPACE_VERSION_KEY = 'REFERENCE_SPACE_VERSION'

STRUCTURES_KEY = 'STRUCTURES'

STRUCTURE_MASK_KEY = 'STRUCTURE_MASK'

STRUCTURE_MESH_KEY = 'STRUCTURE_MESH'

STRUCTURE_TREE_KEY = 'STRUCTURE_TREE'

TEMPLATE_KEY = 'TEMPLATE'

add_manifest_paths(*manifest_builder*)

Construct a manifest for this Cache class and save it in a file.

Parameters

file_name: string
File location to save the manifest.

get_annotation_volume(*file_name=None*)

Read the annotation volume. Download it first if it doesn't exist.

Parameters

file_name: string
File name to store the annotation volume. If it already exists, it will be read from this file. If file_name is None, the file_name will be pulled out of the manifest. Default is None.

get_reference_space(*structure_file_name=None*, *annotation_file_name=None*)

Build a ReferenceSpace from this cache's annotation volume and structure tree. The ReferenceSpace does operations that relate brain structures to spatial domains.

Parameters

structure_file_name: string
File name to save/read the structures table. If file_name is None, the file_name will be pulled out of the manifest. If caching is disabled, no file will be saved. Default is None.

annotation_file_name: string
File name to store the annotation volume. If it already exists, it will be read from this file. If file_name is None, the file_name will be pulled out of the manifest. Default is None.

get_structure_mask(*structure_id*, *file_name=None*, *annotation_file_name=None*)

Read a 3D numpy array shaped like the annotation volume that has non-zero values where voxels belong to a particular structure. This will take care of identifying substructures.

Parameters

structure_id: int
ID of a structure.

file_name: string
File name to store the structure mask. If it already exists, it will be read from this file. If file_name is None, the file_name will be pulled out of the manifest. Default is None.

annotation_file_name: string
File name to store the annotation volume. If it already exists, it will be read from this file. If file_name is None, the file_name will be pulled out of the manifest. Default is None.

Notes

This method downloads structure masks from the Allen Institute. To make your own locally, see ReferenceSpace.many_structure_masks.

get_structure_mesh(*structure_id*, *file_name=None*)

Obtain a 3D mesh specifying the surface of an annotated structure.

Parameters

structure_id: int
ID of a structure.

file_name: string
File name to store the structure mesh. If it already exists, it will be read from this file. If file_name is None, the file_name will be pulled out of the manifest. Default is None.

Returns

vertices
[np.ndarray] Dimensions are (nSamples, nCoordinates=3). Locations in the reference space of vertices

vertex_normals
[np.ndarray] Dimensions are (nSample, nElements=3). Vectors normal to vertices.

face_vertices
[np.ndarray] Dimensions are (sample, nVertices=3). References are given in indices (0-indexed here, but 1-indexed in the file) of vertices that make up each face.

face_normals
[np.ndarray] Dimensions are (sample, nNormals=3). References are given in indices (0-indexed here, but 1-indexed in the file) of vertex normals that make up each face.

Notes

These meshes are meant for 3D visualization and as such have been smoothed. If you are interested in performing quantative analyses, we recommend that you use the structure masks instead.

get_structure_tree(*file_name=None*, *structure_graph_id=1*)

Read the list of adult mouse structures and return an StructureTree instance.

Parameters

file_name: string
File name to save/read the structures table. If file_name is None, the file_name will be pulled out of the manifest. If caching is disabled, no file will be saved. Default is None.

structure_graph_id: int
Build a tree using structure only from the identified structure graph.

get_template_volume(*file_name=None*)

Read the template volume. Download it first if it doesn't exist.

Parameters

file_name: string
File name to store the template volume. If it already exists, it will be read from this file. If file_name is None, the file_name will be pulled out of the manifest. Default is None.

classmethod validate_structure_id(*structure_id*)

classmethod validate_structure_ids(*structure_ids*)

allensdk.core.simple_tree module

class allensdk.core.simple_tree.**SimpleTree**(*nodes*, *node_id_cb*, *parent_id_cb*)

Bases: object

ancestor_ids(*node_ids*)

Obtain the ids of one or more nodes' ancestors

Parameters

node_ids
[list of hashable] Items are ids of nodes whose ancestors you wish to find.

Returns

list of list of hashable
Items are lists of input nodes' ancestors' ids.

Notes

Given the tree: A -> B -> C

`-> D

The ancestors of C are [C, B, A]. The ancestors of A are [A]. The ancestors of D are [D, A]

ancestors(*node_ids*)

Get one or mode nodes' ancestor nodes

Parameters

node_ids
[list of hashable] Items are ids of nodes whose ancestors will be found.

Returns

list of list of dict
Items are lists of ancestor nodes corresponding to argued ids.

child_ids(*node_ids*)

Obtain the ids of one or more nodes' children

Parameters

node_ids
[list of hashable] Items are ids of nodes whose children you wish to find.

Returns

list of list of hashable
Items are lists of input nodes' children's ids.

children(*node_ids*)

Get one or mode nodes' child nodes

Parameters

node_ids
[list of hashable] Items are ids of nodes whose children will be found.

Returns

list of list of dict
Items are lists of child nodes corresponding to argued ids.

descendant_ids(*node_ids*)

Obtain the ids of one or more nodes' descendants

Parameters

node_ids
[list of hashable] Items are ids of nodes whose descendants you wish to find.

Returns

list of list of hashable
Items are lists of input nodes' descendants' ids.

Notes

Given the tree: A -> B -> C

`-> D

The descendants of A are [B, C, D]. The descendants of C are [].

descendants(*node_ids*)

Get one or mode nodes' descendant nodes

Parameters

node_ids
[list of hashable] Items are ids of nodes whose descendants will be found.

Returns

list of list of dict
Items are lists of descendant nodes corresponding to argued ids.

filter_nodes(*criterion*)

Obtain a list of nodes filtered by some criterion

Parameters

criterion
[function | node dict => bool] Only nodes for which criterion returns true will be returned.

Returns

list of dict
Items are node dictionaries that passed the filter.

node(*node_ids=None*)

node_ids()

Obtain the node ids of each node in the tree

Returns

list
elements are node ids

nodes(*node_ids=None*)

Get one or more nodes' full dictionaries from their ids.

Parameters

node_ids
[list of hashable] Items are ids of nodes to be returned. Default is all.

Returns

list of dict
Items are nodes corresponding to argued ids.

nodes_by_property(*key*, *values*, *to_fn=None*)

Get nodes by a specified property

Parameters

key
[hashable or function] The property used for lookup. Should be unique. If a function, will be invoked on each node.

values
[list] Select matching elements from the lookup.

to_fn
[function, optional] Defines the outputs, on a per-node basis. Defaults to returning the whole node.

Returns

list
outputs, 1 for each input value.

`parent`(*node_ids*)

`parent_id`(*node_ids*)

`parent_ids`(*node_ids*)

Obtain the ids of one or more nodes' parents

Parameters

node_ids
[list of hashable] Items are ids of nodes whose parents you wish to find.

Returns

list of hashable
Items are ids of input nodes' parents in order.

`parents`(*node_ids*)

Get one or mode nodes' parent nodes

Parameters

node_ids
[list of hashable] Items are ids of nodes whose parents will be found.

Returns

list of dict
Items are parents of nodes corresponding to argued ids.

`value_map`(*from_fn*, *to_fn*)

Obtain a look-up table relating a pair of node properties across nodes

Parameters

from_fn
[function | node dict => hashable value] The keys of the output dictionary will be obtained by calling from_fn on each node. Should be unique.

to_fn
[function | node_dict => value] The values of the output function will be obtained by calling to_fn on each node.

Returns

dict
Maps the node property defined by from_fn to the node property defined by to_fn across nodes.

allensdk.core.sitk_utilities module

allensdk.core.sitk_utilities.**fix_array_dimensions**(*array*, *ncomponents=1*)

Convenience function that reorders ndarray dimensions for io with SimpleITK

Parameters

array
[np.ndarray] The array to be reordered

ncomponents
[int, optional] Number of components per pixel, default 1.

Returns

np.ndarray
Reordered array

allensdk.core.sitk_utilities.**get_sitk_image_information**(*image*)

Extract information about a SimpleITK image

Parameters

image
[sitk.Image] Extract information about this image.

Returns

dict
Extracted information. Includes spacing, origin, size, direction, and number of components per pixel

allensdk.core.sitk_utilities.**read_ndarray_with_sitk**(*path*)

Read a numpy array from a file using SimpleITK

Parameters

path
[str] Read from this path

Returns

image
[np.ndarray] Obtained array

information
[dict] Additional information about the array

allensdk.core.sitk_utilities.**set_sitk_image_information**(*image*, *information*)

Set information on a SimpleITK image

Parameters

image
[sitk.Image] Set information on this image.

information
[dict] Stores information to be set. Supports spacing, origin, direction. Also checks (but cannot set) size and number of components per pixel

allensdk.core.sitk_utilities.**write_ndarray_with_sitk**(*array*, *path*, ***information*)

Write a numpy array to a file using SimpleITK

Parameters

array
[np.ndarray] Array to be written.

path
[str] Write to here

****information**
[dict] Contains additional information to be stored in the image file. See set_sitk_image_information for more information.

allensdk.core.structure_tree module

class allensdk.core.structure_tree.**StructureTree**(*nodes*)

Bases: *SimpleTree*

static clean_structures(*structures*, *whitelist=None*, *data_transforms=None*, *renames=None*)

Convert structures_with_sets query results into a form that can be used to construct a StructureTree

Parameters

structures
[list of dict] Each element describes a structure. Should have a structure id path field (str values) and a structure_sets field (list of dict).

whitelist
[list of str, optional] Only these fields will be included in the final structure record. Default is the output of StructureTree.whitelist.

data_transforms
[dict, optional] Keys are str field names. Values are functions which will be applied to the data associated with those fields. Default is to map colors from hex to rgb and convert the structure id path to a list of int.

renames
[dict, optional] Controls the field names that appear in the output structure records. Default is to map 'color_hex_triplet' to 'rgb_triplet'.

Returns

list of dict
structures, after conversion of structure_id_path and structure_sets

static collect_sets(*structure*)

Structure sets may be specified by full records or id. This method collects all of the structure set records/ids in a structure record and replaces them with a single list of id records.

static data_transforms()

export_label_description(*alphas=None*, *exclude_label_vis=None*, *exclude_mesh_vis=None*, *label_key='acronym'*)

Produces an itksnap label_description table from this structure tree

Parameters

alphas
[dict, optional] Maps structure ids to alpha levels. Optional - will only use provided ids.

exclude_label_vis
[list, optional] The structures denoted by these ids will not be visible in ITKSnap.

exclude_mesh_vis
[list, optional] The structures denoted by these ids will not have visible meshes in ITKSnap.

label_key: str, optional
Use this column for display labels.

Returns

pd.DataFrame
Contains data needed for loading as an ITKSnap label description file.

`get_ancestor_id_map()`

Get a dictionary mapping structure ids to ancestor ids across all nodes.

Returns

dict
Keys are structure ids. Values are lists of ancestor ids.

`get_colormap()`

Get a dictionary mapping structure ids to colors across all nodes.

Returns

dict
Keys are structure ids. Values are RGB lists of integers.

`get_id_acronym_map()`

Get a dictionary mapping structure acronyms to ids across all nodes.

Returns

dict
Keys are structure acronyms. Values are structure ids.

`get_name_map()`

Get a dictionary mapping structure ids to names across all nodes.

Returns

dict
Keys are structure ids. Values are structure name strings.

`get_structure_sets()`

Lists all unique structure sets that are assigned to at least one structure in the tree.

Returns

list of int
Elements are ids of structure sets.

`get_structures_by_acronym(`*acronyms*`)`

Obtain a list of brain structures from their acronyms

Parameters

names
[list of str] Get structures corresponding to these acronyms.

Returns

list of dict
Each item describes a structure.

`get_structures_by_id`(*structure_ids*)

Obtain a list of brain structures from their structure ids

Parameters

structure_ids
[list of int] Get structures corresponding to these ids.

Returns

list of dict
Each item describes a structure.

`get_structures_by_name`(*names*)

Obtain a list of brain structures from their names,

Parameters

names
[list of str] Get structures corresponding to these names.

Returns

list of dict
Each item describes a structure.

`get_structures_by_set_id`(*structure_set_ids*)

Obtain a list of brain structures from by the sets that contain them.

Parameters

structure_set_ids
[list of int] Get structures belonging to these structure sets.

Returns

list of dict
Each item describes a structure.

`has_overlaps`(*structure_ids*)

Determine if a list of structures contains structures along with their ancestors

Parameters

structure_ids
[list of int] Check this set of structures for overlaps

Returns

set
Ids of structures that are the ancestors of other structures in the supplied set.

`static hex_to_rgb`(*hex_color*)

Convert a hexadecimal color string to a uint8 triplet

Parameters

hex_color
[string] Must be 6 characters long, unless it is 7 long and the first character is #. If hex_color is a triplet of int, it will be returned unchanged.

Returns

list of int
3 characters long - 1 per two characters in the input string.

`static path_to_list(path)`

Structure id paths are sometimes formatted as "/"-seperated strings. This method converts them to a list of integers, if needed.

`static renames()`

`structure_descends_from(child_id, parent_id)`

Tests whether one structure descends from another.

Parameters

child_id
[int] Id of the putative child structure.

parent_id
[int] Id of the putative parent structure.

Returns

bool
True if the structure specified by child_id is a descendant of the one specified by parent_id. Otherwise False.

`static whitelist()`

allensdk.core.swc module

`class allensdk.core.swc.Compartment(*args, **kwargs)`

Bases: `dict`

A dictionary class storing information about a single morphology node

`print_node()`

print out compartment information with field names

`class allensdk.core.swc.Marker(*args, **kwargs)`

Bases: `dict`

Simple dictionary class for handling reconstruction marker objects.

`CUT_DENDRITE = 10`

`NO_RECONSTRUCTION = 20`

`SPACING = [0.1144, 0.1144, 0.28]`

`class allensdk.core.swc.Morphology(compartment_list=None, compartment_index=None)`

Bases: `object`

Keep track of the list of compartments in a morphology and provide a few helper methods (soma, tree information, pruning, etc).

`APICAL_DENDRITE = 4`

`AXON = 2`

BASAL_DENDRITE = 3

DENDRITE = 3

NODE_TYPES = [1, 2, 3, 3, 4]

SOMA = 1

append(*node_list*)

Add additional nodes to this Morphology. Those nodes must originate from another morphology object.

Parameters

node_list: list of Morphology nodes

apply_affine(*aff*, *scale=None*)

Apply an affine transform to all compartments in this morphology. Node radius is adjusted as well.

Format of the affine matrix is:

[x0 y0 z0] [tx] [x1 y1 z1] [ty] [x2 y2 z2] [tz]

where the left 3x3 the matrix defines the affine rotation and scaling, and the right column is the translation vector.

The matrix must be collapsed and stored in a list as follows:

[x0 y0, z0, x1, y1, z1, x2, y2, z2, tx, ty, tz]

Parameters

aff: 3x4 array of floats (python 2D list, or numpy 2D array)
the transformation matrix

change_parent(*child*, *parent*)

Change the parent of a node. The child node is adjusted to point to the new parent, the child is taken off of the previous parent's child list, and it is added to the new parent's child list.

Parameters

child: integer or Morphology Object
The ID of the child node, or the child node itself

parent: integer or Morphology Object
The ID of the parent node, or the parent node itself

Returns

Nothing

children_of(*seg*)

Returns a list of the children of the specified node

Parameters

seg: integer or Morphology Object
The ID of the parent node, or the parent node itself

Returns

A list of the child morphology objects. If the ID of the parent node is invalid, None is returned.

property compartment_index

Return the compartment index. This is a property to ensure that the compartment list and compartment index are in sync.

compartment_index_by_type(*compartment_type*)

Return an dictionary of compartments indexed by id that all have a particular compartment type.

Parameters

compartment_type: int
Desired compartment type

Returns

A dictionary of Morphology Objects, indexed by ID

property compartment_list

Return the compartment list. This is a property to ensure that the compartment list and compartment index are in sync.

compartment_list_by_type(*compartment_type*)

Return an list of all compartments having the specified compartment type.

Parameters

compartment_type: int
Desired compartment type

Returns

A list of of Morphology Objects

convert_type(*old_type*, *new_type*)

Converts all compartments from one type to another. Nodes of the original type are not affected so this procedure can also be used as a merge procedure.

Parameters

old_type: enum
The compartment type to be changed. Use one of the following constants: SOMA, AXON, DENDRITE, BASAL_DENDRITE, or APICAL_DENDRITE

new_type: enum
The target compartment type. Use one of the following constants: SOMA, AXON, DENDRITE, BASAL_DENDRITE, or APICAL_DENDRITE

delete_tree(*n*)

Delete tree, and all of its compartments, from the morphology.

Parameters

n: Integer
The tree number to delete

find(*x*, *y*, *z*, *dist*, *node_type=None*)

Returns a list of Morphology Objects located within 'dist' of coordinate (x,y,z). If node_type is specified, the search will be constrained to return only nodes of that type.

Parameters

x, y, z: float
The x,y,z coordinates from which to search around

dist: float
The search radius

node_type: enum (optional)
One of the following constants: SOMA, AXON, DENDRITE, BASAL_DENDRITE or APICAL_DENDRITE

Returns

A list of all Morphology Objects matching the search criteria

`node`(*n*)

Returns the morphology node having the specified ID.

Parameters

n: integer
ID of desired node

Returns

A morphology object having the specified ID, or None if such a node doesn't exist

`property num_nodes`

Return the number of compartments in the morphology.

`property num_trees`

Return the number of trees in the morphology. A tree is defined as everything following from a single root compartment.

`parent_of`(*seg*)

Returns parent of the specified node.

Parameters

seg: integer or Morphology Object
The ID of the child node, or the child node itself

Returns

A morphology object, or None if no parent exists or if the specified node ID doesn't exist

`property root`

[deprecated] Returns root node of soma, if present. Use 'soma' instead of 'root'

`save`(*file_name*)

Write this morphology out to an SWC file

Parameters

file_name: string
desired name of your SWC file

`property soma`

Returns root node of soma, if present

`sparsify`(*modulo*, *compress_ids=False*)

Return a new Morphology object that has a given number of non-leaf, non-root nodes removed. IDs can be reassigned so as to be continuous.

Parameters

modulo: int
keep 1 out of every modulo nodes.

compress_ids: boolean
Reassign ids so that ids are continuous (no missing id numbers).

Returns

Morphology
A new morphology instance

`strip_all_other_types`(*node_type*, *keep_soma=True*)

Strips everything from the morphology except for the specified type. Parent and child relationships are updated accordingly, creating new roots when necessary.

Parameters

node_type: enum
The compartment type to keep in the morphology. Use one of the following constants: SOMA, AXON, DENDRITE, BASAL_DENDRITE, or APICAL_DENDRITE

keep_soma: Boolean (optional)
True (default) if soma nodes should remain in the morpyhology, and False if the soma should also be stripped

`strip_type`(*node_type*)

Strips all compartments of the specified type from the morphology. Parent and child relationships are updated accordingly, creating new roots when necessary.

Parameters

node_type: enum
The compartment type to strip from the morphology. Use one of the following constants: SOMA, AXON, DENDRITE, BASAL_DENDRITE, or APICAL_DENDRITE

`stumpify_axon`(*count=10*)

Remove all axon compartments except the first 'count' nodes, as counted from the connected axon root.

Parameters

count: Integer
The length of the axon 'stump', in number of compartments

`tree`(*n*)

Returns a list of all Morphology Nodes within the specified tree. A tree is defined as a fully connected graph of nodes. Each tree has exactly one root.

Parameters

n: integer
ID of desired tree

Returns

A list of all morphology objects in the specified tree, or None if the tree doesn't exist

`write`(*file_name*)

`allensdk.core.swc.read_marker_file`(*file_name*)

read in a marker file and return a list of dictionaries

allensdk.core.swc.**read_swc**(*file_name*, *columns='NOT_USED'*, *numeric_columns='NOT_USED'*)

Read in an SWC file and return a Morphology object.

Parameters

file_name: string
SWC file name.

Returns

Morphology
A Morphology instance.

allensdk.core.typing module

class allensdk.core.typing.**SupportsStr**(**args*, ***kwargs*)

Bases: Protocol

Classes that support the __str__ method

allensdk.core.utilities module

allensdk.core.utilities.**df_list_to_tuple**(*df: DataFrame*, *columns: List[str]*) → DataFrame

convert list to tuple so that it can be hashable

allensdk.core.utilities.**literal_col_eval**(*df: DataFrame*, *columns: List[str]*) → DataFrame

Eval string entries of specified columns

Module contents

6.1.5 allensdk.ephys package

Submodules

allensdk.ephys.ephys_extractor module

class allensdk.ephys.ephys_extractor.**EphysCellFeatureExtractor**(*ramps_ext*, *short_squares_ext*, *long_squares_ext*, *subthresh_min_amp=-100*)

Bases: object

SAG_TARGET = -100.0

SUBTHRESH_MAX_AMP = 0

as_dict()

Create dict of cell features.

cell_features()

long_squares_features(*option=None*)

long_squares_stim_amps(*option=None*)

process(*keys=None*)

Processes features. Can take a specific key (or set of keys) to do a subset of processing.

ramps_features(*all=False*)

short_squares_features()

class allensdk.ephys.ephys_extractor.**EphysSweepFeatureExtractor**(*t=None*, *v=None*, *i=None*, *start=None*, *end=None*, *filter=10.0*, *dv_cutoff=20.0*, *max_interval=0.005*, *min_height=2.0*, *min_peak=-30.0*, *thresh_frac=0.05*, *baseline_interval=0.1*, *baseline_detect_thresh=0.3*, *id=None*)

Bases: object

Feature calculation for a sweep (voltage and/or current time series).

as_dict()

Create dict of features and spikes.

burst_metrics()

Find bursts and return max "burstiness" index (normalized max rate in burst vs out).

Returns

max_burstiness_index
[max "burstiness" index across detected bursts]

num_bursts
[number of bursts detected]

delay_metrics()

Calculates ratio of latency to dominant time constant of rise before spike

Returns

delay_ratio
[ratio of latency to tau (higher means more delay)]

tau
[dominant time constant of rise before spike]

estimate_sag(*peak_width=0.005*)

Calculate the sag in a hyperpolarizing voltage response.

Parameters

peak_width
[window width to get more robust peak estimate in sec]

(default 0.005)

Returns

sag
[fraction that membrane potential relaxes back to baseline]

estimate_time_constant()

Calculate the membrane time constant by fitting the voltage response with a single exponential.

Returns

tau
[membrane time constant in seconds]

is_spike_feature_affected_by_clipping(*key*)

pause_metrics()

Estimate average number of pauses and average fraction of time spent in a pause

Attempts to detect pauses with a variety of conditions and averages results together.

Pauses that are consistently detected contribute more to estimates.

Returns

avg_n_pauses
[average number of pauses detected across conditions]

avg_pause_frac
[average fraction of interval (between start and]

end) spent in a pause
max_reliability
[max fraction of times most reliable pause was]

detected given weights tested
n_max_rel_pauses
[number of pauses detected with *max_reliability*]

process_new_spike_feature(*feature_name*, *feature_func*, *affected_by_clipping=False*)

Add new spike-level feature calculation function

The function should take this sweep extractor as its argument. Its results can be accessed by calling the method spike_feature(<feature_name>).

process_new_sweep_feature(*feature_name*, *feature_func*)

Add new sweep-level feature calculation function

The function should take this sweep extractor as its argument. Its results can be accessed by calling the method sweep_feature(<feature_name>).

process_spikes()

Perform spike-related feature analysis

set_stimulus_amplitude_calculator(*function*)

spike_feature(*key*, *include_clipped=False*, *force_exclude_clipped=False*)

Get specified feature for every spike.

Parameters

key
[feature name]

include_clipped: return values for every identified spike, even when clipping means they will be incorrect/undefined

Returns

spike_feature_values
[ndarray of features for each spike]

`spike_feature_keys()`

Get list of every available spike feature.

`spikes()`

Get all features for each spike as a list of records.

`stimulus_amplitude()`

`sweep_feature(key, allow_missing=False)`

Get sweep-level feature (*key*).

Parameters

key
[name of sweep-level feature]

allow_missing
[return np.nan if key is missing for sweep (default]

False)

Returns

sweep_feature
[sweep-level feature value]

`sweep_feature_keys()`

Get list of every available sweep-level feature.

`voltage_deflection(deflect_type=None)`

Measure deflection (min or max, between start and end if specified).

Parameters

deflect_type
[measure minimal ('min') or maximal ('max') voltage]

deflection
If not specified, it will check to see if the current (i) is positive or negative between start and end, then choose 'max' or 'min', respectively If the current is not defined, it will default to 'min'.

Returns

deflect_v
[peak]

deflect_index
[index of peak deflection]

class allensdk.ephys.ephys_extractor.**EphysSweepSetFeatureExtractor**(*t_set=None*, *v_set=None*, *i_set=None*, *start=None*, *end=None*, *filter=10.0*, *dv_cutoff=20.0*, *max_interval=0.005*, *min_height=2.0*, *min_peak=-30.0*, *thresh_frac=0.05*, *baseline_interval=0.1*, *baseline_detect_thresh=0.3*, *id_set=None*)

Bases: object

classmethod from_sweeps(*sweep_list*)

Initialize EphysSweepSetFeatureExtractor object with a list of pre-existing sweep feature extractor objects.

process_spikes()

Analyze spike features for all sweeps.

spike_feature_averages(*key*)

Get nparray of average spike-level feature (*key*) for all sweeps

sweep_features(*key*, *allow_missing=False*)

Get nparray of sweep-level feature (*key*) for all sweeps

Parameters

key
[name of sweep-level feature]

allow_missing
[return np.nan if key is missing for sweep (default]

False)

Returns

sweep_feature
[nparray of sweep-level feature values]

sweeps()

Get list of EphysSweepFeatureExtractor objects.

allensdk.ephys.ephys_extractor.**cell_extractor_for_nwb**(*dataset*, *ramps*, *short_squares*, *long_squares*, *subthresh_min_amp=-100*)

Initialize EphysCellFeatureExtractor object from NWB data set

Parameters

dataset
[NwbDataSet]

ramps
[list of sweep numbers of ramp sweeps]

short_squares
[list of sweep numbers of short square sweeps]

long_squares
[list of sweep numbers of long square sweeps]

allensdk.ephys.ephys_extractor.**extractor_for_nwb_sweeps**(*dataset*, *sweep_numbers*, *fixed_start=None*, *fixed_end=None*, *dv_cutoff=20.0*, *thresh_frac=0.05*)

allensdk.ephys.ephys_extractor.**fit_fi_slope**(*ext*)

Fit the rate and stimulus amplitude to a line and return the slope of the fit.

allensdk.ephys.ephys_extractor.**input_resistance**(*ext*)

Estimate input resistance in MOhms, assuming all sweeps in passed extractor are hyperpolarizing responses.

allensdk.ephys.ephys_extractor.**membrane_time_constant**(*ext*)

Average the membrane time constant values estimated from each sweep in passed extractor.

allensdk.ephys.ephys_extractor.**reset_long_squares_start**(*when*)

allensdk.ephys.ephys_features module

exception allensdk.ephys.ephys_features.**FeatureError**

Bases: Exception

Generic Python-exception-derived object raised by feature detection functions.

allensdk.ephys.ephys_features.**adaptation_index**(*isis*)

Calculate adaptation index of *isis*.

allensdk.ephys.ephys_features.**analyze_trough_details**(*v*, *t*, *spike_indexes*, *peak_indexes*, *clipped=None*, *end=None*, *filter=10.0*, *heavy_filter=1.0*, *term_frac=0.01*, *adp_thresh=0.5*, *tol=0.5*, *flat_interval=0.002*, *adp_max_delta_t=0.005*, *adp_max_delta_v=10.0*, *dvdt=None*)

Analyze trough to determine if an ADP exists and whether the reset is a 'detour' or 'direct'

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

spike_indexes
[numpy array of spike indexes]

peak_indexes
[numpy array of spike peak indexes]

end
[end of time window (optional)]

filter
[cutoff frequency for 4-pole low-pass Bessel filter in kHz (default 1)]

heavy_filter
[lower cutoff frequency for 4-pole low-pass Bessel filter in kHz (default 1)]

thresh_frac
[fraction of average upstroke for threshold calculation (optional, default 0.05)]

adp_thresh: minimum dV/dt in V/s to exceed to be considered to have an ADP (optional, default 1.5)
tol
[tolerance for evaluating whether Vm drops appreciably further after end of spike (default 1.0 mV)]

flat_interval: if the trace is flat for this duration, stop looking for an ADP (default 0.002 s)
adp_max_delta_t: max possible ADP delta t (default 0.005 s)
adp_max_delta_v: max possible ADP delta v (default 10 mV)
dvdt
[pre-calculated time-derivative of voltage (optional)]

Returns

isi_types
[numpy array of isi reset types (direct or detour)]

fast_trough_indexes
[numpy array of indexes at the start of the trough (i.e. end of the spike)]

adp_indexes
[numpy array of adp indexes (np.nan if there was no ADP in that ISI]

slow_trough_indexes
[numpy array of indexes at the minimum of the slow phase of the trough] (if there wasn't just a fast phase)

allensdk.ephys.ephys_features.**average_rate**(*t*, *spikes*, *start*, *end*)

Calculate average firing rate during interval between *start* and *end*.

Parameters

t
[numpy array of times in seconds]

spikes
[numpy array of spike indexes]

start
[start of time window for spike detection]

end
[end of time window for spike detection]

Returns

avg_rate
[average firing rate in spikes/sec]

allensdk.ephys.ephys_features.**average_voltage**(*v*, *t*, *start=None*, *end=None*)

Calculate average voltage between start and end.

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

start
[start of time window for spike detection (optional, default None)]

end
[end of time window for spike detection (optional, default None)]

Returns

v_avg
[average voltage]

allensdk.ephys.ephys_features.**calculate_dvdt**(*v*, *t*, *filter=None*)

Low-pass filters (if requested) and differentiates voltage by time.

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

filter
[cutoff frequency for 4-pole low-pass Bessel filter in kHz (optional, default None)]

Returns

dvdt
[numpy array of time-derivative of voltage (V/s = mV/ms)]

allensdk.ephys.ephys_features.**check_thresholds_and_peaks**(*v*, *t*, *spike_indexes*, *peak_indexes*, *upstroke_indexes*, *end=None*, *max_interval=0.005*, *thresh_frac=0.05*, *filter=10.0*, *dvdt=None*, *tol=1.0*)

Validate thresholds and peaks for set of spikes

Check that peaks and thresholds for consecutive spikes do not overlap Spikes with overlapping thresholds and peaks will be merged.

Check that peaks and thresholds for a given spike are not too far apart.

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

spike_indexes
[numpy array of spike indexes]

peak_indexes
[numpy array of indexes of spike peaks]

upstroke_indexes
[numpy array of indexes of spike upstrokes]

max_interval
[maximum allowed time between start of spike and time of peak in sec (default 0.005)]

thresh_frac
[fraction of average upstroke for threshold calculation (optional, default 0.05)]

filter
[cutoff frequency for 4-pole low-pass Bessel filter in kHz (optional, default 10)]

dvdt
[pre-calculated time-derivative of voltage (optional)]

tol
[tolerance for returning to threshold in mV (optional, default 1)]

Returns

spike_indexes
[numpy array of modified spike indexes]

peak_indexes
[numpy array of modified spike peak indexes]

upstroke_indexes
[numpy array of modified spike upstroke indexes]

clipped
[numpy array of clipped status of spikes]

allensdk.ephys.ephys_features.**detect_bursts**(*isis*, *isi_types*, *fast_tr_v*, *fast_tr_t*, *slow_tr_v*, *slow_tr_t*, *thr_v*, *tol=0.5*, *pause_cost=1.0*)

Detect bursts in spike train.

Parameters

isis
[numpy array of n interspike intervals]

isi_types
[numpy array of n interspike interval types]

fast_tr_v
[numpy array of fast trough voltages for the n + 1 spikes of the train]

fast_tr_t
[numpy array of fast trough times for the n + 1 spikes of the train]

slow_tr_v
[numpy array of slow trough voltages for the n + 1 spikes of the train]

slow_tr_t
[numpy array of slow trough times for the n + 1 spikes of the train]

thr_v
[numpy array of threshold voltages for the n + 1 spikes of the train]

tol
[tolerance for the difference in slow trough voltages and thresholds (default 0.5 mV)] Used to identify "delay" interspike intervals that occur within a burst

Returns

bursts
[list of bursts] Each item in list is a tuple of the form (burst_index, start, end) where *burst_index* is a comparison index between the highest instantaneous rate within the burst vs the highest instantaneous rate outside the burst. *start* is the index of the first ISI of the burst, and *end* is the ISI index immediately following the burst.

allensdk.ephys.ephys_features.**detect_pauses**(*isis*, *isi_types*, *cost_weight=1.0*)

Determine which ISIs are "pauses" in ongoing firing.

Pauses are unusually long ISIs with a "detour reset" among "direct resets".

Parameters

isis
[numpy array of interspike intervals]

isi_types
[numpy array of interspike interval types ('direct' or 'detour')]

cost_weight
[weight for cost function for calling an ISI a pause] Higher cost weights lead to fewer ISIs identified as pauses. The cost function also depends on the difference between the duration of the "pause" ISIs and the average duration and standard deviation of "non-pause" ISIs.

Returns

pauses
[numpy array of indices corresponding to pauses in *isis*]

allensdk.ephys.ephys_features.**detect_putative_spikes**(*v*, *t*, *start=None*, *end=None*, *filter=10.0*, *dv_cutoff=20.0*)

Perform initial detection of spikes and return their indexes.

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

start
[start of time window for spike detection (optional)]

end
[end of time window for spike detection (optional)]

filter
[cutoff frequency for 4-pole low-pass Bessel filter in kHz (optional, default 10)]

dv_cutoff
[minimum dV/dt to qualify as a spike in V/s (optional, default 20)]

dvdt
[pre-calculated time-derivative of voltage (optional)]

Returns

putative_spikes
[numpy array of preliminary spike indexes]

allensdk.ephys.ephys_features.**estimate_adjusted_detection_parameters**(*v_set*, *t_set*, *interval_start*, *interval_end*, *filter=10*)

Estimate adjusted values for spike detection by analyzing a period when the voltage changes quickly but passively (due to strong current stimulation), which can result in spurious spike detection results.

Parameters

v_set
[list of numpy arrays of voltage time series in mV]

t_set
[list of numpy arrays of times in seconds]

interval_start
[start of analysis interval (sec)]

interval_end
[end of analysis interval (sec)]

Returns

new_dv_cutoff
[adjusted dv/dt cutoff (V/s)]

new_thresh_frac
[adjusted fraction of avg upstroke to find threshold]

allensdk.ephys.ephys_features.**filter_putative_spikes**(*v*, *t*, *spike_indexes*, *peak_indexes*, *min_height=2.0*, *min_peak=-30.0*, *filter=10.0*, *dvdt=None*)

Filter out events that are unlikely to be spikes based on:

- Voltage failing to go down between peak and the next spike's threshold
- Height (threshold to peak)
- Absolute peak level

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

spike_indexes
[numpy array of preliminary spike indexes]

peak_indexes
[numpy array of indexes of spike peaks]

min_height
[minimum acceptable height from threshold to peak in mV (optional, default 2)]

min_peak
[minimum acceptable absolute peak level in mV (optional, default -30)]

filter
[cutoff frequency for 4-pole low-pass Bessel filter in kHz (optional, default 10)]

dvdt
[pre-calculated time-derivative of voltage (optional)]

Returns

spike_indexes
[numpy array of threshold indexes]

peak_indexes
[numpy array of peak indexes]

allensdk.ephys.ephys_features.**find_downstroke_indexes**(*v*, *t*, *peak_indexes*, *trough_indexes*, *clipped=None*, *filter=10.0*, *dvdt=None*)

Find indexes of minimum voltage (troughs) between spikes.

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

peak_indexes
[numpy array of spike peak indexes]

trough_indexes
[numpy array of threshold indexes]

clipped: boolean array - False if spike not clipped by edge of window
filter
[cutoff frequency for 4-pole low-pass Bessel filter in kHz (optional, default 10)]

dvdt
[pre-calculated time-derivative of voltage (optional)]

Returns

downstroke_indexes
[numpy array of downstroke indexes]

allensdk.ephys.ephys_features.**find_peak_indexes**(*v*, *t*, *spike_indexes*, *end=None*)

Find indexes of spike peaks.

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

spike_indexes
[numpy array of preliminary spike indexes]

end
[end of time window for spike detection (optional)]

allensdk.ephys.ephys_features.**find_time_index**(*t*, *t_0*)

Find the index value of a given time (t_0) in a time series (t).

allensdk.ephys.ephys_features.**find_trough_indexes**(*v*, *t*, *spike_indexes*, *peak_indexes*, *clipped=None*, *end=None*)

Find indexes of minimum voltage (trough) between spikes.

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

spike_indexes
[numpy array of spike indexes]

peak_indexes
[numpy array of spike peak indexes]

end
[end of time window (optional)]

Returns

trough_indexes
[numpy array of threshold indexes]

allensdk.ephys.ephys_features.**find_upstroke_indexes**(*v*, *t*, *spike_indexes*, *peak_indexes*, *filter=10.0*, *dvdt=None*)

Find indexes of maximum upstroke of spike.

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

spike_indexes
[numpy array of preliminary spike indexes]

peak_indexes
[numpy array of indexes of spike peaks]

filter
[cutoff frequency for 4-pole low-pass Bessel filter in kHz (optional, default 10)]

dvdt
[pre-calculated time-derivative of voltage (optional)]

Returns

upstroke_indexes
[numpy array of upstroke indexes]

allensdk.ephys.ephys_features.**find_widths**(*v*, *t*, *spike_indexes*, *peak_indexes*, *trough_indexes*, *clipped=None*)

Find widths at half-height for spikes.

Widths are only returned when heights are defined

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

spike_indexes
[numpy array of spike indexes]

peak_indexes
[numpy array of spike peak indexes]

trough_indexes
[numpy array of trough indexes]

Returns

widths
[numpy array of spike widths in sec]

allensdk.ephys.ephys_features.**fit_membrane_time_constant**(*v*, *t*, *start*, *end*, *min_rsme=0.0001*)

Fit an exponential to estimate membrane time constant between start and end

Parameters

v
[numpy array of voltages in mV]

t
[numpy array of times in seconds]

start
[start of time window for exponential fit]

end
[end of time window for exponential fit]

min_rsme: minimal acceptable root mean square error (default 1e-4)

Returns

a, inv_tau, y0
[Coeffients of equation y0 + a * exp(-inv_tau * x)]

returns np.nan for values if fit fails

allensdk.ephys.ephys_features.**fit_prespike_time_constant**(*v*, *t*, *start*, *spike_time*, *dv_limit=-0.001*, *tau_limit=0.3*)

Finds the dominant time constant of the pre-spike rise in voltage

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

start
[start of voltage rise (seconds)]

spike_time
[time of first spike (seconds)]

dv_limit
[dV/dt cutoff (default -0.001)] Shortens fit window if rate of voltage drop exceeds this limit

tau_limit
[upper bound for slow time constant (seconds, default 0.3)] If the slower time constant of a double-exponential fit is twice that of the faster and exceeds this limit, the faster one will be considered the dominant one

Returns

tau
[dominant time constant (seconds)]

allensdk.ephys.ephys_features.**get_isis**(*t*, *spikes*)

Find interspike intervals in sec between spikes (as indexes).

allensdk.ephys.ephys_features.**has_fixed_dt**(*t*)

Check that all time intervals are identical.

allensdk.ephys.ephys_features.**latency**(*t*, *spikes*, *start*)

Calculate time to the first spike.

allensdk.ephys.ephys_features.**norm_diff**(*a*)

Calculate average of (a[i] - a[i+1]) / (a[i] + a[i+1]).

allensdk.ephys.ephys_features.**norm_sq_diff**(*a*)

Calculate average of (a[i] - a[i+1])^2 / (a[i] + a[i+1])^2.

allensdk.ephys.ephys_features.**refine_threshold_indexes**(*v*, *t*, *upstroke_indexes*, *thresh_frac=0.05*, *filter=10.0*, *dvdt=None*)

Refine threshold detection of previously-found spikes.

Parameters

v
[numpy array of voltage time series in mV]

t
[numpy array of times in seconds]

upstroke_indexes
[numpy array of indexes of spike upstrokes (for threshold target calculation)]

thresh_frac
[fraction of average upstroke for threshold calculation (optional, default 0.05)]

filter
[cutoff frequency for 4-pole low-pass Bessel filter in kHz (optional, default 10)]

dvdt
[pre-calculated time-derivative of voltage (optional)]

Returns

threshold_indexes
[numpy array of threshold indexes]

allensdk.ephys.extract_cell_features module

allensdk.ephys.extract_cell_features.**extract_cell_features**(*data_set*, *ramp_sweep_numbers*, *short_square_sweep_numbers*, *long_square_sweep_numbers*, *subthresh_min_amp=None*)

allensdk.ephys.extract_cell_features.**extract_sweep_features**(*data_set*, *sweeps_by_type*)

allensdk.ephys.extract_cell_features.**get_ramp_stim_characteristics**(*i*, *t*)

Identify the start time and start index of a ramp sweep.

allensdk.ephys.extract_cell_features.**get_square_stim_characteristics**(*i*, *t*, *no_test_pulse=False*)

Identify the start time, duration, amplitude, start index, and end index of a square stimulus. This assumes that there is a test pulse followed by the stimulus square.

allensdk.ephys.extract_cell_features.**get_stim_characteristics**(*i*, *t*, *no_test_pulse=False*)

Identify the start time, duration, amplitude, start index, and end index of a general stimulus. This assumes that there is a test pulse followed by the stimulus square.

allensdk.ephys.extract_cell_features.**mean_features_spike_zero**(*sweeps*)

Compute mean feature values for the first spike in list of extractors

allensdk.ephys.feature_extractor module

class allensdk.ephys.feature_extractor.**EphysFeatureExtractor**

Bases: object

adaptation_index(*spikes*, *stim_end*)

calculate_trough(*spike*, *v*, *curr*, *t*, *next_idx*)

isicv(*spikes*)

process_instance(*name*, *v*, *curr*, *t*, *onset*, *dur*, *stim_name*)

push_summary(*new_summary*)

score_feature_set(*set_num*)

summarize(*summary*)

class allensdk.ephys.feature_extractor.**EphysFeatures**(*name*)

Bases: object

clone(*param_dict*)

print_out()

Module contents

6.1.6 allensdk.internal package

Subpackages

allensdk.internal.api package

Subpackages

allensdk.internal.api.queries package

Submodules

allensdk.internal.api.queries.behavior_lims_queries module

allensdk.internal.api.queries.behavior_lims_queries.**foraging_id_map_from_behavior_session_id**(*lims_engine: PostgresQueryMixin*, *behavior_session_id_list: List[int]*, *logger: RootLogger | None = None*) → DataFrame

Returns DataFrame with two columns:
foraging_id behavior_session_id

Parameters

lims_engine: PostgresQueryMixin
Means of connecting to the LIMS database

behavior_session_ids: List[int]
List of behavior_session_ids for which we want the foraging_id

allensdk.internal.api.queries.behavior_lims_queries.**stimulus_pickle_paths_from_behavior_session_ids**(*lims* Post gres Que *be- hav- ior_ List[* → Data

Get a DataFrame mapping behavior_session_id to stimulus_pickle_path

Parameters

lims_connection: PostgresQueryMixin
behavior_session_id_list: List[int]

Returns

beh_to_path: pd.DataFrame

with columns
behavior_session_id pkl_path

allensdk.internal.api.queries.biophysical_module_api module

class allensdk.internal.api.queries.biophysical_module_api.**BiophysicalModuleApi**(*base_uri=None*)

Bases: *RmaTemplate*

get_neuronal_model_runs(*neuronal_model_run_ids=None*)

List Neuronal Model Rusn available through LIMS with associated info needed to run in NEURON.

Parameters

neuronal_model_run_ids
[integer or list of integers, optional] only select specific neuronal_model_runs.

Returns

dict
[neuronal model run metadata]

get_neuronal_models(*neuronal_model_ids=None*)

List Neuronal Models available through LIMS with associated info needed to run in NEURON.

Parameters

neuronal_model_ids
[integer or list of integers, optional] only select specific neuronal_models.

Returns

dict
[neuronal model metadata]

```
rma_templates = {'biophysical_lims_queries': [{'name':
'neuronal_model_runs_by_ids', 'description': 'see name', 'model':
'NeuronalModelRun', 'criteria': '[id$in{{ neuronal_model_run_ids }}]', 'include':
'well_known_files(well_known_file_type),
neuronal_model(well_known_files(well_known_file_type),specimen(project,
specimen_tags,ephys_roi_result(ephys_qc_criteria,
well_known_files(well_known_file_type)),
neuron_reconstructions(well_known_files(well_known_file_type)),
ephys_sweeps(ephys_sweep_tags,ephys_stimulus(ephys_stimulus_type))),
neuronal_model_template(neuronal_model_template_type,
well_known_files(well_known_file_type)))', 'num_rows': 'all', 'count': False,
'criteria_params': ['neuronal_model_run_ids']}, {'name': 'neuronal_models_by_ids',
'description': 'see name', 'model': 'NeuronalModel', 'criteria': '[id$in{{
neuronal_model_ids }}]', 'include':
'well_known_files(well_known_file_type),specimen(project,specimen_tags,
ephys_roi_result(ephys_qc_criteria,well_known_files(well_known_file_type)),
neuron_reconstructions(well_known_files(well_known_file_type)),
ephys_sweeps(ephys_sweep_tags,ephys_stimulus(ephys_stimulus_type))),
neuronal_model_template(neuronal_model_template_type,
well_known_files(well_known_file_type))', 'num_rows': 'all', 'count': False,
'criteria_params': ['neuronal_model_ids']}]}
```

allensdk.internal.api.queries.biophysical_module_reader module

class allensdk.internal.api.queries.biophysical_module_reader.**BiophysicalModuleReader**

Bases: object

MOD_FILE_TYPE_ID = 292178729

MORPHOLOGY_TYPE_ID = 303941301

STIMULUS_CONTENT_TYPE = None

fit_parameters_file_entries()

read the fit_parameter file path from the lims result corresponding to the stimulus file :return: well_known_file entries :rtype: array of dicts

fit_parameters_path()

Get the path to the fit parameters file from the lims result. :return: path to file :rtype: string

lims_working_directory()

While this is the same directory as the neuronal_model_run directory, it can be mocked out for testing if the other directory is read only.

mod_file_entries()

read the NERUON .mod file entries from the lims result corresponding to the NeuronModel :return: well known file entries :rtype: array of dicts

mod_file_paths()

Get the paths to the mod files from the lims result. :return: paths to mod files :rtype: array of strings

model_type()

TODO: comment

morphology_file_entries()

read the well known file paths from the lims result corresponding to the morphology

Returns

arrary of dicts:
well known file entries

morphology_path()

Get the path to the morphology file from the lims result. :return: path to morphology file :rtype: string

neuronal_model_run_dir()

read the directory path where output goes from the lims optimization config json

Parameters

Returns

string:
directory path

read_json(*path*)

read_json_string(*json_string*)

read_lims_file(*lims_path*)

read_lims_message(*message*, *lims_path*)

set_workflow_state(*state*)

stimulus_file_entries()

read the well known file path from the lims result corresponding to the stimulus file :return: well_known_file entries :rtype: array of dicts

stimulus_path()

Get the path to the stimulus file from the lims result. :return: path to stimulus file :rtype: string

sweep_entries()

read the sweep entries from the lims result corresponding to the stimulus :return: stimulus sweep entries :rtype: array of dicts

sweep_numbers()

Get the stimulus sweep numbers from the lims result :return: list of sweep numbers :rtype: array of ints

sweep_numbers_by_type()

to_manifest(*manifest_path=None*)

update_well_known_file(*path*, *well_known_file_type_id=None*)

write_file(*path*)

allensdk.internal.api.queries.compound_lims_queries module

allensdk.internal.api.queries.compound_lims_queries.**behavior_sessions_from_ecephys_session_ids**(*lims_conne*
Post-
gres-
QueryMixi
ece-
phys_sessi
List[int])
→
DataFrame

Get a DataFrame listing all of the behavior sessions that mice from a specified list of ecephys sessions went through

Parameters

lims_connection: PostgresQueryMixin
ecephys_session_id_list: List[int]
The ecephys sessions used to find the mice used to find the behavior sessions

Returns

mouse_to_behavior: pd.DataFrame

Dataframe with columns
mouse_id behavior_session_id session_type date_of_acquisition date_of_birth ecephys_session_id genotype sex equipment_name

listing every behavior session the mice in question went through

allensdk.internal.api.queries.ecephys_lims_queries module

allensdk.internal.api.queries.ecephys_lims_queries.**donor_id_list_from_ecephys_session_ids**(*lims_connection: PostgresQueryMixin*, *session_id_list: List[int]*) → List[int]

Get the list of donor IDs associated with a list of ecephys_session_ids

allensdk.internal.api.queries.ecephys_lims_queries.**donor_id_lookup_from_ecephys_session_ids**(*lims_connectio PostgresQueryMixin*, *session_id_list: List[int]*) → DataFrame

Return a dataframe with columns ecephys_session_id donor_id from a specified list of ecephys_session_ids

allensdk.internal.api.queries.equipment_lims_queries module

allensdk.internal.api.queries.equipment_lims_queries.**experiment_configs_from_equipment_id**(*equipment_id: int*, *config_type_id: int*, *lims_connection: PostgresQueryMixin*) → DataFrame

Return the configuration of a piece of experimental equipment as a function of time.

Parameters

equipment_id: int
config_type_id: str
The observatory_experiment_config_types.id corresponding to the configuration you want.

lims_connection: PostrgresQueryMixin

Returns

experiment_config: pd.DataFrame

Columns are
active_date – the date the config took effect center_x_mm center_y_mm center_z_mm rotation_x_deg rotation_y_deg rotation_z_deg

allensdk.internal.api.queries.equipment_lims_queries.**experiment_configs_from_equipment_id_and_type**(*equip*
int,
con-
fig_ty
str,
*lims_*
Post-
gres-
Query
→
DataF

Return the configuration of a piece of experimental equipment as a function of time.

Parameters

equipment_id: int
config_type: str
The observatory_experiment_config_types.name corresponding to the configuration you want. One of 'led position', 'behavior camera position', 'eye camera position', or 'screen position'

lims_connection: PostrgresQueryMixin

Returns

experiment_config: pd.DataFrame

Columns are
active_date – the date the config took effect center_x_mm center_y_mm center_z_mm rotation_x_deg rotation_y_deg rotation_z_deg

allensdk.internal.api.queries.grid_data_api_prerelease module

class allensdk.internal.api.queries.grid_data_api_prerelease.**GridDataApiPrerelease**(*storage_directories*,
resolution=None,
base_uri=None)

Bases: *GridDataApi*

Client for retrieving prereleased mouse connectivity data from lims.

Parameters

base_uri
[string, optional] Does not affect pulling from lims.

file_name
[string, optional] File name to save/read storage_directories dict. Passed to GridDataApiPrerelease constructor.

GRID_DATA_DIRECTORY = 'grid'

download_projection_grid_data(*path*, *experiment_id*, *file_name*)

Copy data from path to file_name.

Parameters

path
[string] path to file in shared directory (copy source)

experiment_id
[int] image series id.

file_name
[string] path to file destination (copy target)

classmethod from_file_name(*file_name*, *cache=True*, ***kwargs*)

Alternative constructor using cache path file_name.

Parameters

file_name
[string] Path where storage_directories will be saved.

****kwargs**
Keyword arguments to be supplied to __init__

Returns

cls
[instance of GridDataApiPrerelease]

allensdk.internal.api.queries.mouse_connectivity_api_prerelease module

class allensdk.internal.api.queries.mouse_connectivity_api_prerelease.**MouseConnectivityApiPrerelease**(*sto*
cac
bas

Bases: *MouseConnectivityApi*

Client for retrieving prereleased mouse connectivity data from lims.

Parameters

base_uri
[string, optional] Does not affect pulling from lims.

file_name
[string, optional] File name to save/read storage_directories dict. Passed to GridDataApiPrerelease constructor.

download_data_mask(*path*, *experiment_id*, *resolution*)

download_injection_density(*path*, *experiment_id*, *resolution*)

download_injection_fraction(*path*, *experiment_id*, *resolution*)

download_projection_density(*path*, *experiment_id*, *resolution*)

get_experiments()

Fetch experiment metadata from the Mouse Brain Connectivity Atlas.

Parameters

structure_ids
[integer or list, optional] injection structure

Returns

url
[string] The constructed URL

get_structure_unionizes()

allensdk.internal.api.queries.optimize_config_reader module

class allensdk.internal.api.queries.optimize_config_reader.**OptimizeConfigReader**

Bases: object

MOD_FILE_TYPE_ID = 292178729

MORPHOLOGY_TYPE_ID = 303941301

NEURONAL_MODEL_PARAMETERS = 329230374

STIMULUS_CONTENT_TYPE = None

build_manifest(*manifest_path=None*)

lims_working_directory()

While this is the same directory as the optimize directory, it can be mocked out for testing if the optimize directory is write only.

mod_file_entries()

read the NERUON .mod file entries from the lims result corresponding to the NeuronModel :return: well known file entries :rtype: array of dicts

mod_file_paths()

Get the paths to the mod files from the lims result. :return: paths to mod files :rtype: array of strings

morphology_file_entries()

read the well known file paths from the lims result corresponding to the morphology

Returns

arrary of dicts:
well known file entries

morphology_path()

Get the path to the morphology file from the lims result. :return: path to morphology file :rtype: string

neuronal_model_optimize_dir()

read the directory path where output goes from the lims optimization config json

Parameters

Returns

string:
directory path

output_directory()

read_json(*path*)

read_json_string(*json_string*)

read_lims_file(*lims_path*)

read_lims_message(*message*, *lims_path*)

stimulus_file_entries()

read the well known file path from the lims result corresponding to the stimulus file :return: well_known_file entries :rtype: array of dicts

stimulus_path()

Get the path to the stimulus file from the lims result. :return: path to stimulus file :rtype: string

sweep_entries()

read the sweep entries from the lims result corresponding to the stimulus :return: stimulus sweep entries :rtype: array of dicts

sweep_numbers()

Get the stimulus sweep numbers from the lims result :return: list of sweep numbers :rtype: array of ints

to_manifest(*manifest_path=None*)

update_well_known_file(*path*, *well_known_file_type_id=None*)

write_file(*path*)

allensdk.internal.api.queries.pre_release module

allensdk.internal.api.queries.utils module

allensdk.internal.api.queries.utils.**build_in_list_selector_query**(*col: str*, *valid_list: List[SupportsStr] | None = None*, *operator: str = 'WHERE'*, *valid: bool = True*) → str

Filter for rows where the value of a column is contained in a list (or, if valid=False, where the value is not contained in that list). If no list is specified in *valid_list*, return an empty string.

Parameters

col: str
The name of the column being filtered on

valid_list: Optional[SupportsStr]
The list of values to test column on

operator: str
The SQL operator that starts the clause ("WHERE", "AND" or "OR")

valid: bool
If True, test for "col IN valid_list"; else, test for "col NOT IN valid_list"

Returns

session_query: str
The clause performing the request filter

allensdk.internal.api.queries.utils.**build_where_clause**(*clauses: List[str]*)

allensdk.internal.api.queries.wkf_lims_queries module

allensdk.internal.api.queries.wkf_lims_queries.**wkf_path_from_attachable**(*lims_connection: PostgresQueryMixin*, *wkf_type_name: List[str]*, *attachable_type: str*, *attachable_id: int*) → Dict[str, str]

Get the path to well known files, selecting files of a specific type with a specified attachable ID and attachable_type.

Parameters

lims_connection: PostgresQueryMixin
wkf_type_name: List[str]
i.e. 'StimulusPickle' or 'RawEyeTrackingVideoMetadata' etc.

attachable_type: str
the value of well_known_file.attachable_type to look for

attachable_id: int
the value of well_known_file.attachable_id to look for

Returns

wkf_path_lookup: Dict[str, str]
a dict mapping attachable_type to absolute file path

Notes

Will raise an error if more than one result is returned for a single type

Module contents

Submodules

allensdk.internal.api.api_prerelease module

class allensdk.internal.api.api_prerelease.**ApiPrerelease**(*api_base_url_string=None*)

Bases: *Api*

Extends allensdk.api.api to copy files 'locally' from shared storage.

retrieve_file_from_storage(*storage_path*, *save_file_path*)

Copy data from path to file_name.

Parameters

storage_path
[string] path to file in shared directory (copy source)

save_file_name
[string] path to file destination (copy target)

allensdk.internal.api.lims_api module

class allensdk.internal.api.lims_api.**LimsApi**(*lims_credentials:* DbCredentials | *None* = *None*)

Bases: object

get_behavior_tracking_video_filepath_df()

get_experiment_id()

get_eye_tracking_video_filepath_df()

allensdk.internal.api.mtrain_api module

Module contents

exception allensdk.internal.api.**OneOrMoreResultExpectedError**

Bases: RuntimeError

class allensdk.internal.api.**PostgresQueryMixin**(*, *dbname*, *user*, *host*, *password*, *port*)

Bases: object

fetchall(*query*, *strict=True*)

fetchone(*query*, *strict=True*)

get_connection()

get_cursor()

select(*query*)

select_one(*query*)

allensdk.internal.api.**db_connection_creator**(*credentials:* DbCredentials | *None* = *None*, *fallback_credentials: dict* | *None* = *None*) → *PostgresQueryMixin*

Create a db connection using credentials. If credentials are not provided then use fallback credentials (which attempt to read from shell environment variables).

Note: Must provide one of either 'credentials' or 'fallback_credentials'. If both are provided, 'credentials' will take precedence.

Parameters

credentials
[Optional[DbCredentials], optional] User specified credentials, by default None

fallback_credentials
[dict] Fallback credentials to use for creating the DB connection in the case that no 'credentials' are provided, by default None.

Fallback credentials will attempt to get db connection info from shell environment variables.

Some examples of environment variables that fallback credentials will try to read from can be found in allensdk.core.auth_config.

Returns

PostgresQueryMixin
A DB connection instance which can execute queries to the DB specified by credentials or fallback_credentials.

Raises

RuntimeError
If neither 'credentials' nor 'fallback_credentials' were provided.

allensdk.internal.api.**psycopg2_select**(*query*, *database*, *host*, *port*, *username*, *password*)

allensdk.internal.brain_observatory package

Subpackages

allensdk.internal.brain_observatory.resources package

Module contents

allensdk.internal.brain_observatory.util package

Submodules

allensdk.internal.brain_observatory.util.multi_session_utils module

Module contents

Submodules

allensdk.internal.brain_observatory.annotated_region_metrics module

Module for calculating annotated region metrics from ISI data

allensdk.internal.brain_observatory.annotated_region_metrics.**create_region_mask**(*image_shape*, *x*, *y*, *width*, *height*, *mask*)

Create mask for region on retinotopic map

Parameters

image_shape
[tuple] (height, width) of retinotopic map

x
[int] x offset of region mask within retinotopic map

y
[int] y offset of region mask within retinotopic map

width
[int] width of region mask

height
[int] height of region mask

mask
[list] region mask as a list of lists

Returns

numpy.ndarray
Region mask

allensdk.internal.brain_observatory.annotated_region_metrics.**eccentricity**(*az*, *alt*, *az_center*, *alt_center*)

Compute eccentricity

Parameters

az
[numpy.ndarray] Azimuth retinotopic map

alt
[numpy.ndarray] Altitude retinotopic map

az_center
[float] Azimuth value to use as center of eccentricity map

alt_center
[float] Altitude value to use as center of eccentricity map

Returns

numpy.ndarray
Eccentricity map

allensdk.internal.brain_observatory.annotated_region_metrics.**get_metrics**(*altitude_phase*, *azimuth_phase*, *x=None*, *y=None*, *width=None*, *height=None*, *mask=None*, *altitude_scale=0.322*, *azimuth_scale=0.383*)

Calculate annotated region metrics

allensdk.internal.brain_observatory.annotated_region_metrics.**retinotopy_metric**(*mask*, *isi_map*)

Compute retinotopic metrics for a responding area

Parameters

mask
[numpy.ndarray] Mask representing the area over which to calculate metrics

isi_map
[numpy.ndarray] Retinotopic map

Returns

(float, float, float, float) tuple
min, max, range, bias of retinotopic map over masked region

allensdk.internal.brain_observatory.demix_report module

allensdk.internal.brain_observatory.demix_report.**background_trace**(*trace*, *save_dir*, *data_set=None*)

allensdk.internal.brain_observatory.demix_report.**compute_correlations**(*dm*, *movie_path*, *movie_dataset*)

allensdk.internal.brain_observatory.demix_report.**compute_correlations_without_masks**(*dm*)

allensdk.internal.brain_observatory.demix_report.**compute_non_overlap_masks**(*dm*)

allensdk.internal.brain_observatory.demix_report.**compute_non_overlap_traces**(*dm*, *movie_path*, *movie_dataset*)

allensdk.internal.brain_observatory.demix_report.**correlation_report**(*dm*, *save_dir*, *without_masks=True*)

parameters:
dm: [DeMix object] without_masks: boolean

allensdk.internal.brain_observatory.demix_report.**plot_masks**(*dm*, *save_dir*, *movie_file*, *movie_dataset*, *window=150*, *add_background=True*)

allensdk.internal.brain_observatory.demixer module

allensdk.internal.brain_observatory.demixer.**demix_time_dep_masks**(*raw_traces*, *stack*, *masks*)

Parameters

- **raw_traces** – extracted traces
- **stack** – movie (same length as traces)
- **masks** – binary roi masks

Returns
demixed traces

allensdk.internal.brain_observatory.demixer.**find_negative_baselines**(*trace*)

allensdk.internal.brain_observatory.demixer.**find_negative_transients_threshold**(*trace*, *window=500*, *length=10*, *std_devs=3*)

allensdk.internal.brain_observatory.demixer.**find_zero_baselines**(*traces*)

allensdk.internal.brain_observatory.demixer.**identify_valid_masks**(*mask_array*)

allensdk.internal.brain_observatory.demixer.**plot_negative_baselines**(*raw_traces*, *demix_traces*, *mask_array*, *roi_ids_mask*, *plot_dir*, *ext='png'*)

allensdk.internal.brain_observatory.demixer.**plot_negative_transients**(*raw_traces*, *demix_traces*, *valid_roi*, *mask_array*, *roi_ids_mask*, *plot_dir*, *ext='png'*)

allensdk.internal.brain_observatory.demixer.**plot_overlap_masks_lengthOne**(*roi_ind*, *masks*, *savefile=None*, *weighted=False*)

allensdk.internal.brain_observatory.demixer.**plot_traces**(*raw_trace*, *demix_trace*, *roi_id*, *roi_ind*, *save_file*)

allensdk.internal.brain_observatory.demixer.**plot_transients**(*roi_ind*, *t_trans*, *masks*, *traces*, *demix_traces*, *savefile*)

allensdk.internal.brain_observatory.demixer.**rolling_window**(*trace*, *window=500*)

Parameters

- **trace** –
- **window** –

Returns

allensdk.internal.brain_observatory.eye_calibration module

class allensdk.internal.brain_observatory.eye_calibration.**EyeCalibration**(*monitor_position=array([11.86, 8.62, 3.16])*, *monitor_rotations=array([0., 0., 0.])*, *led_position=array([25.89, -6.12, 3.21])*, *camera_position=array([13., 0., 0.])*, *camera_rotations=array([0., 0., 0.22863813])*, *eye_radius=0.1682*, *cm_per_pixel=0.001019999999999999*

Bases: object

Class for performing eye-tracking calibration.

Provides methods for estimating the position of the pupil in 3D space and projecting the gaze onto the monitor in both 3D space and monitor space given the experimental geometry.

Parameters

monitor_position
[numpy.ndarray] [x,y,z] position of monitor in cm.

monitor_rotations
[numpy.ndarray] [x,y,z] rotations of monitor in radians.

led_position
[numpy.ndarray] [x,y,z] position of LED in cm.

camera_position
[numpy.ndarray] [x,y,z] position of camera in cm.

camera_rotations
[numpy.ndarray] [x,y,z] rotations for camera in radians. X and Y must be 0.

eye_radius
[float] Radius of the eye in cm.

cm_per_pixel
[float] Pixel size of eye-tracking camera.

`compute_area`(*pupil_parameters*)

Compute the area of the pupil.

Assume the pupil is a circle, and that as it moves off-axis with the camera the observed ellipse major axis remains the diameter of the circle.

Parameters

pupil_parameters
[numpy.ndarray] [nx5] array of pupil parameters.

Returns

numpy.ndarray
[nx1] array of pupil areas in estimated pixels.

`static cr_position_in_mouse_eye_coordinates`(*led_position*, *eye_radius*)

Determine the 3D position of the corneal reflection.

The eye is modeled as a spherical mirror, so the reflection appears to be half the radius of the eye from the origin along the eye-LED axis.

Parameters

led_position
[numpy.ndarray] [x,y,z] position of the LED in eye coordinates.

eye_radius
[float] Radius of the eye in centimeters.

Returns

numpy.ndarray
[x,y,z] location of the corneal reflection in eye coordinates.

`pupil_position_in_mouse_eye_coordinates`(*pupil_parameters*, *cr_parameters*)

Compute the 3D pupil position in mouse eye coordinates.

Parameters

pupil_parameters
[numpy.ndarray] Array of pupil parameters for each eye tracking frame.

cr_paramaeters
[numpy.ndarray] Array of corneal reflection parameters for each eye tracking frame.

Returns

numpy.ndarray
Pupil position estimates in eye coordinates.

`pupil_position_on_monitor_in_cm`(*pupil_parameters*, *cr_parameters*)

Compute the pupil position on the monitor in cm.

Parameters

pupil_parameters
[numpy.ndarray] Array of pupil parameters for each eye tracking frame.

cr_paramaeters
[numpy.ndarray] Array of corneal reflection parameters for each eye tracking frame.

Returns

numpy.ndarray
Pupil position estimates in eye coordinates.

pupil_position_on_monitor_in_degrees(*pupil_parameters*, *cr_parameters*)

Get pupil position on monitor measured in visual degrees.

Parameters

pupil_parameters
[numpy.ndarray] Array of pupil parameters for each eye tracking frame.

cr_paramaeters
[numpy.ndarray] Array of corneal reflection parameters for each eye tracking frame.

Returns

numpy.ndarray
Pupil position estimate in visual degrees.

allensdk.internal.brain_observatory.eye_calibration.**base_object_to_eye_rotation_matrix**(*object_position*)

Rotation matrix to rotate base object frame to eye coordinates.

By convention, any other object's coordinate frame before rotations is set with positive Z pointing from the object's position back to the origin of the eye coordinate system, with X parallel to the eye X-Y plane.

Parameters

object_position
[np.ndarray] [x, y, z] position of object in eye coordinates.

Returns

numpy.ndarray
[3x3] rotation matrix.

allensdk.internal.brain_observatory.eye_calibration.**object_norm_eye_coordinates**(*object_position*, *x_rotation*, *y_rotation*, *z_rotation*)

Get the normal vector for the object plane in eye coordinates.

Parameters

object_position
[numpy.ndarray] [x, y, z] location of the object in eye coordinates.

x_rotation
[float] Rotation about the x-axis in radians.

y_rotation
[float] Rotation about the y-axis in radians.

z_rotation
[float] Rotation about the z-axis in radians.

Returns

numpy.ndarray
Endpoint of the object plane vector in eye coordinates.

allensdk.internal.brain_observatory.eye_calibration.**object_rotation_matrix**(*x_rotation*, *y_rotation*, *z_rotation*)

Rotation matrix in object coordinate frame.

The rotation matrix for rotating the object coordinate frame from the initial position. This is done by rotating around x, then around y', then around z".

Parameters

x_rotation
[float] Rotation about x axis in radians.

y_rotation
[float] Rotation about y axis in radians.

z_rotation
[float] Rotation about z axis in radians.

Returns

numpy.ndarray
[3x3] rotation matrix.

allensdk.internal.brain_observatory.eye_calibration.**project_to_plane**(*plane_normal*, *plane_point*, *points*)

Project from the origin through points onto a plane.

Parameters

plane_normal
[numpy.ndarray] [x, y, z] normal unit vector to the plane.

plane_point
[numpy.ndarray] [x, y, z] point on the plane.

points
[numpy.ndarray] [nx3] points in space through which to project.

Returns

numpy.ndarray
[nx3] points projected on the plane.

allensdk.internal.brain_observatory.fit_ellipse module

class allensdk.internal.brain_observatory.fit_ellipse.**FitEllipse**(*min_points*, *max_iter*, *threshold*, *num_close*)

Bases: object

choose_inliers(*candidate_points*)

fit_ellipse(*inlier_points*)

outlier_cost(*outlier_points*, *params*)

ransac_fit(*candidate_points*)

allensdk.internal.brain_observatory.fit_ellipse.**ellipse_angle_of_rotation**(*a*)

allensdk.internal.brain_observatory.fit_ellipse.**ellipse_angle_of_rotation2**(*a*)

allensdk.internal.brain_observatory.fit_ellipse.**ellipse_axis_length**(*a*)

allensdk.internal.brain_observatory.fit_ellipse.**ellipse_center**(*a*)

allensdk.internal.brain_observatory.fit_ellipse.**fit_ellipse**(*candidate_points*)

allensdk.internal.brain_observatory.fit_ellipse.**rotate_vector**(*y*, *x*, *theta*)

allensdk.internal.brain_observatory.fit_ellipse.**test_fit**()

allensdk.internal.brain_observatory.frame_stream module

class allensdk.internal.brain_observatory.frame_stream.**CvInputStream**(*movie_path*, *num_frames=None*, *block_size=1*, *cache_frames=False*)

Bases: object

close()

open()

class allensdk.internal.brain_observatory.frame_stream.**FfmpegInputStream**(*movie_path*, *frame_shape*, *ffmpeg_bin='ffmpeg'*, *num_frames=None*, *block_size=1*, *cache_frames=False*, *process_frame_cb=None*)

Bases: *FrameInputStream*

close()

create_images(*output_directory*, *image_type*)

open()

class allensdk.internal.brain_observatory.frame_stream.**FfmpegOutputStream**(*frame_shape*, *ffmpeg_bin='ffmpeg'*, *block_size=1*)

Bases: *FrameOutputStream*

close()

open(*movie_path*)

class allensdk.internal.brain_observatory.frame_stream.**FrameInputStream**(*movie_path*, *num_frames=None*, *block_size=1*, *cache_frames=False*, *process_frame_cb=None*)

Bases: object

close()

create_images(*output_directory*, *image_type*)

open()

class allensdk.internal.brain_observatory.frame_stream.**FrameOutputStream**(*block_size=1*)

Bases: object

close()

open(*movie_path*)

write(*frame*)

class allensdk.internal.brain_observatory.frame_stream.**ImageOutputStream**(*block_size=1*)

Bases: *FrameOutputStream*

allensdk.internal.brain_observatory.itracker module

allensdk.internal.brain_observatory.itracker_utils module

allensdk.internal.brain_observatory.itracker_utils.**default_ray**(*n*)

allensdk.internal.brain_observatory.itracker_utils.**eccentricity**(*a1*, *a2*)

allensdk.internal.brain_observatory.itracker_utils.**filter_bad_params**(*params*, *frame_width*, *frame_height*)

Replace positions outside image with nan

allensdk.internal.brain_observatory.itracker_utils.**generate_rays**(*image_array*, *seed_pixel*)

allensdk.internal.brain_observatory.itracker_utils.**initial_cr_point**(*image_array*, *bbox=None*)

bbox is a tuple of (xmin, xmax, ymin, ymax)

allensdk.internal.brain_observatory.itracker_utils.**initial_pupil_point**(*image_array*, *bbox=None*)

bbox is a tuple of (xmin, xmax, ymin, ymax)

allensdk.internal.brain_observatory.itracker_utils.**medfilt_custom**(*x*, *kernel_size=3*)

This median filter returns 'nan' whenever any value in the kernal width is 'nan' and the median otherwise

allensdk.internal.brain_observatory.itracker_utils.**median_absolute_deviation**(*a*, *consistency_constant=1.4826*)

Calculate the median absolute deviation of a univariate dataset.

Parameters

a
[numpy.ndarray] Sample data.

consistency_constant
[float] Constant to make the MAD a consistent estimator of the population standard deviation (1.4826 for a normal distribution).

Returns

float
Median absolute deviation of the data.

allensdk.internal.brain_observatory.itracker_utils.**post_process_cr**(*cr_params*)

This will replace questionable values of the CR x and y position with 'nan'

1) threshold ellipse area by 99th percentile area distribution
2) median filter using custom median filter
3) remove deviations from discontinuous jumps

The 'nan' values likely represent obscured CRs, secondary reflections, merges with the secondary reflection, or visual distortions due to the whisker or deformations of the eye

allensdk.internal.brain_observatory.itracker_utils.**post_process_pupil**(*pupil_params*)

Filter pupil parameters to replace outliers with nan

Parameters

pupil_params
[numpy.ndarray] (Nx5) array of pupil parameters [x, y, angle, axis1, axis2].

Returns

numpy.ndarray
Pupil parameters with outliers replaced with nan

allensdk.internal.brain_observatory.itracker_utils.**rotate_ray**(*ray*, *theta*)

allensdk.internal.brain_observatory.itracker_utils.**sobel_grad**(*image_array*)

allensdk.internal.brain_observatory.mask_set module

class allensdk.internal.brain_observatory.mask_set.**MaskSet**(*masks*)

Bases: object

close(*mask_idxs*, *max_dist*)

close_sets(*set_size*, *max_dist*)

property count

detect_duplicates(*overlap_threshold*)

detect_unions(*set_size=2*, *max_dist=10*, *threshold=0.7*)

distance(*mask_idxs*)

intersection(*mask_idxs*)

intersection_size(*mask_idxs*)

mask(*mask_idx*)

mask_is_union_of_set(*mask_idx*, *set_idxs*, *threshold*)

overlap_fraction(*idx0*, *idx1*)

size(*mask_idx*)

union(*mask_idxs*)

union_size(*mask_idxs*)

allensdk.internal.brain_observatory.mask_set.**bb_dist**(*bbs*)

allensdk.internal.brain_observatory.mask_set.**make_bbs**(*masks*)

allensdk.internal.brain_observatory.mouse module

allensdk.internal.brain_observatory.ophys_session_decomposition module

allensdk.internal.brain_observatory.ophys_session_decomposition.**export_frame_to_hdf5**(*raw_filename*, *data_hdf5_filename*, *auxiliary_hdf5_filename*, *frame_meta*, *compression='gzip'*, *compression_opts=9*)

Export a frame from raw to hdf5.

Data with the channel_description *data* is stored in the data_hdf5_filename, while any other data is stored in the auxiliary_hdf5_filename

allensdk.internal.brain_observatory.ophys_session_decomposition.**load_frame**(*raw_filename*, *json_meta*, *use_memmap=False*)

Load a frame of a multi-frame raw file.

allensdk.internal.brain_observatory.ophys_session_decomposition.**open_view_on_binary**(*file_like*, *dtype=<class 'numpy.uint8'>*, *mode='r'*, *offset=0*, *shape=None*, *order='C'*, *strides=None*)

Open a view into a memory-mapped binary file.

Parameters

file_like
[{string, file object}] File to open.

dtype
[numpy.dtype] Numpy dtype to open the memory-mapped array as.

mode
[string] Mode to open the file in.

offset
[integer] Offset (in bytes) into the file at which to start the memory map.

shape
[{tuple, list}] Shape of the array.

order
[{"C", "F"}] C or Fortran ordering.

strides
[{tuple, list}] Strides along each axis for reading the array.

Returns

numpy.memmap
Strided view into memory-mapped array.

allensdk.internal.brain_observatory.ophys_session_decomposition.**read_strided**(*filename*, *dtype*, *offset*, *shape*, *strides*)

Load a frame without memory-mapping.

allensdk.internal.brain_observatory.roi_filter module

allensdk.internal.brain_observatory.roi_filter_utils module

allensdk.internal.brain_observatory.roi_filter_utils.**CRITERIA**()

class allensdk.internal.brain_observatory.roi_filter_utils.**TrainingLabelClassifier**(*criteria*)

Bases: object

Very basic threshold_based classifier.

Has a decision function that is just the number of distinct criteria met by the classifier. Criteria are defined as a list of strings used with pandas.DataFrame.eval.

Parameters

criteria
[list] List of evaluation strings.

decision_function(*X*)

Get the distance from the decision boundary.

Parameters

X
[array-like] Features for each ROI.

Returns

T
[array-like] Distance for each sample from the decision boundary.

class allensdk.internal.brain_observatory.roi_filter_utils.**TrainingMultiLabelClassifier**(*criteria=None*)

Bases: object

Multilabel classifier using groups of TrainingLabelClassifiers.

This was used to generate labeling for training the original SVM for classification.

Parameters

criteria
[dictionary] Label names and criteria for each label.

get_eXcluded(*X*)

Get the calculated value of the eXcluded column.

This is useful for comparison with the original classifier implementation.

Parameters

X
[pandas.DataFrame] Object features from the object list file.

Returns

numpy.ndarray
Calculated eXcluded score from the classifier.

label_data(*X*, *as_columns=True*)

Generate labels for each row in X.

Parameters

X
[pandas.DataFrame] Object features from the object list file.

Returns

numpy.ndarray
Array of label codes representing the combination of labels found for each row.

allensdk.internal.brain_observatory.roi_filter_utils.**calculate_max_border**(*motion_df*, *max_shift*)

Calculate motion boundary from frame offsets.

When the motion correction algorithm fails to find sufficient matches, it generates very large frame offsets. The use of *max_shift* avoids filtering too many cells due to the large offsets, with the tradeoff that those frames will be noise.

Parameters

motion_df
[pandas.DataFrame] Dataframe containing the x, y offsets from motion correction.

max_shift
[float] Maximum shift to allow when considering motion correction. Any larger shifts are considered outliers.

Returns

list
[right_shift, left_shift, down_shift, up_shift]

allensdk.internal.brain_observatory.roi_filter_utils.**get_indices_by_distance**(*object_list_points*, *mask_points*)

Find indices of nearest neighbor matches.

Require a distance of 0 (perfect match) and a unique match between masks and object_list entries.

allensdk.internal.brain_observatory.roi_filter_utils.**get_rois**(*segmentation_stack*, *border=None*)

Extract a list of rois from the segmentation data array.

Parameters

segmentation_stack
[numpy.ndarray] The array from the maxInt_masks file showing the object masks.

border
[list] [right_shift, left_shift, down_shift, up_shift] bounding box determined from motion correction.

Returns

list
List of RoiMask objects.

allensdk.internal.brain_observatory.roi_filter_utils.**order_rois_by_object_list**(*object_data*, *rois*)

Reorder rois by matching bounding boxes to object list.

Parameters

object_data
[pandas.DataFrame] Object list data.

rois
[list] List of RoiMasks.

Returns

list
The list of rois reordered to index the same as object_data.

allensdk.internal.brain_observatory.run_itracker module

allensdk.internal.brain_observatory.time_sync module

class allensdk.internal.brain_observatory.time_sync.**OphysTimeAligner**(*sync_file*, *scanner=None*, *dff_file=None*, *stimulus_pkl=None*, *eye_video=None*, *behavior_video=None*, *long_stim_threshold=0.2*)

Bases: `object`

property behavior_video_timestamps

property clipped_stim_timestamps

Return the stimulus timestamps with the erroneous initial spike removed (if relevant)

Returns

timestamps: np.ndarray
An array of stimulus timestamps in seconds with th emonitor delay added

delta: int
Difference between the length of timestamps and the number of frames reported in the stimulus pickle file, i.e. len(timestamps) - len(pkl_file['items']['behavior']['intervalsms']

property corrected_behavior_video_timestamps

property corrected_eye_video_timestamps

property corrected_ophys_timestamps

property corrected_stim_timestamps
The stimulus timestamps corrected for monitor delay

Returns

timestamps: np.ndarray
An array of stimulus timestamps in seconds with th emonitor delay added

delta: int
Difference between the length of timestamps and the number of frames reported in the stimulus pickle file, i.e. len(timestamps) - len(pkl_file['items']['behavior']['intervalsms']

delay: float
The monitor delay in seconds

property dataset

property eye_video_timestamps

property monitor_delay
The monitor delay (in seconds) associated with the session

property ophys_timestamps
Get the timestamps for the ophys data.

property stim_timestamps

allensdk.internal.brain_observatory.time_sync.**calculate_monitor_delay**(*sync_dset*, *stim_times*, *photodiode_key*, *transition_frame_interval=60*, *max_monitor_delay=0.07*)
Calculate monitor delay.

allensdk.internal.brain_observatory.time_sync.**corrected_video_timestamps**(*video_name*, *timestamps*, *data_length*)

allensdk.internal.brain_observatory.time_sync.**get_alignment_array**(*ref*, *other*, *int_method=<ufunc 'floor'>*)
Generate an alignment array

allensdk.internal.brain_observatory.time_sync.**get_keys**(*sync_dset:* Dataset) → dict
Gets the correct keys for the sync file by searching the sync file line labels. Removes key from the dictionary if it is not in the sync dataset line labels. Args:

sync_dset: The sync dataset to search for keys within

Returns:

key_dict: dictionary of key value pairs for finding data in the
sync file

allensdk.internal.brain_observatory.time_sync.**get_ophys_data_length**(*filename*)

allensdk.internal.brain_observatory.time_sync.**get_photodiode_events**(*sync_dset*, *photodiode_key*)

Returns the photodiode events with the start/stop indicators and the window init flash stripped off. These transitions occur roughly ~1.0s apart, since the sync square changes state every N frames (where N = 60, and frame rate is 60 Hz). Because there are no markers for when the first transition of this type started, we estimate based on the event intervals. For the first valid event, find the first two events that both meet the following criteria:

The next event occurs ~1.0s later

First the last valid event, find the first two events that both meet the following criteria:

The last valid event occured ~1.0s before

allensdk.internal.brain_observatory.time_sync.**get_real_photodiode_events**(*sync_dset*, *photodiode_key*, *anomaly_threshold=0.5*)

Gets the photodiode events with the anomalies removed.

allensdk.internal.brain_observatory.time_sync.**get_stim_data_length**(*filename: str*) → int

Get stimulus data length from .pkl file.

Parameters

filename
[str] Path of stimulus data .pkl file.

Returns

int
Stimulus data length.

allensdk.internal.brain_observatory.time_sync.**get_video_length**(*filename*)

Module contents

allensdk.internal.core package

Submodules

allensdk.internal.core.lims_pipeline_module module

class allensdk.internal.core.lims_pipeline_module.**PipelineModule**(*description=''*, *parser=None*)

Bases: object

property args

input_data()

write_output_data(*data*)

allensdk.internal.core.lims_pipeline_module.**default_argument_parser**(*description=''*)

allensdk.internal.core.lims_pipeline_module.**run_module**(*module*, *input_data*, *storage_directory*, *optional_args=None*, *python='/shared/utils.x86_64/python-2.7/bin/python'*, *sdk_path='/shared/bioapps/infoapps/lims2_modules/lib/allensd*
local=False, *pbs=None*)

allensdk.internal.core.lims_utilities module

allensdk.internal.core.lims_utilities.**append_well_known_file**(*wkfs*, *path*, *wkf_type_id=None*, *content_type=None*)

allensdk.internal.core.lims_utilities.**connect**(*user='limsreader'*, *host='limsdb2'*, *database='lims2'*, *password='limsro'*, *port=5432*)

allensdk.internal.core.lims_utilities.**convert_from_titan_linux**(*file_name*)

allensdk.internal.core.lims_utilities.**get_input_json**(*object_id*, *object_class*, *strategy_class*, *host='lims2'*, ***kwargs*)

allensdk.internal.core.lims_utilities.**get_well_known_file_by_name**(*wkfs*, *filename*)

allensdk.internal.core.lims_utilities.**get_well_known_file_by_type**(*wkfs*, *wkf_type_id*)

allensdk.internal.core.lims_utilities.**get_well_known_files_by_name**(*wkfs*, *filename*)

allensdk.internal.core.lims_utilities.**get_well_known_files_by_type**(*wkfs*, *wkf_type_id*)

allensdk.internal.core.lims_utilities.**linux_to_windows**(*file_name*)

allensdk.internal.core.lims_utilities.**query**(*query*, *user='limsreader'*, *host='limsdb2'*, *database='lims2'*, *password='limsro'*, *port=5432*)

allensdk.internal.core.lims_utilities.**safe_system_path**(*file_name*)

allensdk.internal.core.lims_utilities.**select**(*cursor*, *query*)

allensdk.internal.core.mouse_connectivity_cache_prerelease module

class allensdk.internal.core.mouse_connectivity_cache_prerelease.**MouseConnectivityCachePrerelease**(*resolut*
cache=
man-
i-
fest_fil
ccf_ver
ver-
sion=N
*cache_*
stor-
age_di

Bases: *MouseConnectivityCache*

Extends MouseConnectivityCache to use prereleased data from lims.

Parameters

resolution: int
Resolution of grid data to be downloaded when accessing projection volume, the annotation volume, and the annotation volume. Must be one of (10, 25, 50, 100). Default is 25.

ccf_version: string
Desired version of the Common Coordinate Framework. This affects the annotation volume (get_annotation_volume) and structure masks (get_structure_mask). Must be one of (MouseConnectivityApi.CCF_2015, MouseConnectivityApi.CCF_2016). Default: MouseConnectivityApi.CCF_2016

cache: boolean
Whether the class should save results of API queries to locations specified in the manifest file. Queries for files (as opposed to metadata) must have a file location. If caching is disabled, those locations must be specified in the function call (e.g. get_projection_density(file_name='file.nrrd')).

manifest_file: string
File name of the manifest to be read. Default is "mouse_connectivity_manifest.json".

Attributes

resolution: int
Resolution of grid data to be downloaded when accessing projection volume, the annotation volume, and the annotation volume. Must be one of (10, 25, 50, 100). Default is 25.

api: MouseConnectivityApiPrerelease instance
Used internally to make API queries.

`EXPERIMENTS_PRERELEASE_KEY = 'EXPERIMENTS_PRERELEASE'`

`STORAGE_DIRECTORIES_PRERELEASE_KEY = 'STORAGE_DIRECTORIES_PRERELEASE'`

`add_manifest_paths`(*manifest_builder*)

Construct a manifest for this Cache class and save it in a file.

Parameters

file_name: string
File location to save the manifest.

`filter_experiments`(*experiments*, *cre=None*, *injection_structure_ids=None*, *age=None*, *gender=None*, *workflow_state=None*, *workflows=None*, *project_code=None*)

Take a list of experiments and filter them by cre status and injection structure.

Parameters

cre: boolean or list
If True, return only cre-positive experiments. If False, return only cre-negative experiments. If None, return all experients. If list, return all experiments with cre line names in the supplied list. Default None.

injection_structure_ids: list
Only return experiments that were injected in the structures provided here. If None, return all experiments. Default None.

age
[list] Only return experiments with specimens with ages provided here. If None, returna all experiments. Default None.

get_experiments(*dataframe=False*, *file_name=None*, *cre=None*, *injection_structure_ids=None*, *age=None*, *gender=None*, *workflow_state=None*, *workflows=None*, *project_code=None*)

Read a list of experiments.

If caching is enabled, this will save the whole (unfiltered) list of experiments to a file.

Parameters

dataframe: boolean
Return the list of experiments as a Pandas DataFrame. If False, return a list of dictionaries. Default False.

file_name: string
File name to save/read the structures table. If file_name is None, the file_name will be pulled out of the manifest. If caching is disabled, no file will be saved. Default is None.

allensdk.internal.core.simpletree module

class allensdk.internal.core.simpletree.**SimpleTree**(*nodes*, *node_id_cb*, *parent_id_cb*)

Bases: object

ancestor_ids(*nid*)

ancestors(*nid*)

child_ids(*nid*)

children(*nid*)

descendant_ids(*nid*)

descendants(*nid*)

node(*nid*)

node_ids()

nodes(*nids=None*)

parent(*nid*)

parent_id(*nid*)

allensdk.internal.core.swc module

class allensdk.internal.core.swc.**Marker**(**args*, ***kwargs*)

Bases: dict

Simple dictionary class for handling reconstruction marker objects.

CUT_DENDRITE = 10

NO_RECONSTRUCTION = 20

SPACING = [0.1144, 0.1144, 0.28]

allensdk.internal.core.swc.**read_marker_file**(*file_name*)

read in a marker file and return a list of dictionaries

allensdk.internal.core.swc.**read_swc**(*file_name*)

Read in an SWC file and return a Morphology object.

Parameters

file_name: string
SWC file name.

Returns

Morphology
A Morphology instance.

Module contents

allensdk.internal.core provides general modules for interacting with on-prem resources

allensdk.internal.ephys package

Submodules

allensdk.internal.ephys.core_feature_extract module

allensdk.internal.ephys.core_feature_extract.**extract_data**(*data*, *nwb_file*)

allensdk.internal.ephys.core_feature_extract.**filter_sweeps**(*sweeps*, *types=None*, *passed_only=True*, *iclamp_only=True*)

allensdk.internal.ephys.core_feature_extract.**filtered_sweep_numbers**(*sweeps*, *types=None*, *passed_only=True*, *iclamp_only=True*)

allensdk.internal.ephys.core_feature_extract.**find_coarse_long_square_amp_delta**(*sweeps*, *decimals=0*)

Find the delta between amplitudes of coarse long square sweeps. Includes failed sweeps.

allensdk.internal.ephys.core_feature_extract.**find_stim_start**(*stim*, *idx0=0*)

Find the index of the first nonzero positive or negative jump in an array.

Parameters

stim: np.ndarray
Array to be searched

idx0: int
Start searching with this index (default: 0).

Returns

int

allensdk.internal.ephys.core_feature_extract.**find_sweep_stim_start**(*data_set*, *sweep_number*)

allensdk.internal.ephys.core_feature_extract.**generate_output_cell_features**(*cell_features*, *sweep_features*, *sweep_index*)

allensdk.internal.ephys.core_feature_extract.**nan_get**(*obj*, *key*)

Return a value from a dictionary. If it does not exist, return None. If it is NaN, return None

allensdk.internal.ephys.core_feature_extract.**save_qc_figures**(*qc_fig_dir*, *nwb_file*, *output_data*, *plot_cell_figures*)

allensdk.internal.ephys.core_feature_extract.**update_output_sweep_features**(*cell_features*, *sweep_features*, *sweep_index*)

allensdk.internal.ephys.plot_qc_figures module

allensdk.internal.ephys.plot_qc_figures.**exp_curve**(*x*, *a*, *inv_tau*, *y0*)

Function used for tau curve fitting

allensdk.internal.ephys.plot_qc_figures.**get_features**(*sweep_features*, *sweep_number*)

allensdk.internal.ephys.plot_qc_figures.**get_spikes**(*sweep_features*, *sweep_number*)

allensdk.internal.ephys.plot_qc_figures.**get_time_string**()

allensdk.internal.ephys.plot_qc_figures.**load_experiment**(*file_name*, *sweep_number*)

allensdk.internal.ephys.plot_qc_figures.**main**()

allensdk.internal.ephys.plot_qc_figures.**make_cell_html**(*image_files*, *ephys_roi_result*, *file_name*, *relative_sweep_link*)

allensdk.internal.ephys.plot_qc_figures.**make_cell_page**(*nwb_file*, *ephys_roi_result*, *working_dir*, *save_cell_plots=True*)

allensdk.internal.ephys.plot_qc_figures.**make_sweep_html**(*sweep_files*, *file_name*)

allensdk.internal.ephys.plot_qc_figures.**make_sweep_page**(*nwb_file*, *ephys_roi_result*, *working_dir*)

allensdk.internal.ephys.plot_qc_figures.**mask_nulls**(*data*)

allensdk.internal.ephys.plot_qc_figures.**plot_cell_figures**(*nwb_file*, *ephys_roi_result*, *image_dir*, *sizes*)

allensdk.internal.ephys.plot_qc_figures.**plot_fi_curve_figures**(*nwb_file*, *cell_features*, *lims_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures.**plot_hero_figures**(*nwb_file*, *cell_features*, *lims_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures.**plot_images**(*ephys_roi_result*, *image_dir*, *sizes*, *image_sets*)

allensdk.internal.ephys.plot_qc_figures.**plot_instantaneous_threshold_thumbnail**(*nwb_file*, *sweep_numbers*, *cell_features*, *lims_features*, *sweep_features*, *color='red'*)

allensdk.internal.ephys.plot_qc_figures.**plot_long_square_summary**(*nwb_file*, *cell_features*, *lims_features*, *sweep_features*)

allensdk.internal.ephys.plot_qc_figures.**plot_ramp_figures**(*nwb_file*, *cell_specimen*, *cell_features*, *lims_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures.**plot_rheo_figures**(*nwb_file*, *cell_features*, *lims_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures.**plot_sag_figures**(*nwb_file*, *cell_features*, *lims_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures.**plot_short_square_figures**(*nwb_file*, *cell_features*, *lims_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures.**plot_single_ap_values**(*nwb_file*, *sweep_numbers*, *lims_features*, *sweep_features*, *cell_features*, *type_name*)

allensdk.internal.ephys.plot_qc_figures.**plot_subthreshold_long_square_figures**(*nwb_file*, *cell_features*, *lims_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures.**plot_sweep_figures**(*nwb_file*, *ephys_roi_result*, *image_dir*, *sizes*)

allensdk.internal.ephys.plot_qc_figures.**plot_sweep_set_summary**(*nwb_file*, *highlight_sweep_number*, *sweep_numbers*, *highlight_color='#0779BE'*, *background_color='#dddddd'*)

allensdk.internal.ephys.plot_qc_figures.**plot_sweep_value_figures**(*cell_specimen*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures.**save_figure**(*fig*, *image_name*, *image_set_name*, *image_dir*, *sizes*, *image_sets*, *scalew=1*, *scaleh=1*, *ext='jpg'*)

allensdk.internal.ephys.plot_qc_figures3 module

allensdk.internal.ephys.plot_qc_figures3.**exp_curve**(*x*, *a*, *inv_tau*, *y0*)

Function used for tau curve fitting

allensdk.internal.ephys.plot_qc_figures3.**get_features**(*sweep_features*, *sweep_number*)

allensdk.internal.ephys.plot_qc_figures3.**get_spikes**(*sweep_features*, *sweep_number*)

allensdk.internal.ephys.plot_qc_figures3.**get_time_string**()

allensdk.internal.ephys.plot_qc_figures3.**load_experiment**(*file_name*, *sweep_number*)

allensdk.internal.ephys.plot_qc_figures3.**make_cell_html**(*image_files*, *file_name*, *relative_sweep_link*, *specimen_info*, *fields*)

allensdk.internal.ephys.plot_qc_figures3.**make_cell_page**(*nwb_file*, *cell_features*, *rheo_features*, *sweep_features*, *sweep_info*, *well_known_files*, *specimen_info*, *working_dir*, *fields_to_show*, *save_cell_plots=True*)

nwb_file: name of nwb file (string)

cell_features:

rheo_features: dict containing extracted features from rheobase sweep

sweep_features:

sweep_info:

well_known_files: LIMS-output information containing graphics file names

working_dir:

save_cell_plots:

allensdk.internal.ephys.plot_qc_figures3.**make_sweep_html**(*sweep_files*, *file_name*)

allensdk.internal.ephys.plot_qc_figures3.**make_sweep_page**(*nwb_file*, *working_dir*, *sweep_data*)

allensdk.internal.ephys.plot_qc_figures3.**mask_nulls**(*data*)

allensdk.internal.ephys.plot_qc_figures3.**plot_cell_figures**(*nwb_file*, *cell_features*, *sweep_features*, *rheo_features*, *image_dir*, *sweep_info*, *sizes*)

allensdk.internal.ephys.plot_qc_figures3.**plot_fi_curve_figures**(*nwb_file*, *cell_features*, *rheo_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures3.**plot_hero_figures**(*nwb_file*, *cell_features*, *rheo_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures3.**plot_images**(*well_known_files*, *image_dir*, *sizes*, *image_sets*)

allensdk.internal.ephys.plot_qc_figures3.**plot_instantaneous_threshold_thumbnail**(*nwb_file*, *sweep_numbers*, *cell_features*, *rheo_features*, *sweep_features*, *color='red'*)

allensdk.internal.ephys.plot_qc_figures3.**plot_long_square_summary**(*nwb_file*, *cell_features*, *rheo_features*, *sweep_features*)

allensdk.internal.ephys.plot_qc_figures3.**plot_ramp_figures**(*nwb_file*, *sweep_info*, *cell_features*, *rheo_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures3.**plot_rheo_figures**(*nwb_file*, *cell_features*, *rheo_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures3.**plot_sag_figures**(*nwb_file*, *cell_features*, *rheo_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures3.**plot_short_square_figures**(*nwb_file*, *cell_features*, *rheo_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures3.**plot_single_ap_values**(*nwb_file*, *sweep_numbers*, *rheo_features*, *sweep_features*, *cell_features*, *type_name*)

allensdk.internal.ephys.plot_qc_figures3.**plot_subthreshold_long_square_figures**(*nwb_file*, *cell_features*, *rheo_features*, *sweep_features*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures3.**plot_sweep_figures**(*nwb_file*, *sweep_data*, *image_dir*, *sizes*)

allensdk.internal.ephys.plot_qc_figures3.**plot_sweep_set_summary**(*nwb_file*, *highlight_sweep_number*, *sweep_numbers*, *highlight_color='#0779BE'*, *background_color='#dddddd'*)

allensdk.internal.ephys.plot_qc_figures3.**plot_sweep_value_figures**(*sweep_info*, *image_dir*, *sizes*, *cell_image_files*)

allensdk.internal.ephys.plot_qc_figures3.**save_figure**(*fig*, *image_name*, *image_set_name*, *image_dir*, *sizes*, *image_sets*, *scalew=1*, *scaleh=1*, *ext='jpg'*)

Module contents

allensdk.internal.model package

Subpackages

allensdk.internal.model.biophysical package

Subpackages

allensdk.internal.model.biophysical.fits package

Subpackages

allensdk.internal.model.biophysical.fits.fit_styles package

Module contents

Module contents

allensdk.internal.model.biophysical.passive_fitting package

Subpackages

allensdk.internal.model.biophysical.passive_fitting.passive package

Module contents

Submodules

allensdk.internal.model.biophysical.passive_fitting.neuron_passive_fit module

allensdk.internal.model.biophysical.passive_fitting.neuron_passive_fit.**arg_parser**()

allensdk.internal.model.biophysical.passive_fitting.neuron_passive_fit.**main**()

allensdk.internal.model.biophysical.passive_fitting.neuron_passive_fit.**process_inputs**(*parser*)

allensdk.internal.model.biophysical.passive_fitting.neuron_passive_fit2 module

allensdk.internal.model.biophysical.passive_fitting.neuron_passive_fit2.**main**()

allensdk.internal.model.biophysical.passive_fitting.neuron_passive_fit_elec module

allensdk.internal.model.biophysical.passive_fitting.neuron_passive_fit_elec.**main**()

allensdk.internal.model.biophysical.passive_fitting.neuron_utils module

allensdk.internal.model.biophysical.passive_fitting.neuron_utils.**get_h**()

allensdk.internal.model.biophysical.passive_fitting.neuron_utils.**load_morphology**(*filename*)

allensdk.internal.model.biophysical.passive_fitting.neuron_utils.**parse_neuron_output**(*output_str*)

allensdk.internal.model.biophysical.passive_fitting.neuron_utils.**read_neuron_fit_stdout**(*func*)

allensdk.internal.model.biophysical.passive_fitting.output_grabber module

class allensdk.internal.model.biophysical.passive_fitting.output_grabber.**OutputGrabber**(*stream=None*, *threaded=False*)

Bases: object

Class used to grab standard output or another stream.

escape_char = '\x08'

readOutput()

Read the stream data (one byte at a time) and save the text in *capturedtext*.

start()

Start capturing the stream data.

stop()

Stop capturing the stream data and save the text in *capturedtext*.

allensdk.internal.model.biophysical.passive_fitting.preprocess module

allensdk.internal.model.biophysical.passive_fitting.preprocess.**get_cap_check_indices**(*i*)

allensdk.internal.model.biophysical.passive_fitting.preprocess.**get_passive_fit_data**(*cap_check_sweeps*, *data_set*)

allensdk.internal.model.biophysical.passive_fitting.preprocess.**main**()

Module contents

Submodules

allensdk.internal.model.biophysical.biophysical_archiver module

class allensdk.internal.model.biophysical.biophysical_archiver.**BiophysicalArchiver**(*archive_dir=None*)

Bases: object

archive_cell(*ephys_result_id*, *specimen_id*, *template*, *neuronal_model_id*)

get_cells()

get_neuronal_models(*specimen_ids*)

get_stimulus_file(*neuronal_model_id*)

get_template_names()

allensdk.internal.model.biophysical.check_fi_shift module

allensdk.internal.model.biophysical.check_fi_shift.**calculate_fi_curves**(*data_set*, *sweeps*)

allensdk.internal.model.biophysical.check_fi_shift.**estimate_fi_shift**(*data_set*, *sweeps*)

allensdk.internal.model.biophysical.deap_utils module

class allensdk.internal.model.biophysical.deap_utils.**Utils**(*description*)

Bases: *HocUtils*

actual_parameters_from_normalized(*params*)

calculate_feature_errors(*t_ms*, *v*, *i*)

generate_morphology(*morph_filename*)

insert_iclamp()

load_cell_parameters()

normalize_actual_parameters(*params*)

record_values()

set_actual_parameters(*params*)

set_iclamp_params(*amp*, *delay*, *dur*)

set_normalized_parameters(*params*)

allensdk.internal.model.biophysical.ephys_utils module

allensdk.internal.model.biophysical.ephys_utils.**get_step_stim_characteristics**(*i*, *t*)

allensdk.internal.model.biophysical.ephys_utils.**get_sweep_v_i_t_from_set**(*data_set*, *sweep_number*)

allensdk.internal.model.biophysical.ephys_utils.**get_sweeps_of_type**(*sweep_type*, *sweeps*)

allensdk.internal.model.biophysical.fit_stage_1 module

allensdk.internal.model.biophysical.fit_stage_2 module

allensdk.internal.model.biophysical.make_deap_fit_json module

class allensdk.internal.model.biophysical.make_deap_fit_json.**Report**(*top_level_description*, *fit_type*)

Bases: object

best_fit_value()

check_org_selections_for_noise_block()

gather_from_seeds()

generate_fit_file()

make_fit_json_file()

setup_model()

allensdk.internal.model.biophysical.neuron_parallel module

allensdk.internal.model.biophysical.optimize module

allensdk.internal.model.biophysical.run_optimize module

class allensdk.internal.model.biophysical.run_optimize.**RunOptimize**(*input_json*, *output_json*)

Bases: object

copy_local()

generate_manifest_lims(*lims_json_path*, *manifest_path*)

generate_manifest_rma(*neuronal_model_id*, *manifest_path*, *api_url=None*)

info(*lims_json_path*)

return a string that a bash script can use to find the working directory, etc. to clean up.

load_manifest()

make_fit()

nrnivmodl()

start_specimen()

allensdk.internal.model.biophysical.run_optimize.**main**(*command*, *input_json*, *output_json*)

Entry point for module. :param command: select behavior, nrnivmodl or simulate :type command: string :param lims_strategy_json: path to json file output from lims. :type lims_strategy_json: string :param lims_response_json: path to json file returned to lims. :type lims_response_json: string

allensdk.internal.model.biophysical.run_optimize_workflow module

allensdk.internal.model.biophysical.run_passive_fit module

allensdk.internal.model.biophysical.run_passive_fit.**main**(*limit*, *manifest_path*)

allensdk.internal.model.biophysical.run_passive_fit.**run_passive_fit**(*description*)

allensdk.internal.model.biophysical.run_simulate_lims module

class allensdk.internal.model.biophysical.run_simulate_lims.**RunSimulateLims**(*input_json*, *output_json*)

Bases: *RunSimulate*

copy_local()

generate_manifest_lims(*lims_data_path*, *manifest_path*)

generate_manifest_rma(*neuronal_model_run_id*, *manifest_path*, *api_url=None*)

allensdk.internal.model.biophysical.run_simulate_lims.**main**(*command*, *lims_strategy_json*, *lims_response_json*)

Entry point for module. :param command: select behavior, nrnivmodl or simulate :type command: string :param lims_strategy_json: path to json file output from lims. :type lims_strategy_json: string :param lims_response_json: path to json file returned to lims. :type lims_response_json: string

allensdk.internal.model.biophysical.run_simulate_workflow module

Module contents

allensdk.internal.model.glif package

Submodules

allensdk.internal.model.glif.ASGLM module

allensdk.internal.model.glif.ASGLM.**ASGLM_pairwise**(*ks_int*, *I_stim*, *voltage*, *spike_ind*, *cinit*, *tauinit*, *SCL*, *dt*, *resting_potential*, *SHORT_RUN=False*, *MAKE_PLOT=False*, *SHOW_PLOT=False*, *BLOCK=False*)

Calculate the resistance and amplitude of the afterspike currents for Parameters ————-

ks_int: list
initial possible k's (k=1/tau, where tau is the time constant of the exponential decay)

I_stim: list of arrays
input stimulus traces of sweeps

voltage: list of arrays
voltage of cell as a result of I_stim

spike_ind: list of arrays
each array contains the index of the spikes

cinit: float
membrane capacitance

tauinit: float
time constant of membrane

SCL: float
number of indicies that should be cut after a spike

dt: float
size of time step of injected current

Returns

allensdk.internal.model.glif.MLIN module

allensdk.internal.model.glif.MLIN.**MLIN**(*voltage*, *current*, *res*, *cap*, *dt*, *MAKE_PLOT=False*, *SHOW_PLOT=False*, *BLOCK=False*, *PUBLICATION_PLOT=False*)

voltage, current input:

voltage: numpy array of voltage with test pulse cut out current: numpy array of stimulus with test pulse cut out

allensdk.internal.model.glif.MLIN.**autocorr**(*x*)

allensdk.internal.model.glif.MLIN.**exp_decay**(*time*, *amp*, *tau*)

allensdk.internal.model.glif.MLIN.**expsymm_cdf**(*v*, *dv*)

allensdk.internal.model.glif.MLIN.**expsymm_pdf**(*v*, *dv*)

allensdk.internal.model.glif.MLIN.**find_bin_center**(*edges*)

allensdk.internal.model.glif.are_two_lists_of_arrays_the_same module

allensdk.internal.model.glif.are_two_lists_of_arrays_the_same.**are_two_lists_of_arrays_the_same**(*data1*, *data2*)

returns False if to lists of arrays are different. otherwise the function returns True.

allensdk.internal.model.glif.configure_model module

allensdk.internal.model.glif.error_functions module

allensdk.internal.model.glif.error_functions.**MLIN_list_error**(*param_guess*, *experiment*, *input_data*)

allensdk.internal.model.glif.find_spikes module

allensdk.internal.model.glif.find_spikes.**align_and_cut_spikes**(*voltage_list*, *current_list*, *dt*, *spike_window=None*)

This function aligns the spikes to some criteria and returns a current and voltage trace of of the spike over a time window. Also returns zero crossing,and threshold in reference to the aligned spikes.

allensdk.internal.model.glif.find_spikes.**find_spikes_list**(*voltage_list*, *dt*)

allensdk.internal.model.glif.find_spikes.**find_spikes_list_old**(*voltage_list*, *dt*)

allensdk.internal.model.glif.find_spikes.**find_spikes_old**(*v*, *dt*)

allensdk.internal.model.glif.find_spikes.**find_spikes_ssq_list**(*voltage_list*, *dt*, *dv_cutoff*, *thresh_frac*)

allensdk.internal.model.glif.find_sweeps module

exception allensdk.internal.model.glif.find_sweeps.**MissingSweepException**

Bases: Exception

allensdk.internal.model.glif.find_sweeps.**find_long_square_sweeps**(*sweeps*)

allensdk.internal.model.glif.find_sweeps.**find_noise_sweeps**(*sweeps*)

Find 1) the noise1 sweeps
2) the noise2 sweeps 4) all noise sweeps

allensdk.internal.model.glif.find_sweeps.**find_ramp_sweeps**(*sweeps*)

Find 1) all ramp sweeps

2) all subthreshold ramps

3) all superthreshold ramps

allensdk.internal.model.glif.find_sweeps.**find_ramp_to_rheo_sweeps**(*sweeps*)

allensdk.internal.model.glif.find_sweeps.**find_ranked_sweep**(*sweep_list*, *key*, *reverse=False*)

allensdk.internal.model.glif.find_sweeps.**find_short_square_sweeps**(*sweeps*)

Find 1) all of the subthreshold short square sweeps

2) all of the superthreshold short square sweeps

3) the subthresholds short square sweep with maximum stimulus amplitude

allensdk.internal.model.glif.find_sweeps.**find_sweeps**(*sweep_list*)

allensdk.internal.model.glif.find_sweeps.**get_sweep_numbers**(*sweep_list*)

allensdk.internal.model.glif.find_sweeps.**get_sweeps_by_name**(*sweeps*, *sweep_type*)

allensdk.internal.model.glif.find_sweeps.**main**()

allensdk.internal.model.glif.find_sweeps.**organize_sweeps_by_name**(*sweeps*, *name*)

allensdk.internal.model.glif.find_sweeps.**parse_arguments**()

allensdk.internal.model.glif.glif_experiment module

class allensdk.internal.model.glif.glif_experiment.**GlifExperiment**(*neuron*, *dt*, *stim_list*, *resp_list*, *spike_time_steps*, *grid_spike_times*, *grid_spike_voltages*, *param_fit_names*, ***kwargs*)

Bases: object

neuron_parameter_count()

run(*param_guess*)

This code will run the loaded neuron model in reference to the target neuron spikes. inputs:

self: is the instance of the neuron model and parameters alone with the values of the target spikes.
NOTE the values in each array of the self.gridSpikeIndexTarge_list and the self.interpolated_spike_times are in reference to the time start of of the stim in each induvidual array (not the universal time)

param_guess: array of scalars of the values that will be inserted into the mapping function below.

returns:

voltage_list: list of array of voltage values. NOTE: IF THE MODEL NEURON SPIKES BEFORE THE TARGET THE VOLTAGE WILL
NOT BE CALCULATED THEREFORE THE RESULTING VECTOR WILL NOT BE AS LONG AS THE TARGET AND ALSO WILL NOT MAKE SENSE WITH THE STIMULUS UNLESS YOU CUT IT AND OUTPUT IT TOO.

grid_spike_times_list: interpolated_spike_time_list: an array of the actual times of the spikes. NOTE: THESE TIMES ARE CALCULATED BY ADDING THE

TIME OF THE INDIVIDUAL SPIKE TO THE TIME OF THE LAST SPIKE.

gridISIFromLastTargSpike_list: list of arrays of spike times of the model in reference to the last target (biological)
spike (not in reference to sweep start)

interpolatedISIFromLastTargSpike_list: list of arrays of spike times of the model in reference to the last target (biological)
spike (not in reference to sweep start)

voltageOfModelAtGridBioSpike_list: list of arrays of scalars that contain the voltage of the model neuron when the target or bio neuron spikes. theshOfModelAtGridBioSpike_list: list of arrays of scalars that contain the threshold of the model neuron when the target or bio neuron spikes.

run_base_model(*param_guess*)

This code will run the loaded neuron model. inputs:

self: is the instance of the neuron model and parameters alone with the values of the target spikes.
NOTE the values in each array of the self.gridSpikeIndexTarge_list and the self.interpolated_spike_times are in reference to the time start of of the stim in each induvidual array (not the universal time)

param_guess: array of scalars of the values that will be inserted into the mapping function below.

returns:

voltage_list: list of array of voltage values. NOTE: IF THE MODEL NEURON SPIKES BEFORE THE TARGET THE VOLTAGE WILL
NOT BE CALCULATED THEREFORE THE RESULTING VECTOR WILL NOT BE AS LONG AS THE TARGET AND ALSO WILL NOT MAKE SENSE WITH THE STIMULUS UNLESS YOU CUT IT AND OUTPUT IT TOO.

gridTime_list: interpolatedTime_list: an array of the actual times of the spikes. NOTE: THESE TIMES ARE CALCULATED BY ADDING THE

TIME OF THE INDIVIDUAL SPIKE TO THE TIME OF THE LAST SPIKE.

grid_ISI_list: list of arrays of spike times of the model in reference to the last target (biological)
spike (not in reference to sweep start)

interpolated_ISI_list: list of arrays of spike times of the model in reference to the last target (biological)
spike (not in reference to sweep start)

grid_spike_voltage_list: list of arrays of scalars that contain the voltage of the model neuron when the target or bio neuron spikes. grid_spike_threshold_list: list of arrays of scalars that contain the threshold of the model neuron when the target or bio neuron spikes.

set_neuron_parameters(*param_guess*)

Maps the parameter guesses to the coefficients of the model. input:

param_guess is vector of values. It is assumed that the length will be

allensdk.internal.model.glif.glif_optimizer module

class allensdk.internal.model.glif.glif_optimizer.**GlifOptimizer**(*experiment*, *dt*, *outer_iterations*, *inner_iterations*, *sigma_outer*, *sigma_inner*, *param_fit_names*, *stim*, *xtol*, *ftol*, *internal_iterations*, *bessel*, *error_function=None*, *error_function_data=None*, *init_params=None*)

Bases: `object`

evaluate(*x*, *dt_multiplier=100*)

initiate_unique_seed(*seed=None*)

randomize_parameter_values(*values*, *sigma*)

run_many(*iteration_finished_callback=None*, *seed=None*)

run_once(*param0*)

@param param0: a list of the initial guesses for the optimizer @return: tuple including parameters that optimize function and value - see fmin docs

run_once_bound(*low_bound*, *high_bound*)

@param low_bound: a scalar initial guess for the optimizer @param high_bound: a scalar high bound for the optimizer @return: tuple including parameters that optimize function and value - see fmin docs

to_dict()

allensdk.internal.model.glif.glif_optimizer_neuron module

exception allensdk.internal.model.glif.glif_optimizer_neuron.**GlifBadInitializationException**(*message*, *dv*, *step*)

Bases: Exception

Exception raised when voltage is above threshold at the beginning of a sweep. i.e. probably caused by the optimizer.

exception allensdk.internal.model.glif.glif_optimizer_neuron.**GlifNeuronException**(*message*, *data*)

Bases: Exception

Exception for catching simulation errors and reporting intermediate data.

class allensdk.internal.model.glif.glif_optimizer_neuron.**GlifOptimizerNeuron**(**args*, ***kwargs*)

Bases: *GlifNeuron*

Contains methods for running the neuron model in a "forced-spike" paradigm used during optimization.

TYPE = 'GLIF'

classmethod from_dict(*d*)

classmethod from_dict_legacy(*d*)

run_until_biological_spike(*voltage_t0*, *threshold_t0*, *AScurrents_t0*, *stimulus*, *response*, *start_index*, *after_end_index*, *bio_spike_time_steps*)

Run the neuron simulation over a segment of a stimulus given initial conditions for use in the "forced spike" optimization paradigm. [Note: the section of stimulus is meant to be between two biological neuron spikes. Thus the stimulus is during the interspike interval (ISI)]. The model is simulated until either the model spikes or the end of the segment is reached. If the model does not spike, a spike time is extrapolated past the end of the simulation segment.

This function also returns the initial conditions for the subsequent stimulus segment. In the forced spike paradigm there are several ways

Parameters

voltage_t0
[float] the current voltage of the neuron

threshold_t0
[float] the current spike threshold level of the neuron

AScurrents_t0
[np.ndarray] the current state of the afterspike currents in the neuron

stimulus
[np.ndarray] the full stimulus array (not just the segment of data being simulated)

response
[np.ndarray] the full response array (not just the segment of data being simulated)

start_index
[int] index of global stimulus at which to start simulation

after_end_index
[int] index of global stimulus *after* the last index to be simulated

bio_spike_time_steps
[list] time steps of input spikes

Returns

dict

a dictionary containing:
'voltage': simulated voltage value 'threshold': simulated threshold values 'AScurrent_matrix': afterspike current values during the simulation 'grid_model_spike_time': model spike time (in units of dt) 'interpolated_model_spike_time': model spike time (in units of dt) interpolated between time steps 'voltage_t0': reset voltage value to be used in subsequent simulation interval 'threshold_t0': reset threshold value to be used in subsequent simulation interval 'AScurrents_t0': reset afterspike current value to be used in subsequent simulation interval 'grid_bio_spike_model_voltage': model voltage at the time of the input spike 'grid_bio_spike_model_threshold': model threshold at the time of the input spike

`run_with_biological_spikes`(*stimulus*, *response*, *bio_spike_time_steps*)

Run the neuron simulation over a stimulus, but do not allow the model to spike on its own. Rather, force the simulation to spike and reset at a given set of spike indices. Dynamics rules are applied between spikes regardless of the simulated voltage and threshold values. Reset rules are applied only at input spike times. This is used during optimization to force the model to follow the spikes of biological data. The model is optimized in this way so that history effects due to spiking can be adequately modeled. For example, every time the model spikes a new set of afterspike currents will be initiated. To ensure that afterspike currents can be optimized, we force them to be initiated at the time of the biological spike.

Parameters

stimulus
[np.ndarray] vector of scalar current values

respones
[np.ndarray] vector of scalar voltage values

bio_spike_time_steps
[list] spike time step indices

Returns

dict

a dictionary containing:
'voltage': simulated voltage values, 'threshold': simulated threshold values, 'AScurrent_matrix': afterspike currents during the simulation, 'grid_model_spike_times': spike times of the model aligned to the simulation grid (when it would have spiked), 'interpolated_model_spike_times': spike times of the model linearly interpolated between time steps, 'grid_ISI': interspike interval between grid model spike times, 'interpolated_ISI': interspike interval between interpolated model spike times, 'grid_bio_spike_model_voltage': voltage of the model at biological/input spike times, 'grid_bio_spike_model_threshold': voltage of the model at biological/input spike times interpolated between time steps

to_dict()

Convert the neuron to a serializable dictionary.

allensdk.internal.model.glif.glif_optimizer_neuron.**extrapolate_model_spike_from_endpoints**(*neuron*, *voltage*, *threshold*, *voltage_t1*, *threshold_t1*, *dt*)

allensdk.internal.model.glif.glif_optimizer_neuron.**extrapolate_model_spike_from_endpoints_single_tau**(*neu*
vol
age
thr
old
vol
age
thr
old
dt)

allensdk.internal.model.glif.glif_optimizer_neuron.**extrapolate_spike_time**(*dt*, *num_time_steps*, *threshold_t0*, *threshold_t1*, *voltage_t0*, *voltage_t1*)

Given two voltage and threshold values and an interval between them, extrapolate a spike time by intersecting lines the thresholds and voltages.

allensdk.internal.model.glif.glif_optimizer_neuron.**extrapolate_spike_voltage**(*dt*, *num_time_steps*, *threshold_t0*, *threshold_t1*, *voltage_t0*, *voltage_t1*)

Given two voltage and threshold values and an interval between them, extrapolate a spike time by intersecting lines the thresholds and voltages.

allensdk.internal.model.glif.glif_optimizer_neuron.**find_first_model_spike**(*voltage*, *threshold*, *voltage_t1*, *threshold_t1*, *dt*)

allensdk.internal.model.glif.glif_optimizer_neuron.**interpolate_spike_voltage**(*dt*, *time_step*, *threshold_t0*, *threshold_t1*, *voltage_t0*, *voltage_t1*)

Given two voltage and threshold values, the dt between them and the initial time step, interpolate a spike time within the dt interval by intersecting the two lines.

allensdk.internal.model.glif.optimize_neuron module

allensdk.internal.model.glif.optimize_neuron.**get_optimize_sweep_numbers**(*sweep_index*)

allensdk.internal.model.glif.optimize_neuron.**main**()

allensdk.internal.model.glif.optimize_neuron.**optimize_neuron**(*model_config*, *sweep_index*, *nwb_file*, *save_callback=None*)

Optimizes a neuron. 1. Loads optimizer and neuron configuration data. 2. Loads the voltage trace sweeps that will be optimized 3. Configures the experiment and optimizer 4. Runs the optimizer 5. TODO: where is data saved

Parameters

model_config
[dictionary] contains values of neuron and optimizer parameters

sweep_index
[list of integers] indices (as labeled in the data configuration file) of sweeps that will be optimized

save_callback
[module] saves output

allensdk.internal.model.glif.plotting module

Written by Corinne Teeter 3-31-14

allensdk.internal.model.glif.plotting.**checkPreprocess**(*originalStim_list*, *processedStim_list*, *originalVoltage_list*, *processedVoltage_list*, *config*, *blockME=False*)

allensdk.internal.model.glif.plotting.**checkSpikeCutting**(*originalStim_list*, *cutStim_list*, *originalVoltage_list*, *cutVoltage_list*, *allindOfNonSpiking_list*, *config*, *blockME=False*)

allensdk.internal.model.glif.plotting.**plotLineRegress1**(*slope*, *intercept*, *r*, *xlim*)

allensdk.internal.model.glif.plotting.**plotLineRegressRed**(*slope*, *intercept*, *r*, *xlim*)

allensdk.internal.model.glif.plotting.**plotSpikes**(*voltage_list*, *spike_ind_list*, *dt*, *blockME=False*, *method=False*)

allensdk.internal.model.glif.preprocess_neuron module

exception allensdk.internal.model.glif.preprocess_neuron.**MissingSpikeException**

Bases: Exception

allensdk.internal.model.glif.preprocess_neuron.**estimate_dv_cutoff**(*voltage_list*, *dt*, *start_t*, *end_t*)

allensdk.internal.model.glif.preprocess_neuron.**find_first_spike_voltage**(*voltage*, *dt*, *ssq=False*, *MAKE_PLOT=False*, *SHOW_PLOT=False*, *BLOCK=False*, *dv_cutoff=20.0*, *thresh_frac=0.05*)

calculate voltage at threshold of first spike Parameters ———- voltage: numpy array

voltage trace

dt: float
sampling time step

ssq: Boolean
whether there is or is not a subrathreshold short square pulse (note that if thes

MAKE_PLOT: Boolean
specifies whether or not a plot should be made

SHOW_PLOT: Boolean
specifies if a visualization should be made

BLOCK: Boolean
if a plot is made this specifies weather to stop the code until the plot is closed

dv_cutoff: float
specifies cut off of the derivative of the voltage

thresh_frac: float
variable that goes into feature extractor

Returns

:float
voltage of threshold of first spike

allensdk.internal.model.glif.preprocess_neuron.**main**()

allensdk.internal.model.glif.preprocess_neuron.**preprocess_neuron**(*nwb_file*, *sweep_list*, *cell_properties=None*, *dt=None*, *cut=None*, *bessel=None*, *save_figure_path=None*)

allensdk.internal.model.glif.preprocess_neuron.**tag_plot**(*tag*, *fs=9*)

allensdk.internal.model.glif.rc module

allensdk.internal.model.glif.rc.**least_squares_RCEl_calc_tested**(*voltage_list*, *current_list*, *dt*)

Calculate resistance, capacitance and resting potential by performing least squares on current and voltage.

Parameters

voltage_list: list of arrays
voltage responses for several sweep repeats

current_list: list of arrays
current injections for several sweep repeats

dt: float
time step size in voltage and current traces

Returns

r_list: list of floats
each value corresponds to the resistance of a sweep

c_list: list of floats
each value corresponds to the capacitance of a sweep

el_list: list of floats
each value corresponds to the resting potential of a sweep

allensdk.internal.model.glif.spike_cutting module

allensdk.internal.model.glif.spike_cutting.**calc_spike_cut_and_v_reset_via_expvar_residuals**(*all_current_list*, *all_voltage_list*, *dt*, *El_reference*, *deltaV*, *max_spike_cut_*, *MAKE_PLOT=*, *SHOW_PLOT=*, *PUB-LI-CA-TION_PLOT=F*, *BLOCK=False*)

This function calculates where the spike should be cut based on explained variance.
The goal is to find a model where the voltage after a spike maximally explains the voltage before a spike. This will also specify the voltage reset rule inputs:

spike_determination_method: string specifing the method used to find threshold all_current_list: list of current (list of current traces injected into neuron) all_voltage_list: list of voltages (list of voltage trace)

The change is that if the slope is greater than one or intercept is greater than zero it forces it. Regardless of required force the residuals are used.

allensdk.internal.model.glif.spike_cutting.**plotLineRegress1**(*slope*, *intercept*, *r*, *xlim*)

allensdk.internal.model.glif.spike_cutting.**plotLineRegressRed**(*slope*, *intercept*, *r*, *xlim*)

allensdk.internal.model.glif.threshold_adaptation module

allensdk.internal.model.glif.threshold_adaptation.**calc_spike_component_of_threshold_from_multiblip**(*multi_*
dt,
dv_cu
thresh
MAK
SHO
BLO
PUB-
LI-
CA-
TION

Calculate the spike components of the threshold by fitting a decaying exponential function to data to threshold versus time since last spike in the multiblip data. The exponential is forced to decay to the local th_inf (calculated as the mean all of the threshold values of the first spikes in each individual triblip stimulus). For each multiblip stimulus in a stimulus set if there is more than one spike the difference in voltages from the first and second spike are plotted versus the separation in time. Note that this algorithm should only be implemented on multiblips sweeps where the neuron spike on the first and second blip. Since there is no easy way to do this, this erroneous data should not be provided to this algorithm (i.e is should be visually checked and eliminated the preprocessor should hold back this data manually for now.)

#TODO: check to see if this is still true. Notes: The standard SDK spike detection algorithm does not work with the multiblip stimulus due to artifacts when the stimulus turns on and off. Please see the find_multiblip_spikes module for more information.

Input:

multi_SS: dictionary
contains multiblip information such as current and stimulus

dt: float
time step in seconds

Returns:

const_to_add_to_thresh_for_reset: float
amplitude of the exponential fit otherwise known as a_spike. Note that this is without any spike cutting

decay_const: float
decay constant of exponential. Note the function fit is a negative exponential which will mean this value will either have to be negated when it is used or the functions used will have to have to include the negative.

thresh_inf: float

allensdk.internal.model.glif.threshold_adaptation.**exp_fit_c**(*t*, *a1*, *k1*, *const*)

allensdk.internal.model.glif.threshold_adaptation.**exp_force_c**(*t_const*, *a1*, *k1*)

allensdk.internal.model.glif.threshold_adaptation.**fit_avoltage_bvoltage**(*x*, *v_trace_list*, *El_list*, *spike_cut_length*, *all_spikeInd_list*, *th_inf*, *dt*, *a_spike*, *b_spike*, *fake=False*)

This is a version of fit_avoltage_bvoltage_debug that does not require the th_trace, v_component_of_thresh_trace, and spike_component_of_thresh_trace needed for debugging. A test should be run to make sure the same output comes out from this and the debug function

This function returns the squared error for the difference between the 'known' voltage component of the threshold obtained from the biological neuron and the voltage component of the threshold of the model obtained with the input parameters (so that the minimum can be searched for via fmin). The overall threshold is the sum of threshold

infinity the spike component of the threshold and the voltage component of the threshold. Therefore threshold infinity and the spike component of the threshold must be subtracted from the threshold of the neuron in order to isolate the voltage component of the threshold. In the evaluation of the model the actual voltage of the neuron is used so that any errors in the other components of the model will not influence the fits here (for example, if a afterspike current was estimated incorrectly)

Notes: * The spike component of the threshold is subtracted from the

voltage which means that the voltage component of the threshold should only be added to rules.

- **b_spike was fit using a negative value in the function therefore the negative is placed in the** equation.
- **values in this function are in 'real' voltage as opposed to voltage** relative to resting potential.
- **current injection during the spike is not taken into account. This seems reasonable as the** ion channels are open during this time and injected current may not greatly influence the neuron.

x: numpy array
x[0]=a_voltage input, x[1] is b_voltage_input, x[2] is th_inf

v_trace_list: list of numpy arrays
voltage traces (v_trace, El, and th_inf must be in the same frame of reference)

El_list: list of floats
reversal potential (v_trace, El, and th_inf must be in the same frame of reference)

spike_cut_length: int
number of indicies removed after initiation of a spike

all_spikeInd_list: list of numpy arrays
indicies of spike trains

th_inf: float
threshold infinity (v_trace, El, and th_inf must be in the same frame of reference)

dt: float
size of time step (SI units)

a_spike: float
amplitude of spike component of threshold.

b_spike: float
decay constant in spike component of the threshold

fake: Boolean
if True makes uses the voltage value of spike step-1 because there is not a voltage value at the spike step because it is set to nan in the simulator.

allensdk.internal.model.glif.threshold_adaptation.**fit_avoltage_bvoltage_th**(*x*, *v_trace_list*, *El_list*, *spike_cut_length*, *all_spikeInd_list*, *dt*, *a_spike*, *b_spike*, *fake=False*)

This is a version of fit_avoltage_bvoltage_th_debug that does not require the th_trace, v_component_of_thresh_trace, and spike_component_of_thresh_trace needed for debugging. A test should be run to make sure the same output comes out from this and the debug function

This function returns the squared error for the difference between the 'known' voltage component of the threshold obtained from the biological neuron and the voltage component of the threshold of the model obtained with the input parameters (so that the minimum can be searched for via fmin). The overall threshold is the sum of threshold infinity the spike component of the threshold and the voltage component of the threshold. Therefore threshold infinity and the spike component of the threshold must be subtracted from the threshold of the neuron in order to isolate the voltage component of the threshold. In the evaluation of the model the actual voltage of the neuron is used so that any errors in the other components of the model will not influence the fits here (for example, if a afterspike current was estimated incorrectly)

Notes: * The spike component of the threshold is subtracted from the

voltage which means that the voltage component of the threshold should only be added to rules.

- **b_spike was fit using a negative value in the function therefore the negative is placed in the** equation.
- **values in this function are in 'real' voltage as opposed to voltage** relative to resting potential.
- **current injection during the spike is not taken into account. This seems reasonable as the** ion channels are open during this time and injected current may not greatly influence the neuron.

x: numpy array
x[0]=a_voltage input, x[1] is b_voltage_input, x[2] is th_inf

v_trace_list: list of numpy arrays
voltage traces (v_trace, El, and th_inf must be in the same frame of reference)

El_list: list of floats
reversal potential (v_trace, El, and th_inf must be in the same frame of reference)

spike_cut_length: int
number of indicies removed after initiation of a spike

all_spikeInd_list: list of numpy arrays
indicies of spike trains

dt: float
size of time step (SI units)

a_spike: float
amplitude of spike component of threshold.

b_spike: float
decay constant in spike component of the threshold

fake: Boolean
if True makes uses the voltage value of spike step-1 because there is not a voltage value at the spike step because it is set to nan in the simulator.

`allensdk.internal.model.glif.threshold_adaptation.`**`get_peaks`**(*voltage*, *aboveValue=0*)

This function was written by Corinne Teeter and calculates the action potential peaks of a voltage equation" inputs

voltage: numpy array of voltages aboveValue: scalar voltage value over which voltage is considered a spike.

outputs:
peakInd: array of indicies of peaks

Module contents

Submodules

allensdk.internal.model.AIC module

allensdk.internal.model.AIC.**AIC**(*RSS*, *k*, *n*)

Computes the Akaike Information Criterion.

RSS-residual sum of squares of the fitting errors. k - number of fitted parameters. n - number of observations.

allensdk.internal.model.AIC.**AICc**(*RSS*, *k*, *n*)

Corrected AIC. formula from Wikipedia.

allensdk.internal.model.AIC.**BIC**(*RSS*, *k*, *n*)

Bayesian information criterion or Schwartz information criterion. Formula from wikipedia.

allensdk.internal.model.GLM module

allensdk.internal.model.GLM.**create_basis_IPSP**(*neye*, *ncos*, *kpeaks*, *ks*, *DTsim*, *t0*, *I_stim*, *nkt*, *flag_exp*, *npcut*)

allensdk.internal.model.GLM.**ff**(*x*, *c*, *dc*)

allensdk.internal.model.GLM.**invnl**(*x*)

allensdk.internal.model.GLM.**makeBasis_StimKernel**(*kbasprs*, *nkt*)

allensdk.internal.model.GLM.**makeBasis_StimKernel_exp**(*kbasprs*, *nkt*)

allensdk.internal.model.GLM.**makeFitStruct_GLM**(*dtsim*, *kbasprs*, *nkt*, *flag_exp*)

allensdk.internal.model.GLM.**nlin**(*x*)

allensdk.internal.model.GLM.**normalizecols**(*A*)

allensdk.internal.model.GLM.**sameconv**(*A*, *B*)

allensdk.internal.model.data_access module

allensdk.internal.model.data_access.**load_sweep**(*file_name*, *sweep_number*, *desired_dt=None*, *cut=0*, *bessel=False*)

load a data sweep and do specified data processing. Inputs:

file_name: string
name of .nwb data file

sweep_number:
number specifying the sweep to be loaded

desired_dt:
the size of the time step the data should be subsampled to

cut:
indicie of which to start reporting data (i.e. cut off data before this indicie)

bessel: dictionary
contains parameters 'N' and 'Wn' to implement standard python bessel filtering

Returns:

dictionary containing
voltage: array current: array dt: time step of the returned data start_idx: the index at which the first stimulus starts (excluding the test pulse)

allensdk.internal.model.data_access.**load_sweeps**(*file_name*, *sweep_numbers*, *dt=None*, *cut=0*, *bessel=False*)

load sweeps and do specified data processing. Inputs:

file_name: string
name of .nwb data file

sweep_numbers:
sweep numbers to be loaded

desired_dt:
the size of the time step the data should be subsampled to

cut:
indicie of which to start reporting data (i.e. cut off data before this indicie)

bessel: dictionary
contains parameters 'N' and 'Wn' to implement standard python bessel filtering

Returns:

dictionary containing
voltage: list of voltage trace arrays current: list of current trace arrays dt: list of time step corresponding to each array of the returned data start_idx: list of the indicies at which the first stimulus starts (excluding

the test pulse) in each returned sweep

allensdk.internal.model.data_access.**subsample_data**(*data*, *method*, *present_time_step*, *desired_time_step*)

Module contents

allensdk.internal.morphology package

Submodules

allensdk.internal.morphology.compartment module

class allensdk.internal.morphology.compartment.**Compartment**(*node1*, *node2*)

Bases: object

allensdk.internal.morphology.morphology module

class allensdk.internal.morphology.morphology.**Morphology**(*node_list=None*)

Bases: object

Keep track of the list of nodes in a morphology and provide a few helper methods (soma, tree information, pruning, etc).

APICAL_DENDRITE = 4

AXON = 2

BASAL_DENDRITE = 3

NODE_TYPES = [1, 2, 3, 4]

SOMA = 1

append(*nodes*)

Add additional nodes to this Morphology. Those nodes must originate from another morphology object.

Parameters

nodes: list of Morphology nodes

apply_affine(*aff*, *scale=None*)

Apply an affine transform to all nodes in this morphology. Compartment radius is adjusted as well.

Format of the affine matrix is:

[x0 y0 z0] [tx] [x1 y1 z1] [ty] [x2 y2 z2] [tz]

where the left 3x3 the matrix defines the affine rotation and scaling, and the right column is the translation vector.

The matrix must be collapsed and stored in a list as follows:

[x0 y0, z0, x1, y1, z1, x2, y2, z2, tx, ty, tz]

Parameters

aff: 3x4 array of floats (python 2D list, or numpy 2D array)
the transformation matrix

apply_affine_only_rotation(*aff*)

Apply an affine transform to all nodes in this morphology. Only the rotation element of the transform is performed (i.e., although the entire transformation and translation matrix is supplied, only the rotation element is used). The morphology is translated to the point where the soma root is at 0,0,0.

Format of the affine matrix is:

[x0 y0 z0] [tx] [x1 y1 z1] [ty] [x2 y2 z2] [tz]

where the left 3x3 the matrix defines the affine rotation and scaling, and the right column is the translation vector.

The matrix must be collapsed and stored in a list as follows:

[x0 y0, z0, x1, y1, z1, x2, y2, z2, tx, ty, tz]

Parameters

aff: 3x4 array of floats (python 2D list, or numpy 2D array)
the transformation matrix

change_parent(*child*, *parent*)

Change the parent of a node. The child node is adjusted to point to the new parent, the child is taken off of the previous parent's child list, and it is added to the new parent's child list.

Parameters

child: integer or Morphology Object
The ID of the child node, or the child node itself

parent: integer or Morphology Object
The ID of the parent node, or the parent node itself

Returns

Nothing

children_of(*seg*)

Returns a list of the children of the specified node

Parameters

seg: integer or Morphology Object
The ID of the parent node, or the parent node itself

Returns

A list of the child morphology objects. If the ID of the parent node is invalid, None is returned.

clone()

Create a clone (deep copy) of this morphology

compartment(*n*)

Returns the morphology Compartment having the specified ID.

Parameters

n: integer
ID of desired compartment

Returns

A morphology object having the specified ID, or None if such a node doesn't exist

property compartment_list

convert_type(*from_type*, *to_type*)

Convert all nodes in morphology from one type to another

Parameters

from_type: enum
The node type that will be eliminated and replaced. Use one of the following constants: SOMA, AXON, BASAL_DENDRITE, or APICAL_DENDRITE

to_type: enum
The new type that will replace it. Use one of the following constants: SOMA, AXON, BASAL_DENDRITE, or APICAL_DENDRITE

delete_tree(*n*)

Delete tree, and all of its nodes, from the morphology.

Parameters

n: Integer
The tree number to delete

`find`(*x*, *y*, *z*, *dist*, *node_type=None*)

Returns a list of Morphology Objects located within 'dist' of coordinate (x,y,z). If node_type is specified, the search will be constrained to return only nodes of that type.

Parameters

x, y, z: float
The x,y,z coordinates from which to search around

dist: float
The search radius

node_type: enum (optional)
One of the following constants: SOMA, AXON, BASAL_DENDRITE or APICAL_DENDRITE

Returns

A list of all Morphology Objects matching the search criteria

`get_dimensions`()

Returns tuple of overall width, height and depth of morphology. WARNING: if locations of nodes in morphology are manipulated then this value can become incorrect. It can be reset and recalculated by programmitcally setting self.dims to None.

Returns

3 real arrays: [width, height, depth], [min_x, min_y, min_z], [max_x, max_y, max_z]

`node`(*n*)

Returns the morphology node having the specified ID.

Parameters

n: integer
ID of desired node

Returns

A morphology node having the specified ID, or None if such a node doesn't exist

`property node_list`

Return the node list. This is a property to ensure that the node list and node index are in sync.

`node_list_by_type`(*node_type*)

Return an list of all nodes having the specified node type.

Parameters

node_type: int
Desired node type

Returns

A list of of Morphology Objects

`property num_nodes`

Return the number of nodes in the morphology.

property num_trees

Return the number of trees in the morphology. A tree is defined as everything following from a single root node.

parent_of(*seg*)

Returns parent of the specified node.

Parameters

seg: integer or Morphology Object
The ID of the child node, or the child node itself

Returns

A morphology object, or None if no parent exists or if the specified node ID doesn't exist

save(*file_name*)

Write this morphology out to an SWC file

Parameters

file_name: string
desired name of your SWC file

soma_root()

Returns root node of soma, if present

sparsify(*modulo*)

Return a new Morphology object that has a given number of non-leaf, non-root nodes removed.

Parameters

modulo: int
keep 1 out of every modulo nodes.

Returns

Morphology
A new morphology instance

strip_all_other_types(*node_type*, *keep_soma=True*)

Strips everything from the morphology except for the specified type. Parent and child relationships are updated accordingly, creating new roots when necessary.

Parameters

node_type: enum
The node type to keep in the morphology. Use one of the following constants: SOMA, AXON, BASAL_DENDRITE, or APICAL_DENDRITE

keep_soma: Boolean (optional)
True (default) if soma nodes should remain in the morpyhology, and False if the soma should also be stripped

strip_type(*node_type*)

Strips all nodes of the specified type from the morphology. Parent and child relationships are updated accordingly, creating new roots when necessary.

Parameters

node_type: enum
The node type to strip from the morphology. Use one of the following constants: SOMA, AXON, BASAL_DENDRITE, or APICAL_DENDRITE

`stumpify_axon`(*count=10*)

Remove all axon nodes except the first 'count' nodes, as counted from the connected axon root.

Parameters

count: Integer
The length of the axon 'stump', in number of nodes

`to_dict`()

Returns a dictionary of Node objects. These Nodes are a copy of the Morphology. Modifying them will not modify anything in the Morphology itself.

`tree`(*n*)

Returns a list of all Morphology nodes within the specified tree. A tree is defined as a fully connected graph of nodes. Each tree has exactly one root.

Parameters

n: integer
ID of desired tree

Returns

A list of all morphology objects in the specified tree, or None if the tree doesn't exist

`write`(*file_name*)

allensdk.internal.morphology.morphvis module

`class` allensdk.internal.morphology.morphvis.**MorphologyColors**

Bases: `object`

`set_apical_color`(*r*, *g*, *b*)

`set_axon_color`(*r*, *g*, *b*)

`set_basal_color`(*r*, *g*, *b*)

`set_soma_color`(*r*, *g*, *b*)

allensdk.internal.morphology.morphvis.**calculate_scale**(*morph*, *pix_width*, *pix_height*)

Calculates scaling factor and x,y insets required to auto-scale and center morphology into box with specified numbers of pixels

Parameters

morph: AISDK Morphology object
pix_width: int
Number of image pixels on X axis
pix_height: int
Number of image pixels on Y axis

Returns

real, real, real
First return value is the scaling factor. Second is the number of pixels needed to adjust x-coordinates so that the morphology is horizontally centered. Third is the number of pixels needed to adjust the y-coordinates so that the morphology is vertically centered.

allensdk.internal.morphology.morphvis.**create_image**(*w*, *h*, *color=None*, *alpha=False*)

allensdk.internal.morphology.morphvis.**draw_density_hist**(*img*, *morph*, *vert_scale*, *inset_left=0*, *inset_right=0*, *inset_top=0*, *inset_bottom=0*, *num_bins=None*, *colors=None*)

Draws density histogram onto image When no scaling is applied, and no insets are provided, the coordinates of the morphology are used directly – i.e., 100 in morphology coordinates is equal to 100 pixels.

The scale factor is multiplied to morphology coordinates before being drawn. If scale_factor=2 then 50 in morphology coordinates is 100 pixels. Left and top insets shift the coordinate axes for drawing. E.g., if left=10 and top=5 then 0,0 in morphology coordinates is 10,5 in pixel space. Bottom and right insets are ignored.

If scale_to_fit is set then scale factor is ignored. The morphology is scaled to be the maximum size that fits in the image, taking into account insets. In a 100x100 image, if all insets=10, then the image is scaled to fit into the center 80x80 pixel area, and nothing is drawn in the inset border areas.

Axons are drawn before soma and dendrite compartments.

Parameters

img: PIL image object
morph: AISDK Morphology object
vert_scale: real
This is the amout required to multiply to a moprhology y-coordinate to convert it to relative cortical depth (on [0,1]). This is the inverse of the cortical thickness.
inset_*: real
This is the number of pixels to use as border on top/bottom/ right/left. If scale_to_fit is false then only the top/left values are used, as the scale_factor will determine how large the morphology is (it can be drawn beyond insets and even beyond image boundaries)
num_bins: int
The number of bins in the histogram
colors: MorphologyColors object
This is the color scheme used to draw the morphology. If colors=None then default coloring is used

Returns

Histogram arrays: [hist, hist2, hist3, hist4]
where hist is the histgram of all neurites, and hist[234] are the histograms of SWC types 2,3,4

allensdk.internal.morphology.morphvis.**draw_morphology**(*img*, *morph*, *inset_left=0*, *inset_right=0*, *inset_top=0*, *inset_bottom=0*, *scale_to_fit=False*, *scale_factor=1.0*, *colors=None*)

Draws morphology onto image When no scaling is applied, and no insets are provided, the coordinates of the morphology are used directly – i.e., 100 in morphology coordinates is equal to 100 pixels.

The scale factor is multiplied to morphology coordinates before being drawn. If scale_factor=2 then 50 in morphology coordinates is 100 pixels. Left and top insets shift the coordinate axes for drawing. E.g., if left=10 and top=5 then 0,0 in morphology coordinates is 10,5 in pixel space. Bottom and right insets are ignored.

If scale_to_fit is set then scale factor is ignored. The morphology is scaled to be the maximum size that fits in the image, taking into account insets. In a 100x100 image, if all insets=10, then the image is scaled to fit into the center 80x80 pixel area, and nothing is drawn in the inset border areas.

Axons are drawn before soma and dendrite compartments.

Parameters

img: PIL image object
morph: AISDK Morphology object
inset_*: real
This is the number of pixels to use as border on top/bottom/ right/left. If scale_to_fit is false then only the top/left values are used, as the scale_factor will determine how large the morphology is (it can be drawn beyond insets and even beyond image boundaries)
scale_to_fit: boolean
If true then morphology is scaled to the inset area of the image and scale_factor is ignored. Morphology is centered in the image in the sense that the top/bottom and left/right edges of the morphology are equidistant from image borders.
scale_factor: real
A scalar amount that is multiplied to morphology coordinates before drawing
colors: MorphologyColors object
This is the color scheme used to draw the morphology. If colors=None then default coloring is used

Returns

2-dimensional array, the pixel coordinates of the soma root [x,y]

allensdk.internal.morphology.node module

class allensdk.internal.morphology.node.**Node**(*n*, *t*, *x*, *y*, *z*, *r*, *pn*, ***kwargs*)

Bases: object

Represents node in SWC morphology file

classmethod from_dict(*d*)

short_string()

create string with node information in succinct, single-line form

to_dict()

Convert the node into a serializable dictionary

allensdk.internal.morphology.node.**euclidean_distance**(*node1*, *node2*)

allensdk.internal.morphology.node.**midpoint**(*node1*, *node2*)

allensdk.internal.morphology.validate_swc module

class allensdk.internal.morphology.validate_swc.**TestNode**(*n*, *t*, *x*, *y*, *z*, *r*, *pn*)

Bases: object

allensdk.internal.morphology.validate_swc.**main**()

allensdk.internal.morphology.validate_swc.**resave_swc**(*orig_swc*, *new_file*)

Reads SWC file into AllenSDK Morphology object and resaves it. This can fix some problems in an SWC file that may disrupt other software tools reading the file (e.g., NEURON)

Parameters

orig_swc: string
Name of SWC file to read
new_file: string
Name of output SWC file

allensdk.internal.morphology.validate_swc.**validate_swc**(*swc_file*)

Tests SWC files for compatibility with AllenSDK

To be compatible with NEURON, SWC files must have the following properties:

1) a single root node with parent ID '-1'
2) sequentially increasing ID numbers
3) immediate children of the soma cannot branch

To be compatible with feature analysis, SWC files can only have node types in the range 1-4:

1 = soma 2 = axon 3 = [basal] dendrite 4 = apical dendrite

Module contents

allensdk.internal.mouse_connectivity package

Subpackages

allensdk.internal.mouse_connectivity.interval_unionize package

Submodules

allensdk.internal.mouse_connectivity.interval_unionize.cav_unionize module

allensdk.internal.mouse_connectivity.interval_unionize.cav_unionizer module

allensdk.internal.mouse_connectivity.interval_unionize.data_utilities module

allensdk.internal.mouse_connectivity.interval_unionize.data_utilities.**get_cav_density**(*cav_density_path*)

allensdk.internal.mouse_connectivity.interval_unionize.data_utilities.**get_injection_data**(*injection_fraction_path*, *injection_density_path*, *injection_energy_path*)

Read nrrd files containing injection signal data

allensdk.internal.mouse_connectivity.interval_unionize.data_utilities.**get_projection_data**(*projection_density_path*, *projection_energy_path*, *aav_exclusion_fr*

Read nrrd files containing global signal data

allensdk.internal.mouse_connectivity.interval_unionize.data_utilities.**get_sum_pixel_intensities**(*sum_pixels_path*, *injection_sum*

allensdk.internal.mouse_connectivity.interval_unionize.data_utilities.**get_sum_pixels**(*sum_pixels_path*)

allensdk.internal.mouse_connectivity.interval_unionize.data_utilities.**load_annotation**(*annotation_path*, *data_mask_path=None*)

Read data files segmenting the reference space into regions of valid and invalid data, then further among brain structures

allensdk.internal.mouse_connectivity.interval_unionize.data_utilities.**read**(*path*)

allensdk.internal.mouse_connectivity.interval_unionize.interval_unionizer module

class allensdk.internal.mouse_connectivity.interval_unionize.interval_unionizer.**IntervalUnionizer**(*exclude*)

Bases: `object`

direct_unionize(*data_arrays*, *pre_sorted=False*, ***kwargs*)

Obtain unionize records from directly annotated regions.

Parameters

data_arrays
[dict] Keys identify types of data volume. Values are flattened arrays.

sorted
[bool, optional] If False, data arrays will be sorted.

extract_data(*data_arrays*, *low*, *high*, ***kwargs*)

Given flattened data arrays and a specified interval, generate summary data

Parameters

data_arrays
[dict] Keys identify types of data volume. Values are flattened, sorted arrays.

low
[int] Index at which interval of interest begins. Inclusive.

high
[int] Index at which interval of interest ends. Exclusive.

`postprocess_unionizes`(*raw_unionizes*, ***kwargs*)

Carry out additional calculations/formatting derivative of core unionization.

Parameters

raw_unionizes
[list of unionizes] Each entry is a unionize record.

`classmethod propagate_record`(*child_record*, *ancestor_record*, *copy_all=False*)

Updates one unionize corresponding to a rootward structure with information from a unionize corresponding to a leafward structure

Parameters

child_record
[unionize] Data will be drawn from this record

ancestor_record
[unionize] This record will be updated

`classmethod propagate_to_bilateral`(*lateral_unionizes*)

`classmethod propagate_unionizes`(*direct_unionizes*, *ancestor_id_map*)

Structures are arranged in a tree, whose leafward-oriented edges indicate physical containment. This method updates rootward unionize records with information from leafward ones.

Parameters

direct_unionizes
[list of unionizes] Each entry is a unionize record produced from a collection of directly labeled voxels in the segmentation volume.

ancestor_id_map
[dict] Keys are structure ids. Values are ids of all structures rootward in

the tree, including the key node

Returns

output_unionizes
[list of unionizes] Contains completed unionize records at all depths in the structure tree

`classmethod record_cb`()

`setup_interval_map`(*annotation*)

Build a map from structure ids to intervals in the sorted flattened reference space.

Parameters

annotation
[np.ndarray] Segmentation label array.

`sort_data_arrays`(*data_arrays*)

Apply the precomputed sort to flattened data arrays

Parameters

data_arrays
[dict] Keys identify types of data volume. Values are flattened, unsorted arrays.

Returns

dict
As input, but values are sorted

allensdk.internal.mouse_connectivity.interval_unionize.run_tissuecyte_unionize_cav module

allensdk.internal.mouse_connectivity.interval_unionize.run_tissuecyte_unionize_classic module

allensdk.internal.mouse_connectivity.interval_unionize.run_tissuecyte_unionize_classic.**get_ancestor_id_**

allensdk.internal.mouse_connectivity.interval_unionize.run_tissuecyte_unionize_classic.**get_volume_scale**

allensdk.internal.mouse_connectivity.interval_unionize.run_tissuecyte_unionize_classic.**run**(*input_data*)

allensdk.internal.mouse_connectivity.interval_unionize.tissuecyte_unionize_record module

class allensdk.internal.mouse_connectivity.interval_unionize.tissuecyte_unionize_record.**TissuecyteBaseUnionize**

Bases: *Unionize*

direct_sum_projection_pixels

max_voxel_density

max_voxel_index

output(*output_spacing_iso*, *volume_scale*, *target_shape*, *sort*)

Generate derived data for this unionize

Parameters

output_spacing_iso
[numeric] Isometric spacing of reference space in microns

volume_scale
[numeric] Scale factor mapping pixels to microns^3

target_shape
[array-like of numeric] Shape of reference space

projection_density

projection_energy

projection_intensity

propagate(*ancestor*, *copy_all=False*)

Update a rootward unionize with data from this unionize record

Parameters

ancestor
[TissuecyteBaseUnionize] will be updated

Returns

ancestor
[TissuecyteBaseUnionize]

`set_max_voxel`(*density_array*, *low*)

Find the voxel of greatest density in this unionizes spatial domain

Parameters

density_array
[ndarray] Float values are densities per voxel

low
[int] index in full flattened, sorted array of starting voxel

`sum_pixel_intensity`

`sum_pixels`

`sum_projection_pixel_intensity`

`sum_projection_pixels`

class allensdk.internal.mouse_connectivity.interval_unionize.tissuecyte_unionize_record.**TissuecyteInjectionUnionize**

Bases: *TissuecyteBaseUnionize*

`calculate`(*low*, *high*, *data_arrays*)

`direct_sum_projection_pixels`

`max_voxel_density`

`max_voxel_index`

`projection_density`

`projection_energy`

`projection_intensity`

`sum_pixel_intensity`

`sum_pixels`

`sum_projection_pixel_intensity`

`sum_projection_pixels`

class allensdk.internal.mouse_connectivity.interval_unionize.tissuecyte_unionize_record.**TissuecyteProjectionUnionize**

Bases: *TissuecyteBaseUnionize*

`calculate`(*low*, *high*, *data_arrays*, *ij_record*)

`direct_sum_projection_pixels`

`max_voxel_density`

`max_voxel_index`

`projection_density`

projection_energy

projection_intensity

sum_pixel_intensity

sum_pixels

sum_projection_pixel_intensity

sum_projection_pixels

allensdk.internal.mouse_connectivity.interval_unionize.tissuecyte_unionizer module

class allensdk.internal.mouse_connectivity.interval_unionize.tissuecyte_unionizer.**TissuecyteUnionizer**(*ex*

Bases: *IntervalUnionizer*

A specialization of the IntervalUnionizer set up for unionizing Tissuecyte-derived projection data.

extract_data(*data_arrays*, *low*, *high*)

As parent

postprocess_unionizes(*raw_unionizes*, *image_series_id*, *output_spacing_iso*, *volume_scale*, *target_shape*, *sort*)

As parent

classmethod propagate_record(*child_record*, *ancestor_record*, *copy_all=False*)

As parent

classmethod record_cb()

allensdk.internal.mouse_connectivity.interval_unionize.unionize_record module

class allensdk.internal.mouse_connectivity.interval_unionize.unionize_record.**Unionize**(**args*, ***kwargs*)

Bases: object

Abstract base class for unionize records.

calculate(**args*, ***kwargs*)

output(**args*, ***kwargs*)

propagate(*ancestor*, *copy_all*, **args*, ***kwargs*)

slice_arrays(*low*, *high*, *data_arrays*)

Extract a slice from several aligned arrays

Parameters

low
[int] start of slice, inclusive

high
[int] end of slice, exclusive

data_arrays
[dict] keys are varieties of data. values are sorted, flattened data arrays

Module contents

allensdk.internal.mouse_connectivity.projection_thumbnail package

Submodules

allensdk.internal.mouse_connectivity.projection_thumbnail.generate_projection_strip module

allensdk.internal.mouse_connectivity.projection_thumbnail.generate_projection_strip.**apply_colormap**(*image*, *colormap*)

allensdk.internal.mouse_connectivity.projection_thumbnail.generate_projection_strip.**blend_with_background**

allensdk.internal.mouse_connectivity.projection_thumbnail.generate_projection_strip.**do_blur**(*image*, *blur*)

allensdk.internal.mouse_connectivity.projection_thumbnail.generate_projection_strip.**handle_output_image**

allensdk.internal.mouse_connectivity.projection_thumbnail.generate_projection_strip.**max_cb**(*max_sheet*, *depth_sheet*, *volume*, *axis*, **a*, ***k*)

allensdk.internal.mouse_connectivity.projection_thumbnail.generate_projection_strip.**run**(*volume*, *imin*, *imax*, *rotations*, *colormap*)

allensdk.internal.mouse_connectivity.projection_thumbnail.generate_projection_strip.**simple_rotation**(*from_a*, *to_a*, *start*, *end*, *nsteps*)

allensdk.internal.mouse_connectivity.projection_thumbnail.image_sheet module

class allensdk.internal.mouse_connectivity.projection_thumbnail.image_sheet.**ImageSheet**

Bases: object

append(*new_cell*)

apply(*fn*, **args*, ***kwargs*)

static build_from_image(*image*, *n*, *axis*)

copy()

get_output(*axis*)

allensdk.internal.mouse_connectivity.projection_thumbnail.projection_functions module

allensdk.internal.mouse_connectivity.projection_thumbnail.projection_functions.**convert_axis**(*axis*)

allensdk.internal.mouse_connectivity.projection_thumbnail.projection_functions.**max_projection**(*volume*, *axis*, **a*, ***k*)

allensdk.internal.mouse_connectivity.projection_thumbnail.projection_functions.**template_projection**(*volum*
axis,
gain=
maxv=
**a*,
***k*)

allensdk.internal.mouse_connectivity.projection_thumbnail.visualization_utilities module

allensdk.internal.mouse_connectivity.projection_thumbnail.visualization_utilities.**blend**(*image_stack*, *weight_stack*)

Parameters

image_stack :: list of np.ndarray
The images to be blended. Shapes cannot differ

weight_stack :: list of np.ndarray
The weight of each image at each pixel. Will be normalized.

allensdk.internal.mouse_connectivity.projection_thumbnail.visualization_utilities.**convert_discrete_colo**

Generates a matplotlib continuous colormap on [0, 1] from a discrete colormap at N evenly spaced points.

Parameters

data
[list of list] Sublists are [r, g, b].

Returns

matplotlib.colors.LinearSegmentedColormap
Gamma is 1. Output space is 3 X [0, 1]

allensdk.internal.mouse_connectivity.projection_thumbnail.visualization_utilities.**minmax_norm**(*data*)

allensdk.internal.mouse_connectivity.projection_thumbnail.visualization_utilities.**normalize_intensity**(*da*
in
in
ou
ou

allensdk.internal.mouse_connectivity.projection_thumbnail.visualization_utilities.**sitk_safe_ln**(*data*,
minimum=
10)

allensdk.internal.mouse_connectivity.projection_thumbnail.volume_projector module

class allensdk.internal.mouse_connectivity.projection_thumbnail.volume_projector.**VolumeProjector**(*view_vo*

Bases: object

build_rotation_transform(*from_axis*, *to_axis*, *angle*)

extract(*cb*, *volume=None*)

classmethod fixed_factory(*volume*, *size*)

rotate(*from_axis*, *to_axis*, *angle*)

rotate_and_extract(*from_axes*, *to_axes*, *angles*, *cb*)

classmethod safe_factory(*volume*)

allensdk.internal.mouse_connectivity.projection_thumbnail.volume_utilities module

allensdk.internal.mouse_connectivity.projection_thumbnail.volume_utilities.**sitk_get_center**(*volume*)

allensdk.internal.mouse_connectivity.projection_thumbnail.volume_utilities.**sitk_get_diagonal_length**(*volu*

allensdk.internal.mouse_connectivity.projection_thumbnail.volume_utilities.**sitk_get_image_parameters**(*vol*

allensdk.internal.mouse_connectivity.projection_thumbnail.volume_utilities.**sitk_get_size_parity**(*volume*)

allensdk.internal.mouse_connectivity.projection_thumbnail.volume_utilities.**sitk_paste_into_center**(*smalle*
larger)

Module contents

allensdk.internal.mouse_connectivity.tissuecyte_stitching package

Submodules

allensdk.internal.mouse_connectivity.tissuecyte_stitching.stitcher module

class allensdk.internal.mouse_connectivity.tissuecyte_stitching.stitcher.**Stitcher**(*image_dimensions*, *tiles*, *average_tiles*, *channels*)

Bases: object

run(*cb=<built-in function array>*)

stitch(*slice_image*, *stitched_indicator*, *tile*, *cb=<built-in function array>*)

allensdk.internal.mouse_connectivity.tissuecyte_stitching.stitcher.**blend_component_from_point**(*point*, *mesh*, *lg*)

Obtains a normalized component of the blend, which describes depth of overlap along a specified axis in a specified direction

allensdk.internal.mouse_connectivity.tissuecyte_stitching.stitcher.**get_blend**(*indicator_region*, *stup*, *cb=<built-in function array>*)

allensdk.internal.mouse_connectivity.tissuecyte_stitching.stitcher.**get_blend_component**(*indicator*, *lg*, *axis*, *meshes*)

allensdk.internal.mouse_connectivity.tissuecyte_stitching.stitcher.**get_indicator_bound_point**(*indicator*, *lg*, *axis*)

Finds the index of first change in a binary mask along a specified axis in a specified direction

allensdk.internal.mouse_connectivity.tissuecyte_stitching.stitcher.**get_overall_blend**(*indicator*, *meshes*)

allensdk.internal.mouse_connectivity.tissuecyte_stitching.stitcher.**initialize_image**(*dimensions*, *nchannels*, *dtype*, *order='C'*)

allensdk.internal.mouse_connectivity.tissuecyte_stitching.stitcher.**initialize_images**(*dimensions*, *nchannels*)

allensdk.internal.mouse_connectivity.tissuecyte_stitching.stitcher.**make_blended_tile**(*blend*, *tile*, *current_region*)

allensdk.internal.mouse_connectivity.tissuecyte_stitching.tile module

class allensdk.internal.mouse_connectivity.tissuecyte_stitching.tile.**Tile**(*index*, *image*, *is_missing*, *bounds*, *channel*, *size*, *margins*, **args*, ***kwargs*)

Bases: object

apply_average_tile(*average_tile*)

apply_average_tile_to_self(*average_tile*)

average_tile_is_untrimmed(*average_tile*)

get_image_region()

get_missing_path()

initialize_image()

trim(*image*)

trim_self()

Module contents

Module contents

allensdk.internal.notebooks package

Submodules

allensdk.internal.notebooks.execute_notebooks module

Module contents

allensdk.internal.pipeline_modules package

Subpackages

allensdk.internal.pipeline_modules.gbm package

Submodules

allensdk.internal.pipeline_modules.gbm.generate_gbm_analysis_run_records module

allensdk.internal.pipeline_modules.gbm.generate_gbm_analysis_run_records.**main**(*analysis_records_json_location*, *db_host*, *db_port*, *db_name*, *db_user*, *db_passwd*)

allensdk.internal.pipeline_modules.gbm.generate_gbm_heatmap module

allensdk.internal.pipeline_modules.gbm.generate_gbm_heatmap.**create_gene_fpkm_table**(*analysis_run_records*)

Creates a a matrix ("rows x columns = genes x samples") of fpkm gene expression values for each particular (gene, sample) pair. Rows are sorted by entrez_id and columns are by rna_well_id

allensdk.internal.pipeline_modules.gbm.generate_gbm_heatmap.**create_genes_for_transcripts**(*analysis_run_trans*

Creates a list that contains the associated gene for each transcript sorted alphabetically

allensdk.internal.pipeline_modules.gbm.generate_gbm_heatmap.**create_sample_metadata**(*sample_metadata_records*

Creates a table of sample metadata sorted by rna_well_id

allensdk.internal.pipeline_modules.gbm.generate_gbm_heatmap.**create_transcript_fpkm_table**(*analysis_run_reco*

Creates a a matrix ("rows x columns = transcripts x samples") of fpkm gene expression values for each particular (transcript, sample) pair. Rows are sorted by transcript id and columns are by rna_well_id

allensdk.internal.pipeline_modules.gbm.generate_gbm_heatmap.**create_transcripts_for_genes**(*analysis_run_gene*

Creates a list that contains the associated transcript for each gene sorted by entrez_id

allensdk.internal.pipeline_modules.gbm.generate_gbm_heatmap.**main**()

allensdk.internal.pipeline_modules.gbm.generate_gbm_sample_metadata module

allensdk.internal.pipeline_modules.gbm.generate_gbm_sample_metadata.**main**(*sample_metadata_json_location*, *db_host*, *db_port*, *db_name*, *db_user*, *db_passwd*)

Module contents

Submodules

allensdk.internal.pipeline_modules.run_annotated_region_metrics module

Run annotated region metrics calculations

allensdk.internal.pipeline_modules.run_annotated_region_metrics.**debug**(*region_id*, *storage_directory='./'*, *local=True*, *sdk_path='/data/informatics/CAM/isi_metr*
script_path='/data/informatics/CAM/isi_me
lims_host='lims2')

allensdk.internal.pipeline_modules.run_annotated_region_metrics.**load_arrays**(*h5_file*)

allensdk.internal.pipeline_modules.run_annotated_region_metrics.**main**()

allensdk.internal.pipeline_modules.run_demixing module

allensdk.internal.pipeline_modules.run_demixing.**assert_exists**(*file_name*)

allensdk.internal.pipeline_modules.run_demixing.**debug**(*experiment_id*, *local=False*)

allensdk.internal.pipeline_modules.run_demixing.**get_path**(*obj*, *key*, *check_exists*)

allensdk.internal.pipeline_modules.run_demixing.**main**()

allensdk.internal.pipeline_modules.run_demixing.**parse_input**(*data*, *exclude_labels*)

allensdk.internal.pipeline_modules.run_dff_computation module

allensdk.internal.pipeline_modules.run_dff_computation.**main**()

allensdk.internal.pipeline_modules.run_dff_computation.**parse_input**(*data*)

allensdk.internal.pipeline_modules.run_eye_tracking module

allensdk.internal.pipeline_modules.run_neuropil_correction module

allensdk.internal.pipeline_modules.run_neuropil_correction.**adjust_r_for_negativity**(*r*, *F_C*, *F_M*, *F_N*)

allensdk.internal.pipeline_modules.run_neuropil_correction.**debug**(*experiment_id*, *local=False*)

allensdk.internal.pipeline_modules.run_neuropil_correction.**debug_plot**(*file_name*, *roi_trace*, *neuropil_trace*, *corrected_trace*, *r*, *r_vals=None*, *err_vals=None*)

allensdk.internal.pipeline_modules.run_neuropil_correction.**main**()

allensdk.internal.pipeline_modules.run_observatory_analysis module

allensdk.internal.pipeline_modules.run_observatory_analysis.**debug**(*experiment_ids*, *local=False*, *OUTPUT_DIR='/data/informatics/CAM/analysis/'*, *SDK_PATH='/data/informatics/CAM/analysis/al*
walltime='10:00:00', *python='/shared/utils.x86_64/python-2.7/bin/python'*, *queue='braintv'*)

allensdk.internal.pipeline_modules.run_observatory_analysis.**get_experiment_nwb_file**(*experiment_id*)

allensdk.internal.pipeline_modules.run_observatory_analysis.**get_experiment_session**(*experiment_id*)

allensdk.internal.pipeline_modules.run_observatory_analysis.**main**()

allensdk.internal.pipeline_modules.run_observatory_container_thumbnails module

allensdk.internal.pipeline_modules.run_observatory_thumbnails module

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_cell_plots**(*cell_specimen_ids*, *prefix*, *aspect*, *configs*, *output_dir*, *axes=None*, *transparent=False*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_correlation_plots**(*data_set*, *analysis_file*, *configs*, *output_dir*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_drifting_gratings**(*dga*, *configs*, *output_dir*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_experiment_thumbnails**(*nwb_file*, *analysis_file*, *output_directory*, *types=None*, *threads=4*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_eye_tracking_plots**(*data_set*, *configs*, *output_dir*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_locally_sparse_noise**(*lsna*, *configs*, *output_dir*, *on*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_natural_movie**(*nma*, *configs*, *output_dir*, *name*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_natural_scenes**(*nsa*, *configs*, *output_dir*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_plots**(*prefix*, *aspect*, *configs*, *output_dir*, *axes=None*, *transparent=False*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_receptive_field**(*lsna*, *configs*, *output_dir*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_speed_tuning**(*analysis*, *configs*, *output_dir*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_static_gratings**(*sga*, *configs*, *output_dir*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**build_type**(*nwb_file*, *data_file*, *configs*, *output_dir*, *type_name*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**debug**(*experiment_id*, *plots=None*, *local=False*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**get_experiment_analysis_file**(*experiment_id*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**get_experiment_files**(*experiment_id*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**get_experiment_nwb_file**(*experiment_id*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**get_input_data**(*experiment_id*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**lsna_check_hvas**(*data_set*, *data_file*)

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**main**()

allensdk.internal.pipeline_modules.run_observatory_thumbnails.**parse_input**(*data*)

allensdk.internal.pipeline_modules.run_ophys_eye_calibration module

allensdk.internal.pipeline_modules.run_ophys_eye_calibration.**debug**(*experiment_id*, *local=False*)

allensdk.internal.pipeline_modules.run_ophys_eye_calibration.**get_wkf**(*wkf_type*, *experiment_id*)

allensdk.internal.pipeline_modules.run_ophys_eye_calibration.**main**()

allensdk.internal.pipeline_modules.run_ophys_eye_calibration.**parse_input_data**(*data*)

allensdk.internal.pipeline_modules.run_ophys_eye_calibration.**write_output**(*filename*, *position_degrees*, *position_cm*, *areas*)

allensdk.internal.pipeline_modules.run_ophys_session_decomposition module

allensdk.internal.pipeline_modules.run_ophys_session_decomposition.**convert_frame**(*conversion_definition*)

allensdk.internal.pipeline_modules.run_ophys_session_decomposition.**create_fake_metadata**(*exp_dir*, *raw_path*, *channels=None*, *width=512*, *height=256*, *itemsize=2*, *n_planes=6*)

allensdk.internal.pipeline_modules.run_ophys_session_decomposition.**debug**(*experiment_id*, *local=False*, *raw_path=None*)

allensdk.internal.pipeline_modules.run_ophys_session_decomposition.**main**()

allensdk.internal.pipeline_modules.run_ophys_session_decomposition.**parse_input**(*data*)

Load all input data from the input json.

allensdk.internal.pipeline_modules.run_ophys_time_sync module

class allensdk.internal.pipeline_modules.run_ophys_time_sync.**TimeSyncOutputs**(*experiment_id: int*, *stimulus_delay: float*, *ophys_delta: int*, *stimulus_delta: int*, *eye_delta: int*, *behavior_delta: int*, *ophys_times: ndarray*, *stimulus_times: ndarray*, *eye_times: ndarray*, *behavior_times: ndarray*, *stimulus_alignment: ndarray*, *eye_alignment: ndarray*, *behavior_alignment: ndarray*)

Bases: tuple

Schema for synchronization outputs

behavior_alignment: ndarray

Alias for field number 12

behavior_delta: int

Alias for field number 5

behavior_times: ndarray

Alias for field number 9

experiment_id: int

Alias for field number 0

eye_alignment: ndarray

Alias for field number 11

eye_delta: int

Alias for field number 4

eye_times: ndarray

Alias for field number 8

ophys_delta: int

Alias for field number 2

ophys_times: ndarray

Alias for field number 6

stimulus_alignment: ndarray

Alias for field number 10

stimulus_delay: float

Alias for field number 1

stimulus_delta: int

Alias for field number 3

stimulus_times: ndarray

Alias for field number 7

class allensdk.internal.pipeline_modules.run_ophys_time_sync.**TimeSyncWriter**(*output_h5_path: str*, *output_json_path: str | None = None*)

Bases: object

validate_paths()

Determines whether we can actually write to the specified paths, allowing for creation of intermediate directories. It is a good idea to run this beore doing any heavy calculations!

write(*outputs:* TimeSyncOutputs)

Convenience for writing both an output h5 and (if applicable) an output json.

Parameters

outputs
[the data to be written]

write_output_h5(*outputs*)

Write (mainly) heaviweight data to an h5 file.

Parameters

outputs
[the data to be written]

write_output_json(*outputs*)

Write lightweight data to a json

Parameters

outputs
[the data to be written]

allensdk.internal.pipeline_modules.run_ophys_time_sync.**check_stimulus_delay**(*obt_delay: float*, *min_delay: float*, *max_delay: float*)

Raise an exception if the monitor delay is not within specified bounds

Parameters

obt_delay
[obtained monitor delay (s)]

min_delay
[lower threshold (s)]

max_delay
[upper threshold (s)]

allensdk.internal.pipeline_modules.run_ophys_time_sync.**main**()

allensdk.internal.pipeline_modules.run_ophys_time_sync.**run_ophys_time_sync**(*aligner:* OphysTimeAligner, *experiment_id: int*, *min_stimulus_delay: float*, *max_stimulus_delay: float*) → *TimeSyncOutputs*

Carry out synchronization of timestamps across the data streams of an ophys experiment.

Parameters

aligner
[drives alignment. See OphysTimeAligner for details of the] attributes and properties that must be implemented.

experiment_id
[unique identifier for the experiment being aligned]

min_stimulus_delay
[reject alignment run (raise a ValueError) if the] calculated monitor delay is below this value (s).

max_stimulus_delay
[reject alignment run (raise a ValueError) if the] calculated monitor delay is above this value (s).

Returns

A TimeSyncOutputs (see definintion for more information) of output
parameters and arrays of aligned timestamps.

allensdk.internal.pipeline_modules.run_roi_filter module

allensdk.internal.pipeline_modules.run_tissuecyte_projection_thumbnail_from_json module

allensdk.internal.pipeline_modules.run_tissuecyte_stitching_classic module

allensdk.internal.pipeline_modules.run_tissuecyte_unionize_cav_from_json module

allensdk.internal.pipeline_modules.run_tissuecyte_unionize_classic_counts_from_json module

allensdk.internal.pipeline_modules.run_tissuecyte_unionize_classic_from_json module

allensdk.internal.pipeline_modules.run_tissuecyte_unionize_classic_from_json.**main**()

Module contents

Module contents

6.1.7 allensdk.model package

Subpackages

allensdk.model.biophys_sim package

Subpackages

allensdk.model.biophys_sim.neuron package

Submodules

allensdk.model.biophys_sim.neuron.hoc_utils module

class allensdk.model.biophys_sim.neuron.hoc_utils.**HocUtils**(*description*)

Bases: object

A helper class for containing references to NEUORN.

Attributes

h
[object] The NEURON hoc object.

nrn
[object] The NEURON python object.

neuron
[module] The NEURON module.

h = None

initialize_hoc()
Basic setup for NEURON.

neuron = None

nrn = None

Module contents

allensdk.model.biophys_sim.scripts package

Module contents

Submodules

allensdk.model.biophys_sim.bps_command module

allensdk.model.biophys_sim.bps_command.**choose_bps_command**(*command='bps_simple'*, *conf_file=None*)

allensdk.model.biophys_sim.bps_command.**run_module**(*description*, *module_name*, *function_name*)

allensdk.model.biophys_sim.config module

class allensdk.model.biophys_sim.config.**Config**

Bases: *ApplicationConfig*

load(*config_path*, *disable_existing_logs=False*)

Parse the application configuration then immediately load the model configuration files.

Parameters

disable_existing_logs
[boolean, optional] If false (default) leave existing logs after configuration.

read_model_description()

parse the model_file field of the application configuration and read the files.

The model_file field of the application configuration is first split at commas, since it may list more than one file.

The files may be uris of the form `file:filename?section=name`, in which case a bare configuration object is read from filename into the configuration section with key 'name'.

A simple filename without a section option is treated as a standard multi-section configuration file.

Returns

description
[Description] Configuration object.

Module contents

allensdk.model.biophysical package

Submodules

allensdk.model.biophysical.run_simulate module

class allensdk.model.biophysical.run_simulate.**RunSimulate**(*input_json*, *output_json*)

Bases: `object`

load_manifest()

nrnivmodl()

simulate()

allensdk.model.biophysical.run_simulate.**main**(*command*, *lims_strategy_json*, *lims_response_json*)

Entry point for module. :param command: select behavior, nrnivmodl or simulate :type command: string :param lims_strategy_json: path to json file output from lims. :type lims_strategy_json: string :param lims_response_json: path to json file returned to lims. :type lims_response_json: string

allensdk.model.biophysical.runner module

allensdk.model.biophysical.runner.**load_description**(*args_dict*)

Read configurations.

Parameters

args_dict
[dict] Parsed arguments dictionary with following keys.

manifest_file
[string] .json file with containing the experiment configuration

axon_type
[string] Axon handling for the all-active models

Returns

Config
Object with all information needed to run the experiment.

allensdk.model.biophysical.runner.**prepare_nwb_output**(*nwb_stimulus_path*, *nwb_result_path*)

Copy the stimulus file, zero out the recorded voltages and spike times.

Parameters

nwb_stimulus_path
[string] NWB file name

nwb_result_path
[string] NWB file name

allensdk.model.biophysical.runner.**run**(*args*, *sweeps=None*, *procs=6*)

Main function for simulating sweeps in a biophysical experiment.

Parameters

args
[dict] Parsed arguments to run the experiment.

procs
[int] number of sweeps to simulate simultaneously.

sweeps
[list] list of experiment sweep numbers to simulate. If None, simulate all sweeps.

allensdk.model.biophysical.runner.**run_sync**(*description*, *sweeps=None*)

Single-process main function for simulating sweeps in a biophysical experiment.

Parameters

description
[Config] All information needed to run the experiment.

sweeps
[list] list of experiment sweep numbers to simulate. If None, simulate all sweeps.

allensdk.model.biophysical.runner.**save_nwb**(*output_path*, *v*, *sweep*, *sweeps_by_type*)

Save a single voltage output result into an existing sweep in a NWB file. This is intended to overwrite a recorded trace with a simulated voltage.

Parameters

output_path
[string] file name of a pre-existing NWB file.

v
[numpy array] voltage

sweep
[integer] which entry to overwrite in the file.

allensdk.model.biophysical.utils module

class allensdk.model.biophysical.utils.**AllActiveUtils**(*description*, *axon_type*)

Bases: *Utils*

generate_morphology(*morph_filename*)

Load a neurolucida or swc-format cell morphology file.

Parameters

morph_filename
[string] Path to morphology.

load_cell_parameters()

Configure a neuron after the cell morphology has been loaded.

class allensdk.model.biophysical.utils.**Utils**(*description*)

Bases: *HocUtils*

A helper class for NEURON functionality needed for biophysical simulations.

Attributes

h
[object] The NEURON hoc object.

nrn
[object] The NEURON python object.

neuron
[module] The NEURON module.

generate_morphology(*morph_filename*)

Load a swc-format cell morphology file.

Parameters

morph_filename
[string] Path to swc.

get_recorded_data(*vec*)

Extract recorded voltages and timestamps given the recorded Vector instance. If self.stimulus_sampling_rate is smaller than self.simulation_sampling_rate, resample to self.stimulus_sampling_rate.

Parameters

vec
[neuron.Vector] constructed by self.record_values

Returns

dict with two keys: 'v' = numpy.ndarray with voltages,
't' = numpy.ndarray with timestamps

`load_cell_parameters`()

Configure a neuron after the cell morphology has been loaded.

`static nearest_neuron_sampling_rate`(*hz*, *target_hz=40000*)

`read_stimulus`(*stimulus_path*, *sweep=0*)

Load current values for a specific experiment sweep and setup simulation and stimulus sampling rates.

NOTE: NEURON only allows simulation timestamps of multiples of 40KHz. To avoid aliasing, we set the simulation sampling rate to the least common multiple of the stimulus sampling rate and 40KHz.

Parameters

stimulus path
[string] NWB file name

sweep
[integer, optional] sweep index

`record_values`()

Set up output voltage recording.

`setup_iclamp`(*stimulus_path*, *sweep=0*)

Assign a current waveform as input stimulus.

Parameters

stimulus_path
[string] NWB file name

`update_default_cell_hoc`(*description*, *default_cell_hoc='cell.hoc'*)

replace the default 'cell.hoc' path in the manifest with 'cell.hoc' packaged within AllenSDK if it does not exist

`allensdk.model.biophysical.utils.create_utils`(*description*, *model_type=None*)

Factory method to create a Utils subclass.

Parameters

description
[Config instance] used to initialize Utils subclass

model_type
[string] Must be one of [PERISOMATIC_TYPE, ALL_ACTIVE_TYPE]. If none, defaults to PERISOMATIC_TYPE

Returns

Utils instance

Module contents

allensdk.model.glif package

Submodules

allensdk.model.glif.glif_neuron module

exception allensdk.model.glif.glif_neuron.**GlifBadResetException**(*message*, *dv*)

Bases: Exception

Exception raised when voltage is still above threshold after a reset rule is applied.

class allensdk.model.glif.glif_neuron.**GlifNeuron**(*El*, *dt*, *asc_tau_array*, *R_input*, *C*, *asc_amp_array*, *spike_cut_length*, *th_inf*, *th_adapt*, *coeffs*, *AScurrent_dynamics_method*, *voltage_dynamics_method*, *threshold_dynamics_method*, *AScurrent_reset_method*, *voltage_reset_method*, *threshold_reset_method*, *init_voltage*, *init_threshold*, *init_AScurrents*, ***kwargs*)

Bases: object

Implements the current-based Mihalas Neiber GLIF neuron. Simulations model the voltage, threshold, and afterspike currents of a neuron given an input stimulus. A set of modular dynamics rules are applied until voltage crosses threshold, at which point a set of modular reset rules are applied. See glif_neuron_methods.py for a list of what options there are for voltage, threshold, and afterspike current dynamics and reset rules.

Parameters

El
[float] resting potential

dt
[float] duration between time steps

asc_tau_array: np.ndarray
TODO

R_input
[float] input resistance

C
[float] capacitance

asc_amp_arrap
[np.ndarray] afterspike current vector. one element per element of asc_tau_array.

spike_cut_length
[int] how many time steps to replace with NaNs when a spike occurs.

th_inf
[float] instantaneous threshold

coeffs
[dict] dictionary coefficients premultiplied to neuron properties during simulation. used for optimization.

AScurrent_dynamics_method
[dict] dictionary containing the 'name' of the afterspike current dynamics method to use and a 'params' dictionary parameters to pass to that function.

voltage_dynamics_method
[dict] dictionary containing the 'name' of the voltage dynamics method to use and a 'params' dictionary parameters to pass to that function.

threshold_dynamics_method
[dict] dictionary containing the 'name' of the threshold dynamics method to use and a 'params' dictionary parameters to pass to that function.

AScurrent_reset_method
[dict] dictionary containing the 'name' of the afterspike current dynamics method to use and a 'params' dictionary parameters to pass to that function.

voltage_reset_method
[dict] dictionary containing the 'name' of the voltage dynamics method to use and a 'params' dictionary parameters to pass to that function.

threshold_reset_method
[dict] dictionary containing the 'name' of the threshold dynamics method to use and a 'params' dictionary parameters to pass to that function.

init_voltage
[float] initial voltage value

init_threshold
[float] initial spike threshold value

init_AScurrents
[np.ndarray] initial afterspike current vector. one element per element of asc_tau_array.

`TYPE = 'GLIF'`

`append_threshold_components`(*spike*, *voltage*)

`static configure_library_method`(*method_type*, *params*)

Create a GlifNeuronMethod instance out of a library of functions organized by type name. This refers to the METHOD_LIBRARY in glif_neuron_methods.py, which lays out the available functions that can be used for dynamics and reset rules.

Parameters

method_type
[string] the name of a function category (e.g. 'AScurrent_dynamics_method' for the afterspike current dynamics methods)

params
[dict] a dictionary with two members. 'name': the string name of function you want, and 'params': parameters you want to pass to that function

Returns

GlifNeuronMethod
a GlifNeuronMethod instance

`static configure_method`(*method_name*, *method*, *method_params*)

Create a GlifNeuronMethod instance given a name, a function, and function parameters. This is just a shortcut to the GlifNeuronMethod constructor.

Parameters

method_name
[string] name for refering to this method later

method
[function] a python function

method_parameters
[dict] function arguments whose values should be fixed

Returns

GlifNeuronMethod
a GlifNeuronMethod instance

`dynamics`(*voltage_t0*, *threshold_t0*, *AScurrents_t0*, *inj*, *time_step*, *spike_time_steps*)

Update the voltage, threshold, and afterspike currents of the neuron for a single time step.

Parameters

voltage_t0
[float] the current voltage of the neuron

threshold_t0
[float] the current spike threshold level of the neuron

AScurrents_t0
[np.ndarray] the current state of the afterspike currents in the neuron

inj
[float] the current value of the current injection into the neuron

time_step
[int] the current time step of the neuron simulation

spike_time_steps
[list] a list of all of the time steps of spikes in the neuron

Returns

tuple
voltage_t1 (voltage at next time step), threshold_t1 (threshold at next time step), AScurrents_t1 (afterspike currents at next time step)

`classmethod from_dict`(*d*)

`reset`(*voltage_t0*, *threshold_t0*, *AScurrents_t0*)

Apply reset rules to the neuron's voltage, threshold, and afterspike currents assuming a spike has occurred (voltage is above threshold).

Parameters

voltage_t0
[float] the current voltage of the neuron

threshold_t0
[float] the current spike threshold level of the neuron

AScurrents_t0
[np.ndarray] the current state of the afterspike currents in the neuron

Returns

tuple
voltage_t1 (voltage at next time step), threshold_t1 (threshold at next time step), AScurrents_t1 (afterspike currents at next time step)

run(*stim*)

Run neuron simulation over a given stimulus. This steps through the stimulus applying dynamics equations. After each step it checks if voltage is above threshold. If so, self.spike_cut_length NaNs are inserted into the output voltages, reset rules are applied to the voltage, threshold, and afterspike currents, and the simulation resumes.

Parameters

stim
[np.ndarray] vector of scalar current values

Returns

dict

a dictionary containing:
'voltage': simulated voltage values, 'threshold': threshold values during the simulation, 'AScurrents': afterspike current values during the simulation, 'grid_spike_times': spike times (in uits of self.dt) aligned to simulation time steps, 'interpolated_spike_times': spike times (in units of self.dt) linearly interpolated between time steps, 'spike_time_steps': the indices of grid spike times, 'interpolated_spike_voltage': voltage of the simulation at interpolated spike times, 'interpolated_spike_threshold': threshold of the simulation at interpolated spike times

property tau_m

to_dict()

Convert the neuron to a serializable dictionary.

allensdk.model.glif.glif_neuron.**interpolate_spike_time**(*dt*, *time_step*, *threshold_t0*, *threshold_t1*, *voltage_t0*, *voltage_t1*)

Given two voltage and threshold values, the dt between them and the initial time step, interpolate a spike time within the dt interval by intersecting the two lines.

allensdk.model.glif.glif_neuron.**interpolate_spike_value**(*dt*, *interpolated_spike_time_offset*, *v0*, *v1*)

Take a value at two adjacent time steps and linearly interpolate what the value would be at an offset between the two time steps.

allensdk.model.glif.glif_neuron.**line_crossing_x**(*dx*, *a0*, *a1*, *b0*, *b1*)

Find the x value of the intersection of two lines.

allensdk.model.glif.glif_neuron.**line_crossing_y**(*dx*, *a0*, *a1*, *b0*, *b1*)

Find the y value of the intersection of two lines.

allensdk.model.glif.glif_neuron_methods module

The methods in this module are used for configuring dynamics and reset rules for the GlifNeuron. For more details on how to use these methods, see *Generalized LIF Models*.

class allensdk.model.glif.glif_neuron_methods.**GlifNeuronMethod**(*method_name*, *method*, *method_params*)

Bases: object

A simple class to keep track of the name and parameters associated with a neuron method. This class is initialized with a name, function, and parameters to pass to the function. The function then has those passed parameters fixed to a partial function using functools.partial. This class then mimics a function itself using the __call__ convention. Parameters that are not fixed in this way are assumed to be passed into the method when it is called. If the passed parameters contain an argument that is not part of the function signature, an exception will be raised.

Parameters

method_name
[string] A shorthand name that will be used to reference this method in the *GlifNeuron*.

method
[function] A python function to be called when this instance is called.

method_params
[dict] A dictionary mapping function arguments to values for values that should be fixed.

`modify_parameter`(*param*, *operator*)

Modify a function parameter needs to be modified after initialization.

Parameters

param
[string] the name of the parameter to modify

operator
[callable] a function or lambda that returns the desired modified value

Returns

type
the new value of the variable that was just modified.

`to_dict`()

allensdk.model.glif.glif_neuron_methods.**dynamics_AScurrent_exp**(*neuron*, *AScurrents_t0*, *time_step*, *spike_time_steps*)

Exponential afterspike current dynamics method takes a current at t0 and returns the current at a time step later.

allensdk.model.glif.glif_neuron_methods.**dynamics_AScurrent_none**(*neuron*, *AScurrents_t0*, *time_step*, *spike_time_steps*)

This method always returns zeros for the afterspike currents, regardless of input.

allensdk.model.glif.glif_neuron_methods.**dynamics_threshold_inf**(*neuron*, *threshold_t0*, *voltage_t0*, *AScurrents_t0*, *inj*)

Set threshold to the neuron's instantaneous threshold.

Parameters

neuron
[class]

threshold_t0
[not used here]

voltage_t0
[not used here]

AScurrents_t0
[not used here]

inj
[not used here]

AScurrents_t0
[not used here]

inj
[not used here]

allensdk.model.glif.glif_neuron_methods.**dynamics_threshold_spike_component**(*neuron*, *threshold_t0*, *voltage_t0*, *AScurrents_t0*, *inj*, *a_spike*, *b_spike*, *a_voltage*, *b_voltage*)

Analytical solution for spike component of threshold. The threshold will adapt via a component initiated by a spike which decays as an exponential. The component is in reference to threshold infinity and are recorded in the neuron's threshold components. The voltage component of the threshold is set to zero in the threshold components because it is zero here The third component refers to th_inf which is added separately as opposed to being included in the voltage component of the threshold as is done in equation 2.1 of Mihalas and Nieber 2009. Threshold infinity is removed for simple optimization.

Parameters

neuron
[class]

threshold_t0
[float] threshold input to function

voltage_t0
[float] voltage input to function

AScurrents_t0
[vector] values of after spike currents

inj
[float] current injected into the neuron

allensdk.model.glif.glif_neuron_methods.**dynamics_threshold_three_components_exact**(*neuron*, *threshold_t0*, *voltage_t0*, *AScurrents_t0*, *inj*, *a_spike*, *b_spike*, *a_voltage*, *b_voltage*)

Analytical solution for threshold dynamics. The threshold will adapt via two mechanisms: 1. a voltage dependent adaptation. 2. a component initiated by a spike which decays as an exponential. These two component are in reference to threshold infinity and are recorded in the neuron's threshold components. The third component refers to th_inf which is added separately as opposed to being included in the voltage component of the threshold as is done in equation 2.1 of Mihalas and Nieber 2009. Threshold infinity is removed for simple optimization.

Parameters

neuron
[class]

threshold_t0
[float] threshold input to function

voltage_t0
[float] voltage input to function

AScurrents_t0
[vector] values of after spike currents

inj
[float] current injected into the neuron

allensdk.model.glif.glif_neuron_methods.**dynamics_voltage_linear_exact**(*neuron*, *voltage_t0*, *AScurrents_t0*, *inj*)

(TODO) Linear voltage dynamics.

allensdk.model.glif.glif_neuron_methods.**dynamics_voltage_linear_forward_euler**(*neuron*, *voltage_t0*, *AScurrents_t0*, *inj*)

(TODO) Linear voltage dynamics.

allensdk.model.glif.glif_neuron_methods.**max_of_line_and_const**(*x*, *b*, *c*, *d*)

Find the maximum of a value and a position on a line

Parameters

x: float
x position on line 1

c: float
slope of line 1

d: float
y-intercept of line 1

b: float
y-intercept of line 2

Returns

float
the max of a line value and a constant

allensdk.model.glif.glif_neuron_methods.**min_of_line_and_zero**(*x*, *c*, *d*)

Find the minimum of a value and a position on a line

Parameters

x: float
x position on line 1

c: float
slope of line 1

d: float
y-intercept of line 1

b: float
y-intercept of line 2

Returns

float
the max of a line value and a constant

allensdk.model.glif.glif_neuron_methods.**reset_AScurrent_none**(*neuron*, *AScurrents_t0*)

Reset afterspike currents to zero.

allensdk.model.glif.glif_neuron_methods.**reset_AScurrent_sum**(*neuron*, *AScurrents_t0*, *r*)

Reset afterspike currents by adding summed exponentials. Left over currents from last spikes as well as newly initiated currents from current spike. Currents amplitudes in neuron.asc_amp_array need to be the amplitudes advanced though the spike cutting. I.e. In the preprocessor if the after spike currents are calculated via the GLM from spike initiation the amplitude at the time after the spike cutting needs to be calculated and neuron.asc_amp_array needs to be set to this value.

Parameters

r
[np.ndarray] a coefficient vector applied to the afterspike currents

allensdk.model.glif.glif_neuron_methods.**reset_threshold_inf**(*neuron*, *threshold_t0*, *voltage_v1*)

Reset the threshold to instantaneous threshold.

allensdk.model.glif.glif_neuron_methods.**reset_threshold_three_components**(*neuron*, *threshold_t0*, *voltage_v1*, *a_spike*, *b_spike*)

This method calculates the two components of the threshold: a spike (fast) component and a voltage (slow) component. The threshold_components vectors are then updated so that the traces match the voltage, current, and total threshold traces. The spike component of the threshold decays via an exponential fit specified by the amplitude a_spike and the time constant b_spike fit via the multiblip data. The voltage component does not change during the duration of the spike. The spike component are threshold component are summed along with threshold infinity to return the total threshold. Note that in the current implementation a_spike is added to the last value of the threshold_components which means that a_spike is the amplitude after spike cutting (if there is any).

Inputs:

neuron: class
contains attributes of the neuron

threshold_t0, voltage_t0: float
are not used but are here for consistency with other methods

a_spike: float
amplitude of the exponential decay of spike component of threshold after spike cutting has been implemented.

b_spike: float
amplitude of the exponential decay of spike component of threshold

Outputs:

Returns: float
the total threshold which is the sum of the spike component of threshold, the voltage component of threshold and threshold infinity (with it's corresponding coefficient)

neuron.threshold_components: dictionary containing

a spike: list
vector of spiking component of threshold that corresponds to the voltage, current, and total threshold traces

b_spike: list

vector of voltage component of threshold that corresponds to the voltage, current,
and total threshold traces.

Note that this function can be changed to use a_spike at the time of the spike and then have the the spike component plus the residual decay thought the spike. There are benefits and drawbacks to this. This potential change would be beneficial as it perhaps makes more biological sense for the threshold to go up at the time of spike if the traces are ever used. Also this would mean that a_spike would not have to be adjusted thought the spike cutting after the multiblip fit. However the current implementation makes sense in that it is similar to how afterspike currents are implemented.

allensdk.model.glif.glif_neuron_methods.**reset_voltage_v_before**(*neuron*, *voltage_t0*, *a*, *b*)

Reset voltage to the previous value with a scale and offset applied.

Parameters

a
[float] voltage scale constant

b
[float] voltage offset constant

allensdk.model.glif.glif_neuron_methods.**reset_voltage_zero**(*neuron*, *voltage_t0*)

Reset voltage to zero.

allensdk.model.glif.glif_neuron_methods.**spike_component_of_threshold_exact**(*th0*, *b_spike*, *t*)

Spike component of threshold modeled as an exponential decay. Implemented here as exact analytical solution.

Parameters

th0
[float] threshold input to function

b_spike
[float] decay constant of exponential

t
[float or array] time step if used in an Euler setup time if used analytically

allensdk.model.glif.glif_neuron_methods.**spike_component_of_threshold_forward_euler**(*th_t0*, *b_spike*, *dt*)

Spike component of threshold modeled as an exponential decay. Implemented here for forward Euler

Parameters

th_t0
[float] threshold input to function

b_spike
[float] decay constant of exponential

dt
[float] time step

allensdk.model.glif.glif_neuron_methods.**voltage_component_of_threshold_exact**(*th0*, *v0*, *I*, *t*, *a_voltage*, *b_voltage*, *C*, *g*, *El*)

Note this function is the exact formulation; however, dt is used because t0 is the initial time and dt is the time the function is exactly evaluated at. Note: that here, this equation is in reference to th_inf. Therefore th0 is the total

threshold-thr_inf (threshold_inf replaced with 0 in the equation to be verbose). This is done so that th_inf can be optimized without affecting this function.

Parameters

th0
[float] threshold input to function

v0
[float] voltage input to function

I
[float] total current entering neuron (note if there are after spike currents these must be included in this value)

t
[float or array] time step if used in an Euler setup time if used analytically

a_voltage
[float] constant a

b_voltage
[float] constant b

C
[float] capacitance

g
[float] conductance (1/resistance)

El
[float] reversal potential

allensdk.model.glif.glif_neuron_methods.**voltage_component_of_threshold_forward_euler**(*th_t0*, *v_t0*, *dt*, *a_voltage*, *b_voltage*, *El*)

Equation 2.1 of Mihalas and Nieber, 2009 implemented for use in forward Euler. Note here all variables are in reference to threshold infinity. Therefore thr_inf is zero here (replaced threshold_inf with 0 in the equation to be verbose). This is done so that th_inf can be optimized without affecting this function.

Parameters

th_t0
[float] threshold input to function

v_t0
[float] voltage input to function

dt
[float] time step

a_voltage
[float] constant a

b_voltage
[float] constant b

El
[float] reversal potential

allensdk.model.glif.simulate_neuron module

allensdk.model.glif.simulate_neuron.**load_sweep**(*file_name*, *sweep_number*)

Load the stimulus for a sweep from file.

allensdk.model.glif.simulate_neuron.**main**()

allensdk.model.glif.simulate_neuron.**parse_arguments**()

Use argparse to get required arguments from the command line

allensdk.model.glif.simulate_neuron.**simulate_neuron**(*neuron*, *sweep_numbers*, *input_file_name*, *output_file_name*, *spike_cut_value*)

allensdk.model.glif.simulate_neuron.**simulate_sweep**(*neuron*, *stimulus*, *spike_cut_value*)

Simulate a neuron given a stimulus and initial conditions.

allensdk.model.glif.simulate_neuron.**simulate_sweep_from_file**(*neuron*, *sweep_number*, *input_file_name*, *output_file_name*, *spike_cut_value*)

Load a sweep stimulus, simulate the response, and write it out.

allensdk.model.glif.simulate_neuron.**write_sweep_response**(*file_name*, *sweep_number*, *response*, *spike_times*)

Overwrite the response in a file.

Module contents

A Generalized Linear Integrate and Fire (GLIF) neuron modeling package. Use this code to run the GLIF models available in the Allen Cell Types Atlas. See *Generalized LIF Models* for more details.

Module contents

6.1.8 allensdk.morphology package

Submodules

allensdk.morphology.validate_swc module

allensdk.morphology.validate_swc.**main**()

allensdk.morphology.validate_swc.**validate_swc**(*swc_file*)

To be compatible with NEURON, SWC files must have the following properties:

1) a single root node with parent ID '-1'
2) sequentially increasing ID numbers
3) immediate children of the soma cannot branch

Module contents

6.1.9 allensdk.mouse_connectivity package

Subpackages

allensdk.mouse_connectivity.grid package

Subpackages

allensdk.mouse_connectivity.grid.subimage package

Submodules

allensdk.mouse_connectivity.grid.subimage.base_subimage module

class allensdk.mouse_connectivity.grid.subimage.base_subimage.**IntensitySubImage**(*reduce_level*, *in_dims*, *in_spacing*, *coarse_spacing*, *intensity_paths*, **args*, ***kwargs*)

Bases: *SubImage*

get_intensity()

required_intensities = []

setup_images()

class allensdk.mouse_connectivity.grid.subimage.base_subimage.**PolygonSubImage**(*reduce_level*, *in_dims*, *in_spacing*, *coarse_spacing*, *polygon_info*, **args*, ***kwargs*)

Bases: *SubImage*

get_polygons()

optional_polys = []

required_polys = []

setup_images()

class allensdk.mouse_connectivity.grid.subimage.base_subimage.**SegmentationSubImage**(*reduce_level*, *in_dims*, *in_spacing*, *coarse_spacing*, *segmentation_paths*, **args*, ***kwargs*)

Bases: *SubImage*

extract_injection_from_segmentation(*segmentation_name='segmentation'*, *injection_name='injection'*)

Notes

Currently, the segmentation uses a series of codes to map 8-bit values onto meaningful classifications. The code for signal pixels is a 1 in at least one of of the 5 rightmost bits.

extract_signal_from_segmentation(*segmentation_name='segmentation'*, *signal_name='signal'*)

Notes

Currently, the segmentation uses a series of codes to map 8-bit values onto meaningful classifications. The code for signal pixels is a 1 in the leftmost bit.

In some cases, bit 5 indicates that the pixel was not removed in a posfiltering process. Optionally, this postfilter can be applied in gridding.

get_segmentation()

process_segmentation()

read_segmentation_image(*segmentation_name='segmentation'*)

Notes

We downsample in memory rather than using the jp2 pyramid because the segmentation is a label image.

required_segmentations = []

setup_images()

class allensdk.mouse_connectivity.grid.subimage.base_subimage.**SubImage**(*reduce_level*, *in_dims*, *in_spacing*, *coarse_spacing*, **args*, ***kwargs*)

Bases: `object`

apply_mask(*image_name*, *mask_name*, *positive=True*)

apply_pixel_counter(*accumulator_name*, *image*)

binarize(*image_name*)

compute_coarse_planes()

make_pixel_counter()

property pixel_counter

setup_images()

allensdk.mouse_connectivity.grid.subimage.base_subimage.**run_subimage**(*input_data*)

allensdk.mouse_connectivity.grid.subimage.cav_subimage module

class allensdk.mouse_connectivity.grid.subimage.cav_subimage.**CavSubImage**(*reduce_level*, *in_dims*, *in_spacing*, *coarse_spacing*, *polygon_info*, **args*, ***kwargs*)

Bases: *PolygonSubImage*

compute_coarse_planes()

required_polys = ['missing_tile', 'cav_tracer']

allensdk.mouse_connectivity.grid.subimage.classic_subimage module

class allensdk.mouse_connectivity.grid.subimage.classic_subimage.**ClassicSubImage**(*reduce_level*, *in_dims*, *in_spacing*, *coarse_spacing*, *polygon_info*, *segmentation_paths*, *intensity_paths*, *injection_polygon_key='aav_trace*, **args*, ***kwargs*)

Bases: *IntensitySubImage*, *SegmentationSubImage*, *PolygonSubImage*

compute_coarse_planes()

compute_injection()

compute_intensity()

compute_projection()

compute_sum_pixels()

optional_polys = ['aav_tracer']

process_segmentation()

required_intensities = ['green']

required_polys = ['missing_tile', 'no_signal', 'aav_exclusion']

required_segmentations = ['segmentation']

allensdk.mouse_connectivity.grid.subimage.count_subimage module

class allensdk.mouse_connectivity.grid.subimage.count_subimage.**CountSubImage**(*reduce_level*, *in_dims*, *in_spacing*, *coarse_spacing*, *polygon_info*, *segmentation_paths*, *injection_polygon_key='aav_tracer'*, **args*, ***kwargs*)

Bases: *SegmentationSubImage*, *PolygonSubImage*

compute_coarse_planes()

compute_injection()

compute_projection()

compute_sum_pixels()

process_segmentation()

required_polys = ['missing_tile', 'no_signal', 'aav_exclusion']

required_segmentations = ['segmentation']

Module contents

allensdk.mouse_connectivity.grid.subimage.**run_subimage**(*input_data*)

allensdk.mouse_connectivity.grid.utilities package

Submodules

allensdk.mouse_connectivity.grid.utilities.downsampling_utilities module

allensdk.mouse_connectivity.grid.utilities.downsampling_utilities.**apply_divisions**(*image*, *window_size*)

allensdk.mouse_connectivity.grid.utilities.downsampling_utilities.**block_average**(*volume*, *factor*)

allensdk.mouse_connectivity.grid.utilities.downsampling_utilities.**conv**(*image*, *factor*, *window_size*)

allensdk.mouse_connectivity.grid.utilities.downsampling_utilities.**downsample_average**(*volume*, *current_spacing*, *target_spacing*)

allensdk.mouse_connectivity.grid.utilities.downsampling_utilities.**extract**(*image*, *factor*, *window_size*, *window_step*, *output_shape*)

allensdk.mouse_connectivity.grid.utilities.downsampling_utilities.**window_average**(*volume*, *factor*)

allensdk.mouse_connectivity.grid.utilities.image_utilities module

allensdk.mouse_connectivity.grid.utilities.image_utilities.**block_apply**(*in_image*, *out_shape*, *dtype*, *blocks*, *fn*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**build_affine_transform**(*aff_params*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**build_composite_transform**(*dfmfield=None*, *aff_params=None*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**compute_coarse_parameters**(*in_dims*, *in_spacing*, *out_spacing*, *reduce_level*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**grid_image_blocks**(*in_shape*, *in_spacing*, *out_spacing*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**image_from_array**(*array*, *spacing*, *origin=True*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**new_image**(*dims*, *spacing*, *dtype*, *origin=True*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**np_sitk_convert**(*np_type*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**rasterize_polygons**(*shape*, *scale*, *polys*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**read_intensity_image**(*path*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**read_segmentation_image**(*path*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**resample_into_volume**(*image*, *transform*, *z*, *vol*, *dtype=8*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**resample_volume**(*volume*, *dims*, *spacing*, *interpolator=None*, *transform=None*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**set_image_spacing**(*image*, *spacing*, *origin=True*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**sitk_np_convert**(*sitk_type*)

allensdk.mouse_connectivity.grid.utilities.image_utilities.**write_volume**(*volume*, *name*, *prefix=None*, *specify_resolution=None*, *extension='.nrrd'*, *paths=None*)

Module contents

allensdk.mouse_connectivity.grid.writers package

Module contents

allensdk.mouse_connectivity.grid.writers.**cav_writer**(*gridder*, *grid_prefix*, *accumulator_prefix*, ***kwargs*)

allensdk.mouse_connectivity.grid.writers.**classic_writer**(*gridder*, *grid_prefix*, *accumulator_prefix*, *target_spacings*, ***kwargs*)

allensdk.mouse_connectivity.grid.writers.**count_writer**(*gridder*, *grid_prefix*, *accumulator_prefix*, *target_spacings*, ***kwargs*)

allensdk.mouse_connectivity.grid.writers.**handle_pyramid**(*isg*, *key*, *target_spacings*, *prefix*, *paths*)

allensdk.mouse_connectivity.grid.writers.**ratio_and_pyramid**(*isg*, *num*, *den*, *out*, *accumulator_prefix*, *grid_prefix*, *target_spacings*, *paths*)

Submodules

allensdk.mouse_connectivity.grid.image_series_gridder module

class allensdk.mouse_connectivity.grid.image_series_gridder.**ImageSeriesGridder**(*in_dims*, *in_spacing*, *out_dims*, *out_spacing*, *reduce_level*, *subimages*, *subimage_kwargs*, *nprocesses*, *affine_params*, *dfmfld_path*)

Bases: object

accumulator_to_numpy(*key*, *cb*)

build_coarse_grids()

consume_volume(*key*, *cb*)

initialize_coarse_volume(*key*, *dtype*)

make_ratio_volume(*num_key*, *den_key*, *ratio_key*)

assume parents numpified

paste_slice(*key*, *index*, *slice_array*)

paste_subimage(*index*, *output*)

Inserts planar accumulators into coarse grid volumes

resample_volume(*key*)

set_coarse_grid_parameters()

setup_subimages()

property transform

Module contents

Module contents

6.1.10 allensdk.test_utilities package

Submodules

allensdk.test_utilities.custom_comparators module

class allensdk.test_utilities.custom_comparators.**WhitespaceStrippedString**(*string: str*, *whitespace_chars: str = '\\s'*, *ASCII: bool = False*)

Bases: object

Comparator class to compare strings that have been stripped of whitespace. By default removes any unicode whitespace character that matches the regex s, (which includes [tnrfv], and other unicode whitespace characters).

allensdk.test_utilities.custom_comparators.**safe_df_comparison**(*expected: DataFrame*, *obtained: DataFrame*, *expect_identical_column_order: bool = False*)

Compare two dataframes in a way that is agnostic to column order and datatype of NULL values

Parameters

expected: pd.DataFrame
obtained: pd.DataFrame
expect_identical_column_order: bool
If True, raise an error if columns are not in the same order (default=False)

Raises

RuntimeError

If:

- dataframes do not have the same columns
- dataframes do not have identical indexes
- dataframe columns do not have identical contents

When comparing the contents of dataframe columns, the function:

- verifies that NULL values (whether None or NaN) are in the same location
- loops over non-null values, casts arrays into lists, and compares with ==

allensdk.test_utilities.custom_comparators.**stimulus_pickle_equivalence**(*data0: dict*, *data1: dict*) → bool

Compare two sets of data loaded from a stimulus pickle file. Return True if they are identical. Return False otherwise

allensdk.test_utilities.regression_fixture module

allensdk.test_utilities.regression_fixture.**get_list_of_path_dict**()

allensdk.test_utilities.temp_dir module

allensdk.test_utilities.temp_dir.**temp_dir**(*request*)

Module contents

6.2 Submodules

6.2.1 allensdk.deprecated module

allensdk.deprecated.**class_deprecated**(*message=None*)

allensdk.deprecated.**deprecated**(*message=None*)

allensdk.deprecated.**legacy**(*message=None*)

6.3 Module contents

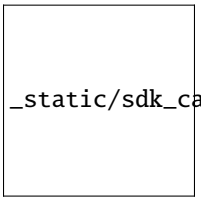
exception allensdk.**OneResultExpectedError**

Bases: RuntimeError

allensdk.**one**(*x*)

The Allen Software Development Kit houses source code for reading and processing Allen Brain Atlas data. The Allen SDK focuses on the Allen Brain Observatory, Cell Types Database, and Mouse Brain Connectivity Atlas.

Attention: As of October 2019, we have dropped Python 2 support and any files with a py2 dependency (for example analysis files) have been updated.



CHAPTER

SEVEN

ALLEN BRAIN OBSERVATORY

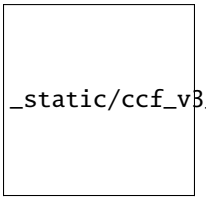
The Allen Brain Observatory is a collection of data resources for understanding sensory processing in the mouse visual cortex. These resources systematically measure visual responses in multiple cortical areas and layers using two-photon calcium imaging or high-density extracellular electrophysiology (Neuropixels) probes. Recordings are performed on mice passively viewing visual stimuli or trained to actively perform an image change detection task.

Behavior	Modality	Resource	Initial Release
Passive	Optical physiology	*Visual Coding - Optical Physiology*	June 2016
Passive	Extracellular electrophysiology	*Visual Coding - Neuropixels*	October 2019
Active	Optical physiology	*Visual Behavior - Optical Physiology*	March 2021
Active	Extracellular electrophysiology	Visual Behavior - Neuropixels	July 2022

Experiment and stimulus data are provided in Neurodata Without Borders (NWB) files. The AllenSDK provides code to:

- download and organize experiment data according to cortical area, imaging depth, and Cre line
- access experiment metadata and data streams
- transform and analyze data

More information about each study is provided in the linked pages. A web-based entry point to the Visual Coding – Optical physiology data is available at http://observatory.brain-map.org/visualcoding .



CHAPTER

EIGHT

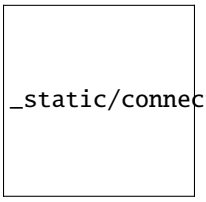
ALLEN CELL TYPES DATABASE

The Allen Cell Types Database contains electrophysiological and morphological characterizations of individual neurons in the mouse primary visual cortex. The Allen SDK provides Python code for accessing electrophysiology measurements (NWB files) for all neurons and morphological reconstructions (SWC files) for a subset of neurons.

The Database also contains two classes of models fit to this data set: biophysical models produced using the NEURON simulator and generalized leaky integrate and fire models (GLIFs) produced using custom Python code provided with this toolkit.

The Allen SDK provides sample code demonstrating how to download neuronal model parameters from the Allen Brain Atlas API and run your own simulations using stimuli from the Allen Cell Types Database or custom current injections:

- *Biophysical Models*
- *Generalized LIF Models*



CHAPTER

NINE

ALLEN MOUSE BRAIN CONNECTIVITY ATLAS

The Allen Mouse Brain Connectivity Atlas is a high-resolution map of neural connections in the mouse brain. Built on an array of transgenic mice genetically engineered to target specific cell types, the Atlas comprises a unique compendium of projections from selected neuronal populations throughout the brain. The primary data of the Atlas consists of high-resolution images of axonal projections targeting different anatomic regions or various cell types using Cre-dependent specimens. Each data set is processed through an informatics data analysis pipeline to obtain spatially mapped quantified projection information.

The Allen SDK provides Python code for accessing experimental metadata along with projection signal volumes registered to a common coordinate framework. This framework has structural annotations, which allows users to compute structure-level signal statistics.

See the mouse connectivity section for more details.

CHAPTER

TEN

WHAT'S NEW - 2.16.1

- See full release notes on Github
- Updated testing and readthedocs configuration
- Fixed bug in loading trials change times.

CHAPTER

ELEVEN

WHAT'S NEW - 2.16.0

- See release notes on Github
- compatibility for python 3.10, 3.11

CHAPTER

TWELVE

WHAT'S NEW - 2.15.0

- See release notes on Github

CHAPTER

THIRTEEN

WHAT'S NEW - 2.14.0

- Support for updated vbn release containing probe, lfp and behavior only data.
- Updates to Ophys data in anticipation of a forthcoming updated data release
- Various notebook updates
- Python 3.9 support
- Various bug fixes and quality of life improvements.

CHAPTER

FOURTEEN

WHAT'S NEW - 2.13.6

- bugfix when accessing stimulus presentations table for vcn data
- updates vbn notebooks

CHAPTER

FIFTEEN

WHAT'S NEW - 2.13.5

- Support for visual behavior neuropixels data

CHAPTER

SIXTEEN

WHAT'S NEW - 2.13.4

- Bug fix in ecephys
- Added support for VBN source density jobs.
- Bug fix for pg8000

CHAPTER

SEVENTEEN

WHAT'S NEW - 2.13.3

- Add ability to extract running speed from mulit-stimulus experiments
- Compatible with pandas 1.4

CHAPTER

EIGHTEEN

WHAT'S NEW - 2.13.2

- Fixes bug that caused file paths on windows machines to be incorrect in Visual behavior user-facing classes
- Updates to support MESO.2
- Loosens/updates required versions for several dependencies
- Updates in order to generate valid NWB files for Neuropixels Visual Coding data collected between 2019 and 2021

CHAPTER

NINETEEN

WHAT'S NEW - 2.13.1

- Fixes bug that was preventing the BehaviorSession from properly instantiating passive sessions.

CHAPTER

TWENTY

WHAT'S NEW - 2.13.0

- Major internal refactor to BehaviorSession, BehaviorOphysExperiment classes. Implements DataObject pattern for fetching and serialization of data.

CHAPTER

TWENTYONE

WHAT'S NEW - 2.12.4

- Documentation changes ahead of SWDB 2021
- Bugfix to CloudCache; it is now possible for multiple users to share a cache.

CHAPTER

TWENTYTWO

WHAT'S NEW - 2.12.3

- Reordered columns in Visual Behavior metadata tables to be more helpful

CHAPTER

TWENTYTHREE

WHAT'S NEW - 2.12.2

- fix to how from_lims API gets OPhys experiment metadata. Preserves relationship between OPhys experiments and failed containers

CHAPTER

TWENTYFOUR

WHAT'S NEW - 2.12.1

- minor fix to cloud cache consistency check

CHAPTER

TWENTYFIVE

WHAT'S NEW - 2.12.0

- Added ability to specify a static cache directory (use_static_cache=True) to instantiate VisualBehaviorOphysProjectCache.from_local_cache()
- Added 'experience_level', 'passive' and 'image_set' columns to ophys_experiments_table
- Added 'ophys_cells_table' metadata table to track the relationship between ophys_experiment_id and cell_specimen_id

CHAPTER

TWENTYSIX

WHAT'S NEW - 2.11.3

- Bugfixes related to NWB creation for BehaviorSessions

CHAPTER

TWENTYSEVEN

WHAT'S NEW - 2.11.2

- Fixed mkdir error for non-existing ecephys upload directory

CHAPTER

TWENTYEIGHT

WHAT'S NEW - 2.11.1

- Refactored the schema for the Ecephys copy utility to avoid raising an error when a previous output file already exists.

CHAPTER

TWENTYNINE

WHAT'S NEW - 2.11.0

- python 3.8 compatibility
- CloudCache (the class supporting cloud-based data releases) is now smart enough to construct symlinks between files that are identical across dataset versions (rather than downloading duplicate copies of files).
- VisualBehavioOphysProjectCache supports user-controlled switching between dataset versions.

CHAPTER

THIRTY

WHAT'S NEW - 2.10.3

- Adds restriction to require hdmf version to be strictly less than 2.5.0 which accidentally introduced a major version breaking change

CHAPTER

THIRTYONE

WHAT'S NEW - 2.10.2

- This version marks the release of Visual Behavior Optical Physiology data! For more details please visit the: Visual Behavior - Optical Physiology Project Page
- Update documentation to support visual behavior data release
- Fixes a bug with the dictionary returned by BehaviorSession get get_performance_metrics() method
- Adds docstrings to the BehaviorSession get_performance_metrics(), get_rolling_performance_df(), and get_reward_rate() methods

CHAPTER

THIRTYTWO

WHAT'S NEW - 2.10.1

- Changes name of BehaviorProjectCache to VisualBehaviorOphysProjectCache
- Changes VisualBehaviorOphysProjectCache method get_session_table() to get_ophys_session_table()
- Changes VisualBehaviorOphysProjectCache method get_experiment_table() to get_ophys_experiment_table()
- VisualBehaviorOphysProjectCache is enabled to instantiate from_s3_cache() and from_local_cache()
- Improvements to BehaviorProjectCache
- Adds project metadata writer

CHAPTER

THIRTYTHREE

WHAT'S NEW - 2.9.0

- Updates to Session metadata; refactors implementation to use class rather than dict internally
- Fixes a bug that was preventing Allen Institute Windows users from accessing gratings images

CHAPTER

THIRTYFOUR

WHAT'S NEW - 2.8.0

- Created lookup table to get monitor_delay for cases where calculation from data fails
- If sync timestamp file has more timestamps than eye tracking moving has frame, trim excess timestamps (up to 15)
- Session API returns both warped and unwarped stimulus images, and both are written to NWB

CHAPTER

THIRTYFIVE

WHAT'S NEW - 2.7.0

- Refactored behavior and ophys session and data APIs to remove a circular inheritance issue
- Fixed segmentation mask and roi_mask misregistration in 'BehaviorOphysSession'
- Replaces BehaviorOphysSession.get_roi_masks() method with roi_masks property
- Fixes bug which prevented the SDK from loading stimuli dataframes for static gratings
- Return event detection data through session API
- Read/write event detection data from/to NWB
- Time stamps for events in trial_log are set to the exact sync timestamp of the corresponding frame.
- For behavior-only sessions, sync-like timestamp of the first frame is set to zero.
- Refactored BehaviorOphysSession to inherit methods and properties from BehaviorSession
- Fixed a test for checking that Behavior and BehaviorOphysSessions contain the same data regardless of which API (LIMS/JSON/NWB) is used. Also fixed resulting failure cases.

CHAPTER

THIRTYSIX

WHAT'S NEW - 2.6.0

- Adds ability to write and read behavior only experiments
- Adds eye tracking ellipse fits and metadata as new NWB data stream
- OPhys Behavior data retrieval methods no longer depend on ROIs being ordered identically in different files.

CHAPTER

THIRTYSEVEN

WHAT'S NEW - 2.5.0 (JANUARY 29, 2021)

- Adds unfiltered running speed as new data stream
- run_demixing gracefully ignores any ROIs that are not in the input trace file

CHAPTER

THIRTYEIGHT

WHAT'S NEW - 2.4.0 (DECEMBER 21, 2020)

As of the 2.4.0 release: - When running raster_plot on a spike_times dataframe, the spike times from each unit are plotted twice. (thank you @dgmurx) - improvements and fixes to behavior ophys NWB files. - improvements and fixes to BehaviorProjectCache tables including new column "donor_id" - implemented a timeout to obtaining an ecephys session. (thank you @wesley-jones) - big overhaul of how Behavior and BehaviorOphys classes are structured for the visual behavior project. See https://github.com/AllenInstitute/AllenSDK/pull/1789

CHAPTER

THIRTYNINE

WHAT'S NEW - 2.3.2 (OCTOBER 19, 2020)

As of the 2.3.2 release:

- (Internal) Fixed a running_processing bug for behavior ophys experiments when the input data would have one more encoder entry than timestamp. The behavior of the code now matches what the warning says.

CHAPTER

FORTY

WHAT'S NEW - 2.3.1 (OCTOBER 13, 2020)

As of the 2.3.1 release:

- (Internal) Fixed a write_nwb bug for behavior ophys experiments involving the BehaviorOphysJsonApi expecting a mesoscope-specific method.

CHAPTER

FORTYONE

WHAT'S NEW - 2.3.0 (OCTOBER 9, 2020)

As of the 2.3.0 release:

- Visual behavior running speed is now low-pass filtered at 10Hz. The raw running speed data is still available. The running speed is corrected for encoder threshold croissing artifacts.
- Support for stimulus gratings for visual behavior.
- Fixed an eye-tracking sync problem.
- Updates to some visual behavior pynwb implementations.
- Adds load sync data for individual plane sets to relate accurate event timings to mesoscope data.
- Adds public API method to access the behavior_session_id from an instance of BehaviorOphysSession.

CHAPTER

FORTYTWO

WHAT'S NEW - 2.2.0 (SEPTEMBER 3, 2020)

As of the 2.2.0 release:

- AllenSDK HTTP engine streaming requests now include a progress bar
- *ImportError: cannot import name 'MultiContainerInterface' from 'hdmf.container'* errors should now be resolved (by removing explicit version bounds on the *hdmf* package).
- The optical physiology 2-photon trace demixer has been modified to be more memory friendly and should no longer result in out of memory errors when trying to demix very large movie stacks.

CHAPTER

FORTYTHREE

WHAT'S NEW - 2.1.0 (JULY 16, 2020)

As of the 2.1.0 release:

- behavior ophys nwb files can now be written using updated pynwb and hdmf
- A warning has been added if you are using AllenSDK with outdated NWB files
- A new documentation file has been added which will contain Visual Behavior specific terms for quick lookup

CHAPTER

FORTYFOUR

WHAT'S NEW - 2.0.0 (JUNE 11, 2020)

As of the 2.0.0 release:

- pynwb and hdmf version requirements have been made less strict
- The organization of data for ecephys neuropixels Neurodata Without Borders (NWB) files has been significantly changed to conform with NWB specifications and best practices
- CCF locations for ecephys neuropixels electrodes are now written to NWB files
- Examples for accessing eye tracking ellipse fit and screen gaze location data have been added to ecephys example notebooks

Important Note: Due to newer versions of pynwb/hdmf having issues reading previously released Visual Coding Neuropixels NWB files and due to the significant reorganization of their NWB file contents, this release contains breaking changes that necessitate a major version revision. NWB files released prior to 6/11/2020 are not guaranteed to work with the 2.0.0 version of AllenSDK. If you cannot or choose not to re-download the updated NWB files, you can continue using a prior version of AllenSDK (< 2.0.0) to access them. However, no further features or bugfixes for AllenSDK (< 2.0.0) are planned. Data released for other projects (Cell Types, Mouse Connectivity, etc.) are *NOT* affected and will *NOT* need to be re-downloaded

CHAPTER

FORTYFIVE

PREVIOUS RELEASE NOTES

BIBLIOGRAPHY

[1] Allen Brain Atlas Data Portal: Downloading a WellKnownFile.

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